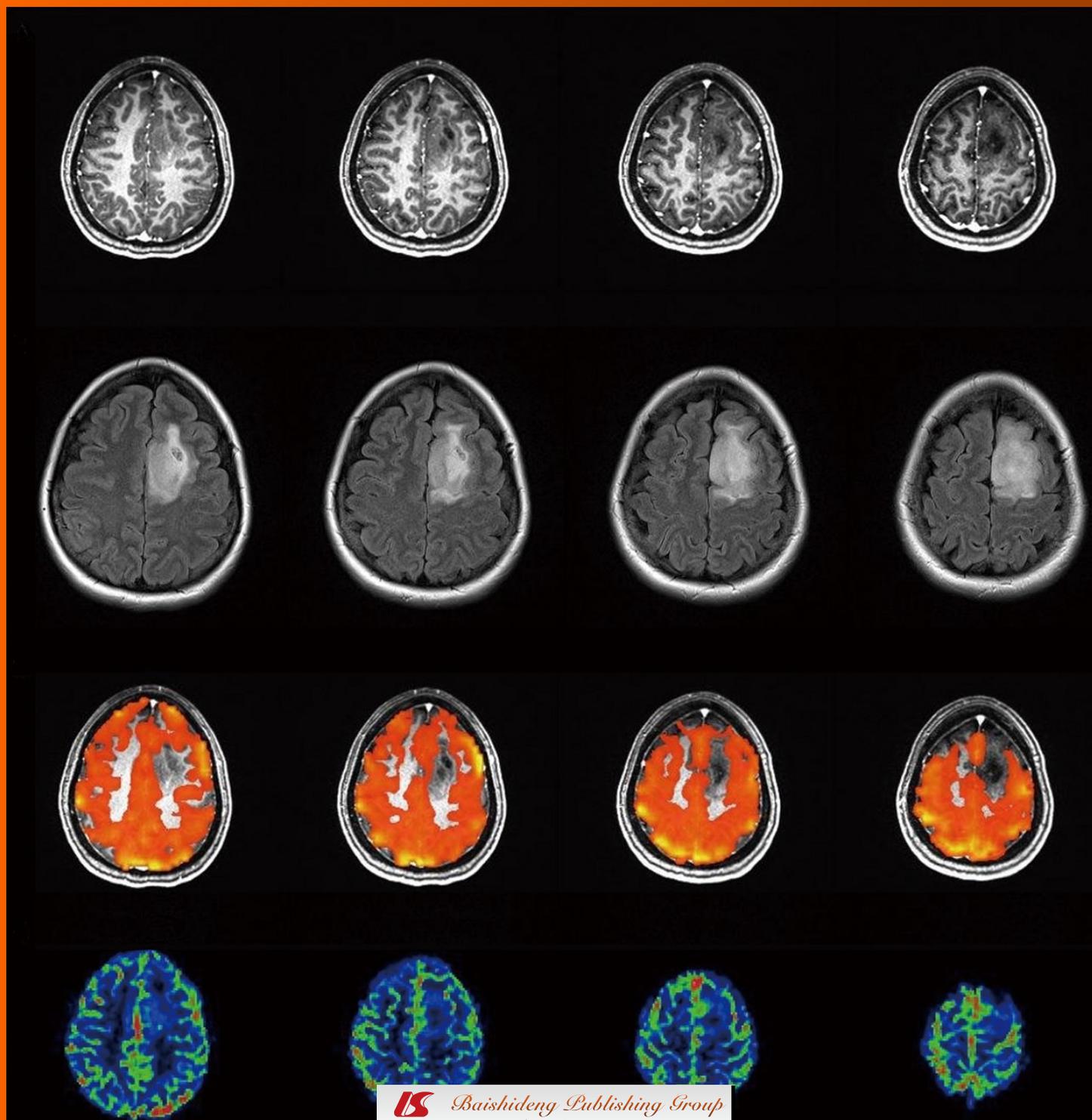


World Journal of *Clinical Oncology*

World J Clin Oncol 2011 December 10; 2(12): 377-403



Editorial Board

2010-2014

The *World Journal of Clinical Oncology* Editorial Board consists of 316 members, representing a team of worldwide experts in oncology. They are from 33 countries, including Australia (6), Belgium (2), Brazil (1), Canada (5), China (34), Egypt (2), Finland (1), France (4), Germany (14), Greece (7), Hungary (1), India (5), Iran (1), Israel (2), Italy (27), Japan (20), Malaysia (1), Mexico (1), Netherlands (6), New Zealand (1), Peru (1), Poland (1), Portugal (4), Saudi Arabia (1), Singapore (9), South Korea (7), Spain (7), Sweden (1), Switzerland (2), Thailand (2), Turkey (6), United Kingdom (11), and United States (123).

PRESIDENT AND EDITOR-IN-CHIEF

Lian-Sheng Ma, *Beijing*

STRATEGY ASSOCIATE EDITORS-IN-CHIEF

Robert J Amato, *Houston*

Kapil Mehta, *Houston*

E YK Ng, *Singapore*

Masahiko Nishiyama, *Saitama*

María Paez de la Cadena, *Vigo*

GJ Peters, *Amsterdam*

Bruno Sangro, *Pamplona*

Wolfgang A Schulz, *Düsseldorf*

Vaclav Vetvicka, *Louisville*

Giuseppe Visani, *Pesaro*

GUEST EDITORIAL BOARD MEMBERS

Shih-Chieh Chang, *Taichung*

How-Ran Guo, *Tainan*

Chao-Cheng Huang, *Kaohsiung*

Chia-Hung Kao, *Taichung*

Shiu-Ru Lin, *Kaohsiung*

Chih-Hsin Tang, *Taichung*

Chih-En Tseng, *Chiayi*

Jaw-Yuan Wang, *Kaohsiung*

Tzu-Chen Yen, *Taoyuan*

Mei-Chin Yin, *Taichung*

Shyng-Shiou F Yuan, *Kaohsiung*

MEMBERS OF THE EDITORIAL BOARD



Australia

Suzanne K Chambers, *Brisbane*

Thomas Grewal, *Sydney*

Peter Hersey, *Newcastle*

Liang Qiao, *Sydney*

Des R Richardson, *Sydney*



Belgium

Tim Van den Wyngaert, *Edegem*

Jan B Vermorken, *Edegem*



Brazil

Gustavo Arruda Viani, *Marilia*



Canada

Dimcho Bachvarov, *Quebec*

Slimane Belbraouet, *Moncton*

Vera Hirsh, *Montreal*

Jennifer Spratlin, *Edmonton*

Seang Lin Tan, *Montreal*



China

Xiao-Tian Chang, *Jinan*

George G Chen, *Hong Kong*

Lei Chen, *Beijing*

Xiao-Ping Chen, *Wuhan*

Yick-Pang Ching, *Hong Kong*

William CS Cho, *Hong Kong*

Yong-Song Guan, *Chengdu*

Lun-Xiu Qin, *Shanghai*

John A Rudd, *Hong Kong*

Jian-Yong Shao, *Guangzhou*

Eric Tse, *Hong Kong*

Gary M Tse, *Hong Kong*

Cheuk Wah, *Hong Kong*

Ming-Rong Wang, *Beijing*

Wei-Hong Wang, *Beijing*

Xun-Di Xu, *Changsha*

Thomas Yau, *Hong Kong*

Qi-Nong Ye, *Beijing*

Anthony PC Yim, *Hong Kong*

Man-Fung Yuen, *Hong Kong*

Ke Zen, *Nanjing*

Xue-Wu Zhang, *Guangzhou*



Egypt

Mohamed Nasser Elsheikh, *Tanta*

Ashraf A Khalil, *Alexandria*



Finland

Veli-Matti Kähäri, *Turku*



France

René Adam, *Villejuif*

Claude Caron de Fromental, *Lyon*

Nathalie Lassau, *Villejuif*

Michel Meignan, *Créteil*



Germany

Thomas Bock, *Berlin*

Christiane Josephine Bruns, *Munich*

Markus W Büchler, *Heidelberg*

André Eckardt, *Hannover*

Felix JF Herth, *Heidelberg*

Georg Kähler, *Mannheim*

Robert Mandic, *Marburg*
Klaus Mross, *Freiburg*
Lars Mueller, *Kiel*
Katharina Pachmann, *Jena*
Matthias Peiper, *Düsseldorf*
Gerd J Ridder, *Freiburg*
Harun M Said, *Wuerzburg*



Greece

Leonidas Duntas, *Athens*
Nicholas Pavlidis, *Ioannina*
Professor A Polyzos, *Athens*
Alexander D Rapidis, *Athens*
Evangelia Razis, *Athens*
Dimitrios Roukos, *Ioannina*
Kostas Syrigos, *Athens*



Hungary

Zsuzsa Schaff, *Budapest*



India

Tanya Das, *Kolkata*
G Arun Maiya, *Manipal*
Ravi Mehrotra, *Allahabad*
Sanjeeb K Sahoo, *Bhubaneswar*
Sarwat Sultana, *New Delhi*



Iran

Ali Kabir, *Tehran*



Israel

Avi Hefetz Khafif, *Tel-Aviv*
Doron Kopelman, *Caesarea*



Italy

Luca Arcaini, *Pavia*
Enrico Benzoni, *Tolmezzo*
Rossana Berardi, *Ancona*
Valentina Bollati, *Milan*
Emilio Bria, *Rome*
Guido Cavaletti, *Monza*
Paolo Chieffi, *Naples*
Marco Ciotti, *Rome*
Giuseppe G Di Lorenzo, *Naples*
Alfio Ferlito, *Udine*
Daris Ferrari, *Abbiategrasso*
Alessandro Franchi, *Florence*
Gennaro Galizia, *Naples*
Roberto Mazzanti, *Firenze*
Michele N Minuto, *Pisa*
Simone Mocellin, *Padova*
Nicola Normanno, *Naples*
Marco G Paggi, *Rome*
Domenico Rubello, *Rovigo*
Antonio Russo, *Palermo*
Daniele Santini, *Rome*
Bruna Scaggiante, *Trieste*

Riccardo Schiavina, *Bologna*
Enzo Spisni, *Bologna*
Bruno Vincenzi, *Rome*
Giovanni Vitale, *Cusano Milanino*



Japan

Hidefumi Aoyama, *Niigata*
Takaaki Arigami, *Kagoshima*
Narikazu Boku, *Shizuoka*
Kazuaki Chikamatsu, *Chuo*
Toru Hiyama, *Higashihiroshima*
Satoru Kakizaki, *Gunma*
Shuichi Kaneko, *Kanazawa*
Koji Kawakami, *Kyoto*
Hiroki Kuniyasu, *Kashihara*
Eiji Miyoshi, *Suita*
Toru Mukohara, *Kobe*
Atsushi Nakajima, *Tokyo*
Takahide Nakazawa, *Sagamihara*
Seishi Ogawa, *Tokyo*
Youngjin Park, *Chiba prefecture*
Naoya Sakamoto, *Tokyo*
Hidekazu Suzuki, *Tokyo*
Michiko Yamagata, *Shimotsuga-gun*
Hiroki Yamaue, *Wakayama*



Malaysia

Min-Tze Liong, *Penang*



Mexico

Rafael Moreno-Sanchez, *Mexico*



Netherlands

Jurgen J Futterer, *Nijmegen*
Bart M Gadella, *Utrecht*
Johannes A Langendijk, *Groningen*
IM Verdonck-de Leeuw, *Amsterdam*
J Voortman, *Amsterdam*



New Zealand

Joanna Skommer, *Auckland*



Peru

Henry L Gomez, *Lima*



Poland

Lukasz Wicherek, *Bydgoszcz*



Portugal

Antonio Araujo, *Porto*
Rui M Medeiros, *Porto*
Paula Ravasco, *Lisbon*
Rui Manuel Reis, *Braga*



Saudi Arabia

Shahab Uddin, *Riyadh*



Singapore

Wei Ning Chen, *Singapore*
John M Luk, *Singapore*
Shu Wang, *Singapore*
Celestial Yap, *Singapore*
Khay-Guan Yeoh, *Singapore*
George W Yip, *Singapore*
Yong Zhang, *Singapore*
Zhan Zhang, *Singapore*



South Korea

Ho-Seong Han, *Seoul*
Young-Seoub Hong, *Busan*
Ja Hyeon Ku, *Seoul*
Geon Kook Lee, *Goyang-si*
Jae Cheol Lee, *Seoul*
Woo Sung Moon, *Jeonju*
Hyun Ok Yang, *Gangeung*



Spain

Maurizio Bendandi, *Pamplona*
Joan Carles, *Barcelona*
Javier Cortés Castán, *Barcelona*
Jose M Cuezva, *Madrid*
Jesús Prieto, *Pamplona*



Sweden

Lalle Hammarstedt, *Stockholm*



Switzerland

A Lugli, *Basel*
Jacqueline Schoumans, *Lausanne*



Thailand

Sueb Wong Chuthapisith, *Bangkok*
Songsak Petmitr, *Bangkok*



Turkey

Nejat Dalay, *Istanbul*
Seher Demirer, *Ankara*
Zafer Özgür Pektaş, *Adana*
Alper Sevinc, *Gaziantep*
Engin Ulukaya, *Gorukle Bursa*
Isik G Yulug, *Ankara*



United Kingdom

Shahriar Behboudi, *London*
Alastair David Burt, *Newcastle*

Barbara Guinn, *Southampton*
Stephen Hiscox, *Cardiff*
Wen G Jiang, *Cardiff*
Youqiang Ke, *Liverpool*
Charles H Lawrie, *Oxford*
T H Marczylo, *Leicester*
Simon N Rogers, *Liverpool*
Abeezar I Sarela, *Leeds*
Alex Tonks, *Cardiff*



United States

Ali Syed Arbab, *Detroit*
Athanassios Argiris, *Pittsburgh*
Raffaele Baffa, *Gaithersburg*
Partha P Banerjee, *Washington*
Scott M Belcher, *Cincinnati*
Heather A Bruns, *Muncie*
Deliang Cao, *Springfield*
William E Carson III, *Columbus*
Disaya Chavalitdhamrong, *Bronx*
Jason Chen, *New York*
Oliver Chen, *Boston*
Jin Q Cheng, *Tampa*
Bruce D Cheson, *Washington*
Mei-Sze Chua, *Stanford*
Muzaffer Cicek, *Rochester*
Ezra EW Cohen, *Chicago*
Hengmi Cui, *Baltimore*
Q Ping Dou, *Detroit*
David W Eisele, *San Francisco*
Wafik S El-Deiry, *Hershey*
Mahmoud El-Tamer, *New York*
Armin Ernst, *Boston*
Zeev Estrov, *Houston*
Marwan Fakih, *Buffalo*
Michelle A Fanale, *Houston*
Xianjun Fang, *Richmond*
Benjamin L Franc, *Sacramento*
Giulia Fulci, *Boston*
David H Garfield, *Denver*
Antonio Giordano, *Philadelphia*
S Murty Goddu, *St. Louis*
Yun Gong, *Houston*
Lei Guo, *Jefferson*
Sanjay Gupta, *Cleveland*
Subrata Haldar, *Cleveland*
Sam M Hanash, *Seattle*
Randall E Harris, *Columbus*
Andrea A Hayes-Jordan, *Houston*
David W Hein, *Louisville*
Paul J Higgins, *Albany*
James R Howe, *Iowa*
Hedvig Hricak, *New York*
Chuanshu Huang, *Tuxedo*
Wendong Huang, *Duarte*
Naijie Jing, *Houston*
Masao Kaneki, *Charlestown*
Hagop Kantarjian, *Houston*
Maria C Katapodi, *Ann Arbor*
Mark R Kelley, *Indianapolis*
Venkateshwar G Keshamouni, *Ann Arbor*
Nikhil Ishwar Khushalani, *Buffalo*
Arianna L Kim, *New York*
K Sean Kimbro, *Atlanta*
Leonidas G Koniaris, *Miami*
Hasan Korkaya, *Ann Arbor*
Sunil Krishnan, *Houston*
Melanie H Kucherlapati, *Boston*
Paul C Kuo, *Maywood*
Andrew C Larson, *Chicago*
Felix Leung, *North Hills*
Ho-Sheng Lin, *Detroit*
Jennifer Lin, *Boston*
Shiaw-Yih Lin, *Houston*
Steven E Lipshultz, *Miami*
Bolin Liu, *Aurora*
Jeri A Logemann, *Evanston*
Bert Lum, *South San Francisco*
Jian-Hua Luo, *Pittsburgh*
Shyamala Maheswaran, *Charlestown*
David L McCormick, *Chicago*
Murielle Mimeault, *Omaha*
Monica Mita, *San Antonio*
Gerard E Mullin, *Baltimore*
Ravi Murthy, *Houston*
Jacques E Nör, *Ann Arbor*
James S Norris, *Charleston*
Scott Okuno, *Rochester*
Timothy Michael Pawlik, *Baltimore*
Joseph A Paydarfar, *Lebanon*
Jay J Pillai, *Baltimore*
Luis F Porrata, *Rochester*
Raj S Pruthi, *Chapel Hill*
Jianyu Rao, *Los Angeles*
Steven A Rosenzweig, *Charleston*
Eric Rowinsky, *Warren*
Jose Russo, *Philadelphia*
Stephen H Safe, *College Station*
Adnan Said, *Madison*
Stewart Sell, *Albany*
Shahrokh F Shariat, *New York*
Jing Shen, *New York*
Dong Moon Shin, *Atlanta*
Haval Shirwan, *Louisville*
Viji Shridhar, *Rochester*
Anurag Singh, *Buffalo*
Lawrence J Solin, *Philadelphia*
David R Spigel, *Nashville*
Brendan Curran Stack, *Little Rock*
Charles F Streckfus, *Houston*
Lu-Zhe Sun, *San Antonio*
Vladimir N Uversky, *Indianapolis*
Jean-Nicolas Vauthey, *Houston*
Hanlin L Wang, *Los Angeles*
Thomas D Wang, *Ann Arbor*
Dennis D Weisenburger, *Omaha*
Robert P Whitehead, *Las Vegas*
Juergen K Willmann, *Stanford*
Jason D Wright, *New York*
Q Jackie Wu, *Durham*
Shenhong Wu, *Stony Brook*
Hang Xiao, *Amherst*
Mingzhao Xing, *Baltimore*
Ronald Xiaorong Xu, *Columbus*
Kaiming Ye, *Fayetteville*
William Andrew Yeudall, *Richmond*
Dihua Yu, *Houston*
Bao-Zhu Yuan, *Morgantown*
Yawei Zhang, *New Haven*
Weixiong Zhong, *Madison*
Shufeng Zhou, *Tampa*
Yue Zou, *Johnson*



EDITORIAL 377 Locally advanced nasopharyngeal carcinoma: Current and emerging treatment strategies
Perri F, Bosso D, Buonerba C, Di Lorenzo G, Della Vittoria Scarpati G

GUIDELINES CLINICAL PRACTICE 384 IL-6/IL-6R as a potential key signaling pathway in prostate cancer development
Azevedo A, Cunha V, Teixeira AL, Medeiros R

BRIEF ARTICLES 397 Clinical utility of cerebrovascular reactivity mapping in patients with low grade gliomas
Pillai JJ, Zacá D

ACKNOWLEDGMENTS I Acknowledgments to reviewers of *World Journal of Clinical Oncology*

APPENDIX I Meetings
 I-V Instructions to authors

ABOUT COVER Jay J Pillai, Domenico Zacá. Clinical utility of cerebrovascular reactivity mapping in patients with low grade gliomas.
World J Clin Oncol 2011; 2(12):397-403
<http://www.wjgnet.com/2218-4333/full/v2/i12/397.htm>

AIM AND SCOPE *World Journal of Clinical Oncology (World J Clin Oncol, WJCO, online ISSN 2218-4333, DOI: 10.5306)* is a monthly peer-reviewed, online, open-access, journal supported by an editorial board consisting of 316 experts in oncology from 33 countries.
 The aim of *WJCO* is to report rapidly new theories, methods and techniques for prevention, diagnosis, treatment, rehabilitation and nursing in the field of oncology. *WJCO* covers etiology, epidemiology, evidence-based medicine, informatics, diagnostic imaging, endoscopy, tumor recurrence and metastasis, tumor stem cells, radiotherapy, chemotherapy, interventional radiology, palliative therapy, clinical chemotherapy, biological therapy, minimally invasive therapy, physiotherapy, psycho-oncology, comprehensive therapy, oncology-related traditional medicine, integrated Chinese and Western medicine, and nursing. *WJCO* covers tumors in various organs/tissues, including the female reproductive system, bone and soft tissue, respiratory system, urinary system, endocrine system, skin, breast, nervous system, head and neck, digestive system, and hematologic and lymphatic system.

FLYLEAF I-III Editorial Board

EDITORS FOR THIS ISSUE Responsible Assistant Editor: *Xiao-Cui Yang*
 Responsible Electronic Editor: *Xiao-Cui Yang*
 Proofing Editor-in-Chief: *Lian-Sheng Ma*
 Responsible Science Editor: *Xing Wu*

NAME OF JOURNAL
World Journal of Clinical Oncology

LAUNCH DATE
 November 10, 2010

SPONSOR
 Beijing Baishideng BioMed Scientific Co., Ltd.,
 Room 903, Building D, Ocean International Center,
 No. 62 Dongsihuan Zhonglu, Chaoyang District,
 Beijing 100025, China
 Telephone: +86-10-8538-1892
 Fax: +86-10-8538-1893
 E-mail: baishideng@wjgnet.com
<http://www.wjgnet.com>

EDITING
 Editorial Board of *World Journal of Clinical Oncology*,
 Room 903, Building D, Ocean International Center,
 No. 62 Dongsihuan Zhonglu, Chaoyang District,
 Beijing 100025, China
 Telephone: +86-10-5908-0036
 Fax: +86-10-8538-1893
 E-mail: wjco@wjgnet.com
<http://www.wjgnet.com>

PUBLISHING
 Baishideng Publishing Group Co., Limited,
 Room 1701, 17/F, Henan Building,
 No.90 Jaffe Road, Wanchai, Hong Kong, China
 Fax: +852-3115-8812
 Telephone: +852-5804-2046

E-mail: baishideng@wjgnet.com
<http://www.wjgnet.com>

SUBSCRIPTION
 Beijing Baishideng BioMed Scientific Co., Ltd.,
 Room 903, Building D, Ocean International Center,
 No. 62 Dongsihuan Zhonglu, Chaoyang District,
 Beijing 100025, China
 Telephone: +86-10-8538-1892
 Fax: +86-10-8538-1893
 E-mail: baishideng@wjgnet.com
<http://www.wjgnet.com>

PUBLICATION DATE
 December 10, 2011

ISSN
 ISSN 2218-4333 (online)

PRESIDENT AND EDITOR-IN-CHIEF
 Lian-Sheng Ma, *Beijing*

STRATEGY ASSOCIATE EDITORS-IN-CHIEF
 Robert J Amato, *Houston*
 María Paez de la Cadena, *Vigo*
 Kapil Mehta, *Houston*
 E YK Ng, *Singapore*
 Masahiko Nishiyama, *Saitama*
 GJ Peters, *Amsterdam*
 Bruno Sangro, *Pamplona*
 Wolfgang A Schulz, *Düsseldorf*
 Vaclav Vetvicka, *Louisville*
 Giuseppe Visani, *Pesaro*

EDITORIAL OFFICE
 Xiao-Cui Yang, Director
World Journal of Clinical Oncology
 Room 903, Building D, Ocean International Center,
 No. 62 Dongsihuan Zhonglu, Chaoyang District,
 Beijing 100025, China
 Telephone: +86-10-8538-1892
 Fax: +86-10-8538-1893
 E-mail: wjco@wjgnet.com
<http://www.wjgnet.com>

COPYRIGHT
 © 2011 Baishideng. Articles published by this Open-Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license.

SPECIAL STATEMENT
 All articles published in this journal represent the viewpoints of the authors except where indicated otherwise.

INSTRUCTIONS TO AUTHORS
 Full instructions are available online at http://www.wjgnet.com/2218-4333/g_info_20100722172206.htm

ONLINE SUBMISSION
<http://www.wjgnet.com/2218-4333office>

Locally advanced nasopharyngeal carcinoma: Current and emerging treatment strategies

Francesco Perri, Davide Bosso, Carlo Buonerba, Giuseppe Di Lorenzo, Giuseppina Della Vittoria Scarpati

Francesco Perri, Oncology Division, Division INT Fondazione "G. Pascale", 80131 Naples, Italy

Davide Bosso, Carlo Buonerba, Giuseppe Di Lorenzo, Giuseppina Della Vittoria Scarpati, Oncology and Rare Cancer Center, Federico II University, 80131 Naples, Italy

Author contributions: Perri F and Della Vittoria Scarpati G collected literature and drafted the first version; Bosso D and Buonerba C critically revised the paper and added additional references; Di Lorenzo G critically revised the paper.

Correspondence to: Giuseppe Di Lorenzo, MD, PhD, Oncology and Rare Cancer Center, Federico II University, 80131

Naples, Italy. giuseppedilorenzoncol@hotmail.com

Telephone: +39-081-7463660 Fax: +39-081-8997370

Received: September 5, 2011 Revised: October 15, 2011

Accepted: October 22, 2011

Published online: December 10, 2011

© 2011 Baishideng. All rights reserved.

Key words: Chemotherapy; Nasopharyngeal carcinoma; Radiotherapy; Treatment

Peer reviewer: Robert Mandic, MD, Associate Professor, Head of Research Laboratory, Department of Otolaryngology, Head and Neck Surgery, University Hospital Giessen and Marburg, Campus Marburg, Deutschhausstrasse 3, D-35037 Marburg, Germany

Perri F, Bosso D, Buonerba C, Di Lorenzo G, Della Vittoria Scarpati G. Locally advanced nasopharyngeal carcinoma: Current and emerging treatment strategies. *World J Clin Oncol* 2011; 2(12): 377-383 Available from: URL: <http://www.wjgnet.com/2218-4333/full/v2/i12/377.htm> DOI: <http://dx.doi.org/10.5306/wjco.v2.i12.377>

Abstract

Although nasopharyngeal carcinoma (NPC) is a widespread malignant tumor, it is particularly frequent in Southeast Asia. Although T1 tumors can be effectively controlled with exclusive radiotherapy, this treatment modality is insufficient for most NPC patients, who present with locally advanced disease at diagnosis. In fact, for stages ranging from T2b N0 to T4 N3, definitive scientific evidence supports the use of concurrent platinum-based chemotherapy with standard external beam radiotherapy. This treatment approach has shown a statistically significant advantage in terms of overall survival, with respect to radiotherapy alone. Several trials have also investigated the use of neoadjuvant and adjuvant chemotherapy in combination with radiotherapy or chemo-radiotherapy. Platinum compounds, anthracyclines and taxanes are among the chemotherapy agents employed. This review focuses on the clinical results obtained in the field of adjuvant/concurrent/neoadjuvant chemotherapy for locally advanced NPC, for which exclusive concurrent chemo-radiotherapy currently represents the standard treatment approach.

INTRODUCTION

Nasopharyngeal carcinoma (NPC), a malignant tumor originating from the epithelium of the nasopharynx, is particularly frequent in Southeast Asia and can be divided into three different histological types, that is, non-keratinizing squamous cell carcinoma, keratinizing squamous cell carcinoma and undifferentiated carcinoma^[1]. At diagnosis, most NPC patients have locally advanced disease, which includes stages ranging from T2b N0 to T4 N3 (Table 1).

Radiotherapy (RT) can control early stage NPC effectively, yielding an excellent 90%-95% 5-year local control rate in clinical trials. However, radiotherapy alone is not the optimal treatment for patients with locally advanced disease, which is the most frequent clinical presentation at diagnosis, since it yields an unsatisfactory 5-year survival rate of about 50%^[2]. For this reason, concurrent platinum-based chemotherapy and radiotherapy has become the standard treatment for locally advanced NPC. While in early stage NPC (T1-2a N0), the addition of chemotherapy to standard radiotherapy has not provided

Table 1 Standard approach to nasopharyngeal carcinoma

Stage	Denomination	Gold standard therapy
T1-2a N0 M0	Early stage	- IMRT alone - Conventional RT alone
From T2b N0 M0 to T4b N3 M0 also every T N2/3 M0	Locally advanced	- Neoadjuvant platinum-based CT followed by IMRT or CCRT (platinum-based) - Concurrent cDDP and RT
Every T every N M1	Metastatic	- Exclusive CT

RT: Radiotherapy; IMRT: Intensity modulated RT; CCRT: Concurrent chemoradiotherapy; CT: Computed tomography.

a survival advantage in clinical trials^[3], a clear superiority has emerged for concurrent chemoradiotherapy when compared to RT alone in patients with locally advanced disease^[4-5].

In locally advanced NPC patients, there are presently few data regarding the use of neoadjuvant/adjuvant chemotherapy, as an alternative to concurrent chemoradiotherapy. The role of neoadjuvant chemotherapy before RT or concurrent chemoradiotherapy is a matter of great interest. In fact, induction chemotherapy is an effective way to control subclinical metastatic foci, especially in patients with lymph node metastasis. Moreover, in some patients with large tumors infiltrating the brain stem, it is often difficult to deliver the total required dose to the clinical target volume (CTV) with preservation of critical tissues. Neoadjuvant chemotherapy is often able to provide objective responses in tumor lesions, which offers the possibility to shrink the CTV and reduce toxicity^[6].

Retrospective studies that used RT alone for NPC indicated that local control was closely linked to the radiation dose delivered to target tissues^[7-8]. Intensity modulated RT (IMRT) is a special type of conformal RT that creates a high dose volume that is precisely shaped around the target volume in order to minimize the radiation dose delivered to surrounding healthy tissues. Investigators compared dosimetric plans of IMRT with conventional RT techniques and concluded that IMRT provided improved tumor coverage and preservation of normal tissues. The proximity of the nasopharynx to critical normal tissues, such as the brainstem and optic structures, makes it challenging for radiotherapists to deliver the optimal radiation dose to the tumor using conventional conformal RT, and underdosing of affected areas is often necessary to preserve healthy tissues. IMRT for locally advanced NPC spares critical portions of the brain stem and of the parotid glands, avoiding neurologic toxicity and permanent xerostomia, respectively. While IMRT has completely replaced conventional conformal RT and has become the standard practice for early stage NPC, its role in the locally advanced setting is not yet well defined^[9].

Intracavitary brachytherapy may be used in patients with residual mass after exclusive upfront radiotherapy, especially in the case of a T2b tumor (parapharyngeal infiltration) at initial diagnosis^[10]. The combination of ex-

ternal beam RT followed by endocavitary brachytherapy may play an important role in patients with T2b disease. In this review, the treatment of patients with locally advanced NPC is reviewed and discussed, with a special focus on novel experimental therapeutic options.

ROLE OF EXCLUSIVE CONCURRENT CHEMORADIO THERAPY

In several phase III trials, radiotherapy with concurrent platinum-based chemotherapy has been compared to standard external beam radiotherapy alone in patients with locally advanced NPC. Concurrent chemo-radiotherapy has shown a statistically significant advantage in terms of survival and response rate when compared with radiotherapy alone, but at the expense of more severe toxicity, mainly mucositis and bone marrow suppression^[4,5,11]. Results of a large meta analysis carried out by Zhang *et al*^[12] which included 1608 patients enrolled in seven studies confirmed the superiority of concurrent chemo-radiotherapy with respect to RT alone. Of note, this meta-analysis was the first to include studies conducted in endemic areas only.

Another meta-analysis included 18 trials enrolling a total of 1993 patients from China. A comparison between concurrent chemo-radiotherapy and RT alone showed that concurrent chemo-radiotherapy was able to obtain a 3-year overall survival rate of 68.5%, compared with 56.4% in the RT alone arm^[13].

More recently, the association of cisplatin and paclitaxel given concurrently with standard radiotherapy was evaluated in a phase II trial. Thirty-one patients with locally advanced NPC received three-weekly 120 mg/m² of paclitaxel and 75 mg/m² of cisplatin concurrently with standard 70 Gy external beam radiotherapy. Three-year overall survival rate was 83.9% and the main grade 3/4 toxicity was neutropenia, reported in 12.9% of patients^[14]. Another way of improving the effectiveness of standard concurrent chemo-radiotherapy may be to modify the radiotherapy scheme. In a phase II trial, Jian *et al*^[15] investigated the activity of hyperfractionated radiotherapy and concomitant platinum-based chemotherapy. As a result, three-year overall survival rate was 72%, with 73% of patients showing grade 3 mucositis, 31% of patients experiencing severe weight loss and 15% requiring a feeding tube. In view of such an unfavorable toxicity profile, further investigation in this direction does not seem justified.

IMRT is widely employed as an alternative to conventional RT in NPC patients with stage I - II disease, but its role in association with chemotherapy is still unknown. Lu *et al*^[16] evaluated the feasibility and efficacy of a weekly cisplatin (40 mg/m²/wk) regimen given concurrently with definitive IMRT in twenty-one locally advanced NPC patients, obtaining a good safety profile and an excellent one-year overall survival of 95.5%^[17]. In another similar trial, the association of three-weekly cisplatin and weekly cetuximab was employed together with standard IMRT

Table 2 Chemo-radiation trials

Trial	Phase	Pts	Study design	Main end- point	Results
Lin JC <i>et al</i> ^[4]	III	284	Exclusive RT alone vs cDDP-5FU + RT	5-year DFS	Experimental arm better ($P < 0.0012$)
Chan AT <i>et al</i> ^[5]	III	350	Exclusive RT alone vs cDDP-5FU + RT	2-year PFS	Experimental arm better ($P < 0.016$)
Zhang L <i>et al</i> ^[12]	III (m)	1608	Exclusive RT alone vs cDDP based CT + RT	5-year OS	Experimental arm better ($P < 0.001$)
Yang AK <i>et al</i> ^[13]	III (m)	1993	Exclusive RT alone vs cDDP based CT + RT	5-year OS	Experimental arm better ($P < 0.05$)
Lu H <i>et al</i> ^[17]	II	22	IMRT + cDDP	1 year OS	96%
Ekenel M <i>et al</i> ^[24]	II	100	IMRT+ cDDP- Cet	ORR	100%

RT: Radiotherapy; IMRT: Intensity modulated RT; CT: Computed tomography; ORR: Overall response rate; DFS: Disease-free survival.

in 100 locally advanced NPC patients. The complete + partial response rate achieved was 100% and the toxicity profile was very low, except for a 64% rate of grade 2 acneiform rash. Clinical studies assessing the efficacy and activity of exclusive concurrent chemo-radiotherapy in locally advanced NPC patients are shown in Table 2.

ROLE OF ADJUVANT CHEMOTHERAPY FOLLOWING CONCURRENT CHEMO-RADIOTHERAPY

The Intergroup-0099 was the first randomized trial to compare concurrent chemo-radiotherapy followed by adjuvant chemotherapy with RT alone^[18]. In this study, concurrent chemo-radiotherapy consisted of cisplatin (100 mg/m² every 21 d) for three cycles, followed by adjuvant cisplatin (80 mg/m² on day 1) and 5-fluorouracil (1000 mg/m² on days 1-4 every 4 wk). A clear and statistically significant advantage in the chemo-radiation arm was seen in terms of overall survival, disease-free-survival, locoregional failure rate and time to distant metastases. However, these encouraging results did not translate into a change in clinical practice for many Asian oncologists, in view of the high rate of well differentiated carcinomas enrolled in both treatment arms (about 25%) which does not reflect common clinical practice. Furthermore, a particularly low compliance was reported, with only 55% undergoing adjuvant treatment and a particularly poor survival observed in the RT-alone arm.

A comparison between concurrent chemo-radiotherapy followed by adjuvant chemotherapy and RT alone was performed in 316 locally advanced NPC patients enrolled in a phase III trial conducted by Chen *et al*^[19]. Patients were assigned to receive concurrent chemo-radiotherapy, consisting of a standard radiation dose of 70 Gy plus cisplatin followed by three adjuvant cycles of cisplatin and 5-fluorouracil, or RT alone. Concurrent chemo-radio-

therapy plus adjuvant chemotherapy yielded better results with respect to RT alone in terms of survival and activity, at the cost of higher toxicity. Similar results were seen in another phase III trial enrolling only non-keratinizing locally advanced NPC patients randomized to concurrent chemo-radiotherapy followed by adjuvant cisplatin and 5-fluorouracil or to RT alone. In this trial, concurrent chemo-radiotherapy was superior in terms of efficacy but also more toxic than RT alone^[20].

Park *et al*^[21] carried out a retrospective analysis in forty-three locally advanced NPC patients treated with concurrent chemo-radiotherapy using cisplatin and 5-fluorouracil followed by adjuvant chemotherapy consisting of three cycles of cisplatin, epirubicin and bleomycin. The overall response rate (ORR) was 95% after concurrent chemo-radiotherapy and 100% after adjuvant therapy. The main toxicities observed were grade 3/4 neutropenia and mucositis occurring during concurrent chemo-radiotherapy.

In a prospective phase II trial conducted by Hu *et al*^[22] fifty-four patients were treated with concomitant weekly paclitaxel and external beam radiation therapy (concurrent chemo-radiotherapy) followed by three cycles of cisplatin (30 mg/m² on days 1-3) and paclitaxel (135 mg/m² on day 1), both given every three weeks. An excellent 100% ORR was obtained after the entire treatment with a complete response (CR) rate of 85%. An acceptable toxicity profile was seen with no grade 3/4 side effects.

In view of the conflicting results reported on the role of adjuvant chemotherapy after concurrent chemo-radiotherapy, it is presently unclear whether the addition of adjuvant therapy may improve the efficacy of concurrent chemo-radiotherapy. Interestingly, a combined analysis of two large studies (NPC-9901 and the NPC-9902) revealed that the dose of cisplatin during the concurrent phase of concurrent chemo-radiotherapy had a significant impact on locoregional control, while additional adjuvant chemotherapy with a fluorouracil-containing combination contributed to improving distant control. Table 3 shows the results of clinical trials assessing the efficacy and/or activity of adjuvant chemotherapy following concurrent chemo-radiotherapy.

ROLE OF NEOADJUVANT CHEMOTHERAPY FOLLOWED BY CONCURRENT CHEMO-RADIOTHERAPY

The role of neoadjuvant chemotherapy followed by concurrent chemo-radiotherapy or RT is a matter of outstanding interest. Several clinical phase III trials from Western countries have proved that induction chemotherapy based on the administration of cisplatin, 5-fluorouracil and taxanes, may significantly improve treatment outcomes in patients with squamous cell carcinoma of the head and neck. An interesting approach may be to employ the same chemotherapy or a similar regimen in locally advanced NPC patients. Amro *et al*^[23] treated 110

Table 3 Adjuvant chemotherapy trials

Trial	Phase	Pts	Study design	Main end-point	Results
Al-Sarraf M <i>et al</i> ^[18]	III	147	Exclusive RT alone vs CCRT followed by cDDP-5FU	3-year PFS	Experimental arm better ($P < 0.01$)
Chen Y <i>et al</i> ^[19]	III	316	Exclusive RT alone vs CCRT followed by cDDP-5FU	2-year OS	Experimental arm better ($P < 0.003$)
Lee AW <i>et al</i> ^[20]	III	348	Exclusive RT alone vs CCRT followed by cDDP-5FU	5-year PFS	Experimental arm better ($P < 0.035$)
Park KH <i>et al</i> ^[21]	II	43	cDDP-5-FU + RT followed by cDDP-Epi-Ble CT	ORR	100%
Hu W <i>et al</i> ^[22]	II	54	w Pac + RT followed by cDDP-Pac CT	ORR	100%
Leung TW <i>et al</i> ^[10]	II	48	HFRT + cDDP based CT followed by cDDP-5FU CT	3-year DFS	71%

RT: Radiotherapy; CT: Computed tomography; ORR: Overall response rate; CCRT: Concurrent chemoradiotherapy; DFS: Disease-free survival.

patients with induction cisplatin and epirubicin followed by a radical course of radiotherapy with three cycles of concurrent cisplatin, and obtained encouraging results in terms of safety and effectiveness. Italian investigators used the same treatment schedule in 40 patients and obtained an overall response rate of 100% and a 5-year disease-free survival of 77%^[24]. In another Italian phase II study, Ferrari *et al*^[25] treated thirty-four patients with three cycles of neoadjuvant cisplatin and 5-fluorouracil followed by concurrent cisplatin and RT. As a result, the overall response rate obtained was a satisfactory 85.3% and the 3-year overall survival rate was 80%.

In the last five years, taxanes have been employed in several phase II and III clinical trials in patients with squamous cell carcinoma of the head and neck, showing a good activity and manageable toxicity profile. Lu *et al*^[26] carried out a trial to compare two different schedules of induction chemotherapy, namely carboplatin-5-fluorouracil (CF) vs docetaxel-carboplatin (TC). Fifty-eight patients with locally advanced NPC were enrolled and randomized to receive CF or TC induction chemotherapy, both followed by concurrent carboplatin and RT. There was no significant difference in terms of response rate and 1-year survival rate. More grade 3/4 neutropenia events were reported in the TC group than in the CF group, whereas less grade 3/4 thrombocytopenia and emesis occurred with the TC regimen than with the CF regimen. An Egyptian study enrolled thirty-six patients who were treated with three cycles of induction paclitaxel (175 mg/m²) and cisplatin (80 mg/m²) given every three weeks, followed by concomitant cisplatin-radiotherapy. The overall response rate after the entire treatment schedule was 89%

and the 3-year overall survival was 68%. The main toxicity encountered was grade 3/4 neutropenia which was observed in 25% of patients^[27]. Hui *et al*^[28] published the results of a randomized phase II trial in which stage III-IVb NPC patients, not previously treated, were randomly assigned to receive either neoadjuvant docetaxel and cisplatin for two cycles followed by concurrent chemo-radiotherapy, or concurrent chemo-radiotherapy alone. A positive impact on survival was observed, since the 3-year overall survival for the neoadjuvant *versus* the control arm was 94.1% *versus* 67.7% ($P = 0.012$). Bossi *et al*^[29] recently presented data of a study on docetaxel, cisplatin and 5-fluorouracil as induction chemotherapy followed by concomitant cisplatin/RT. After completion of treatment, the ORR was 98%, with a complete response rate of 70%. Other authors showed that the same combination had similar results with a 93% response rate and a median time to progression of 39 months^[30]. In a phase II study, induction docetaxel, cisplatin and capecitabine followed by chemo-radiation was tested in 40 patients, and resulted in an ORR of 98% and a complete response rate of 48%^[31]. In a phase II clinical study, Bae *et al*^[32] treated thirty-three patients with induction cisplatin (70 mg/m²), 5-fluorouracil (1000 mg/m² in i.c of 4 d) and docetaxel (75 mg/m²) followed by cisplatin (100 mg/m²) and RT. Twenty-seven patients achieved a partial response and five patients a complete response. An excellent ORR of 98% was achieved and the three-year overall survival rate was 86.1%. Nevertheless, a 72.7% rate of grade 2/3 neutropenia and a 9.1% rate of febrile neutropenia were reported. Xie *et al*^[33] administered induction cisplatin (80 mg/m²) and docetaxel (70 mg/m²) to fifty-seven patients and randomized them to receive either concomitant RT and single agent cisplatin (80 mg/m²) or concomitant cisplatin (80 mg/m²) and docetaxel (60 mg/m²) with RT. After completion of treatment, the complete response rates were very similar in both treatment arms (about 93%), but the occurrence of grade 3/4 neutropenia was significantly higher in the concomitant docetaxel, cisplatin and RT group ($P > 0.05$). In a recent phase II clinical study, fifty-nine locally advanced NPC patients were treated with neoadjuvant cisplatin (75 mg/m²), docetaxel (75 mg/m²) and 5-fluorouracil (500 mg/m² on days 1-5 in i.c) for three cycles, followed by concomitant weekly cisplatin (40 mg/m²) and conventional RT or IMRT. The overall response rate three months after RT was 90.2% and the 1-year overall survival was 100%. The rate of grade 3/4 myelosuppression during induction CT was 55.9% and the corresponding rate during concomitant chemotherapy and RT was 11.9%^[6]. More recently, Ekenel *et al*^[34] published the preliminary results of a phase II trial in which patients with locally advanced NPC received induction cisplatin (75 mg/m²) and docetaxel (75 mg/m²) for three cycles, followed by definitive RT and concomitant cisplatin (100 mg/m²). Fifty-nine patients were evaluable and the ORR obtained after RT was 95%. Three-year overall survival was 93% and the treatment was generally well tolerated with a 10% rate of grade 3/4

Table 4 Neoadjuvant chemotherapy trials

Trial	Phase	Pts	Study design	Main end-point	Results
Al-Amro A <i>et al</i> ^[23]	II	110	Neo cDDP-Epi and followed by cDDP + RT	ORR	100%
Airoldi M <i>et al</i> ^[24]	II	30	Neo cbdca-Pac followed by RT + cbdca-Pac	ORR	87%
Ferrari D <i>et al</i> ^[25]	II	34	Neo cDDP-5FU followed by RT + cDDP	ORR	85.3%
Lu X <i>et al</i> ^[26]	II	58	Neo cbdca-Tax followed by cbdca + RT (arm A) <i>vs</i> neo cbdca-5FU followed by cbdca + RT (armB)	1-year DFS	no difference between arm A and B
Mosatafa E <i>et al</i> ^[27]	II	36	Neo cDDP-Pac followed by cDDP-RT	ORR	89%
Hui EP <i>et al</i> ^[28]	II	65	Neo cDDP-Tax followed by cDDP + RT (arm A) <i>vs</i> cDDP + RT (arm B)	3-year OS	Arm A better than arm B ($P < 0.012$)
Bossi P <i>et al</i> ^[29]	II	45	Neo cDDP-Tax-5FU followed by cDDP + RT	ORR	98%
Cho S <i>et al</i> ^[30]	II	19	Neo cDDP-Tax-5FU followed by cDDP + RT	ORR	93%
Bae WK <i>et al</i> ^[32]	II	33	Neo cDDP-Tax-5FU followed by cDDP + RT	ORR	99%
Kong L <i>et al</i> ^[6]	II	52	Neo cDDP-Tax-5FU followed by cDDP + RT	ORR	90.2%
Ekenel M <i>et al</i> ^[34]	II	59	Neo cDDP-Tax followed by cDDP + RT	ORR	95%
Lin S <i>et al</i> ^[35]	II	370	Neo cDDP based CT followed by IMRT	3-year OS	90%

RT: Radiotherapy; IMRT: Intensity modulated RT; CT: Computed tomography; ORR: Overall response rate; DFS: Disease-free survival.

hematologic toxicity.

In the last three years, a significant effort has been made to incorporate IMRT in treatment protocols for locally advanced NPC. In a phase II study, Lin *et al*^[35] treated a total of 370 patients with locally advanced NPC, with stages ranging from II b to IV, with induction cisplatin-based chemotherapy followed by exclusive IMRT or the association of IMRT and concomitant cisplatin. Drugs more frequently used in combination with cisplatin were paclitaxel and 5-fluorouracil. With a median follow-up of 31 mo, the three-year disease-free-survival and overall survival were 81% and 89%, respectively. A subgroup analysis revealed that concurrent chemotherapy provided no significant benefit to IMRT but was responsible for higher rates of grade 3/4 toxicities^[35]. A recent Italian phase II trial carried out by Palazzi *et al*^[36] enrolled 87 patients with locally advanced NPC and treated them with either conventional RT or with IMRT. Of these

patients, 26% received only concurrent cisplatin and the other 74% received both induction and concurrent CT. Three-year disease-free survival (DFS) and overall survival were 82% and 90%, respectively. Interestingly, a multivariate analysis revealed that histology, N-stage, RT-technique and total dose of RT had the strongest independent impact on DFS.

Further clinical trials assessing activity, efficacy and toxicity of the combination of induction taxane-based chemotherapy followed by exclusive IMRT in locally advanced NPC are warranted. At the present time, this treatment strategy is recommended only in experienced centers. Table 4 shows clinical trials assessing the activity and efficacy of neoadjuvant chemotherapy followed by radiation or chemo-radiation.

CONCLUSION

This review reported in detail the available clinical data regarding the use of chemotherapy in combination with radiotherapy for locally advanced NPC. Although several cytotoxic agents have been used both in the neoadjuvant and adjuvant setting with promising results, exclusive concurrent chemo-radiotherapy remains the recommended approach at the present time, as additional evidence is required to support the use of chemotherapy in the adjuvant/neoadjuvant setting.

REFERENCES

- 1 **Caponigro F**, Longo F, Ionna F, Perri F. Treatment approaches to nasopharyngeal carcinoma: a review. *Anticancer Drugs* 2010; **21**: 471-477
- 2 **Wang CC**. Radiation therapy for head and neck neoplasms. 3rd edition. New York: Wiley-Liss, 1987: 274
- 3 **Song CH**, Wu HG, Heo DS, Kim KH, Sung MW, Park CI. Treatment outcomes for radiotherapy alone are comparable with neoadjuvant chemotherapy followed by radiotherapy in early-stage nasopharyngeal carcinoma. *Laryngoscope* 2008; **118**: 663-670
- 4 **Lin JC**, Jan JS, Hsu CY, Liang WM, Jiang RS, Wang WY. Phase III study of concurrent chemoradiotherapy versus radiotherapy alone for advanced nasopharyngeal carcinoma: positive effect on overall and progression-free survival. *J Clin Oncol* 2003; **21**: 631-637
- 5 **Chan AT**, Teo PM, Ngan RK, Leung TW, Lau WH, Zee B, Leung SF, Cheung FY, Yeo W, Yiu HH, Yu KH, Chiu KW, Chan DT, Mok T, Yuen KT, Mo F, Lai M, Kwan WH, Choi P, Johnson PJ. Concurrent chemotherapy-radiotherapy compared with radiotherapy alone in locoregionally advanced nasopharyngeal carcinoma: progression-free survival analysis of a phase III randomized trial. *J Clin Oncol* 2002; **20**: 2038-2044
- 6 **Kong L**, Zhang YW, Hu CS, Guo Y. Neoadjuvant chemotherapy followed by concurrent chemoradiation for locally advanced nasopharyngeal carcinoma. *Chin J Cancer* 2010; **29**: 551-555
- 7 **Vikram B**, Mishra UB, Strong EW, Manolatos S. Patterns of failure in carcinoma of the nasopharynx: I. Failure at the primary site. *Int J Radiat Oncol Biol Phys*. 1985; **11**: 1455-1459
- 8 **Lee AW**, Tung SY, Chan AT, Chappell R, Fu YT, Lu TX, Tan T, Chua DT, O'sullivan B, Xu SL, Pang ES, Sze WM, Leung TW, Kwan WH, Chan PT, Liu XF, Tan EH, Sham JS, Siu L, Lau WH. Preliminary results of a randomized

- study (NPC-9902 Trial) on therapeutic gain by concurrent chemotherapy and/or accelerated fractionation for locally advanced nasopharyngeal carcinoma. *Int J Radiat Oncol Biol Phys* 2006; **66**: 142-151
- 9 **Lu H**, Yao M. The current status of intensity-modulated radiation therapy in the treatment of nasopharyngeal carcinoma. *Cancer Treat Rev* 2008; **34**: 27-36
 - 10 **Leung TW**, Tung SY, Wong VY, Sze WK, Lui CM, Wong FC, Lee AS, O SK. Nasopharyngeal intracavitary brachytherapy: the controversy of T2b disease. *Cancer* 2005; **104**: 1648-1655
 - 11 **Chua DT**, Sham JS, Au GK, Choy D. Concomitant chemoradiation for stage III-IV nasopharyngeal carcinoma in Chinese patients: results of a matched cohort analysis. *Int J Radiat Oncol Biol Phys* 2002; **53**: 334-343
 - 12 **Zhang L**, Zhao C, Ghimire B, Hong MH, Liu Q, Zhang Y, Guo Y, Huang YJ, Guan ZZ. The role of concurrent chemoradiotherapy in the treatment of locoregionally advanced nasopharyngeal carcinoma among endemic population: a meta-analysis of the phase III randomized trials. *BMC Cancer* 2010; **10**: 558
 - 13 **Yang AK**, Liu TR, Guo X, Qi GL, Chen FJ, Guo ZM, Zhang Q, Zeng ZY, Chen WC, Li QL. [Concurrent chemoradiotherapy versus radiotherapy alone for locoregionally advanced nasopharyngeal carcinoma: a meta-analysis]. *Zhonghua Er Bi Yan Hou Tou Jing Wai Ke Za Zhi* 2008; **43**: 218-223
 - 14 **He XY**, Hu CS, Ying HM, Wu YR, Zhu GP, Liu TF. Paclitaxel with cisplatin in concurrent chemoradiotherapy for locally advanced nasopharyngeal carcinoma. *Eur Arch Otorhinolaryngol* 2010; **267**: 773-778
 - 15 **Jian JJ**, Cheng SH, Tsai SY, Yen KC, Chu NM, Chan KY, Tan TD, Cheng JC, Lin YC, Leu SY, Hsieh CI, Tsou MH, Lin CY, Huang AT. Improvement of local control of T3 and T4 nasopharyngeal carcinoma by hyperfractionated radiotherapy and concomitant chemotherapy. *Int J Radiat Oncol Biol Phys*. 2002; **53**: 344-52
 - 16 **Lu T**, Zhao C, Gao L, Lang J, Pan J, Hu C. Open, multicenter clinical study on cetuximab combined with intensity modulated radiotherapy (IMRT) plus concurrent chemotherapy in nasopharyngeal carcinoma (NPC); preliminary report. *J Clin Oncol* 28: 15s, 2010 (suppl abstr 5577)
 - 17 **Lu H**, Chen J, Huang B, Cheng J, Peng L, Hao Y, Liao C, Mo Y, Wu D, Qin J. Feasibility and efficacy study of weekly cisplatin with concurrent intensity-modulated radiation therapy for nasopharyngeal carcinoma: preliminary results. *Oral Oncol* 2010; **46**: 743-747
 - 18 **Al-Sarraf M**, LeBlanc M, Giri PG, Fu KK, Cooper J, Vuong T, Forastiere AA, Adams G, Sakr WA, Schuller DE, Ensley JF. Chemoradiotherapy versus radiotherapy in patients with advanced nasopharyngeal cancer: phase III randomized intergroup study 0099. *J Clin Oncol* 1998; **16**: 1310-1317
 - 19 **Chen Y**, Liu MZ, Liang SB, Zong JF, Mao YP, Tang LL, Guo Y, Lin AH, Zeng XF, Ma J. Preliminary results of a prospective randomized trial comparing concurrent chemoradiotherapy plus adjuvant chemotherapy with radiotherapy alone in patients with locoregionally advanced nasopharyngeal carcinoma in endemic regions of china. *Int J Radiat Oncol Biol Phys* 2008; **71**: 1356-1364
 - 20 **Lee AW**, Tung SY, Chua DT, Ngan RK, Chappell R, Tung R, Siu L, Ng WT, Sze WK, Au GK, Law SC, O'Sullivan B, Yau TK, Leung TW, Au JS, Sze WM, Choi CW, Fung KK, Lau JT, Lau WH. Randomized trial of radiotherapy plus concurrent-adjuvant chemotherapy vs radiotherapy alone for regionally advanced nasopharyngeal carcinoma. *J Natl Cancer Inst* 2010; **102**: 1188-1198
 - 21 **Park KH**, Kim JS, Park Y, Seo HY, Park YJ, Choi IK, Oh SC, Seo JH, Kim CY, Jung KY, Shin SW, Kim YH, Kim JS, Lee NJ. Concurrent chemoradiation followed by adjuvant chemotherapy in patients with locoregionally advanced nasopharyngeal carcinoma in Korea. *Cancer Chemother Pharmacol* 2010; **66**: 643-651
 - 22 **Hu W**, Ding W, Yang H, Shao M, Wang B, Wang J, Wu S, Wu S, Jin L, Ma CC. Weekly paclitaxel with concurrent radiotherapy followed by adjuvant chemotherapy in locally advanced nasopharyngeal carcinoma. *Radiother Oncol* 2009; **93**: 488-491
 - 23 **Al-Amro A**, Al-Rajhi N, Khafaga Y, Memon M, Al-Hebshi A, El-Enbabi A, El-Husseiny G, Radawi A, Belal A, Allam A, El-Sebaie M. Neoadjuvant chemotherapy followed by concurrent chemo-radiation therapy in locally advanced nasopharyngeal carcinoma. *Int J Radiat Oncol Biol Phys* 2005; **62**: 508-513
 - 24 **Airoldi M**, Gabriele P, Gabriele AM, Garzaro M, Raimondo L, Pedani F, Beatrice F, Pecorari G, Giordano C. Induction chemotherapy with carboplatin and taxol followed by radiotherapy and concurrent weekly carboplatin taxol in locally advanced nasopharyngeal carcinoma. *Cancer Chemother Pharmacol* 2010 Jul 20. [Epub ahead of print]
 - 25 **Ferrari D**, Chiesa F, Codecà C, Calabrese L, Jereczek-Fossa BA, Alterio D, Fiore J, Luciani A, Floriani I, Orecchia R, Foa P. Locoregionally advanced nasopharyngeal carcinoma: induction chemotherapy with cisplatin and 5-fluorouracil followed by radiotherapy and concurrent cisplatin: a phase II study. *Oncology* 2008; **74**: 158-166
 - 26 **Lu X**, Guo X, Hong MH, Chen QY, Zeng Q, Xiang YQ. Comparison of the short-term efficacy of two inductive chemotherapy regimens for locally advanced nasopharyngeal carcinoma: docetaxel plus carboplatin versus 5-fluorouracil plus carboplatin. *Chin J Cancer* 2010; **29**: 140-144
 - 27 **Mostafa E**, Nasar MN, Rabie NA, Ibrahim SA, Barakat HM, Rabie AN. Induction chemotherapy with paclitaxel and cisplatin, followed by concomitant cisplatin and radiotherapy for the treatment of locally advanced nasopharyngeal carcinoma. *J Egypt Natl Canc Inst* 2006; **18**: 348-356
 - 28 **Hui EP**, Ma BB, Leung SF, King AD, Mo F, Kam MK, Yu BK, Chiu SK, Kwan WH, Ho R, Chan I, Ahuja AT, Zee BC, Chan AT. Randomized phase II trial of concurrent cisplatin-radiotherapy with or without neoadjuvant docetaxel and cisplatin in advanced nasopharyngeal carcinoma. *J Clin Oncol* 2009; **27**: 242-249
 - 29 **Bossi P**, Parolini D, Bergamini C, Locati LD, Orlandini E, Franceschini M. TPF induction chemotherapy (CI) followed by concomitant cisplatin/radiotherapy (cTRT) in locally advanced nasopharyngeal cancer (LANPC). abstract 6046. *J Clin Oncol* 2009; **27**
 - 30 **Cho S**, Bae W, Hwang J, Shim H, Lee J, Lim S. Phase II study of docetaxel, cisplatin and 5-FU induction chemotherapy followed by concurrent radiotherapy for locally advanced nasopharyngeal cancer. Abstract 17010. *J Clin Oncol* 2008; **26**
 - 31 **Beldjiladji Y**, Beldjiladji KA, Boukerche A, Kellafi H, Abdelaoui A, Betkaoui Fabstreet. First results of induction chemotherapy with cisplatin, docetaxel and capecitabine for the treatment of nasopharyngeal carcinoma. Abstract 6045. *J Clin Oncol* 2009; **27**.
 - 32 **Bae WK**, Hwang JE, Shim HJ, Cho SH, Lee JK, Lim SC, Chung WK, Chung IJ. Phase II study of docetaxel, cisplatin, and 5-FU induction chemotherapy followed by chemoradiotherapy in locoregionally advanced nasopharyngeal cancer. *Cancer Chemother Pharmacol* 2010; **65**: 589-595
 - 33 **Xie FY**, Qi SN, Hu WH, Zou GR, Peng M, Li JS. [Comparison of efficacy of docetaxel combined cisplatin (TP regimen) and cisplatin combined 5-fluorouracil (PF regimen) on locally advanced nasopharyngeal carcinoma]. *Ai Zheng* 2007; **26**: 880-884
 - 34 **Ekenel M**, Keskin S, Basaran M, Bavbek E, Ozdemir C, Meral R. Clinical outcomes in patients with locally advanced nasopharyngeal cancer treated with neoadjuvant docetaxel and cisplatin followed by radiation treatment and concomitant cisplatin. *J Clin Oncol* abstract 16017. 2010; **28**
 - 35 **Lin S**, Lu JJ, Han L, Chen Q, Pan J. Sequential chemotherapy and intensity-modulated radiation therapy in the manage-

ment of locoregionally advanced nasopharyngeal carcinoma: experience of 370 consecutive cases. *BMC Cancer* 2010; **10**: 39

36 **Palazzi M**, Orlandi E, Bossi P, Pignoli E, Potepan P, Guzzo M, Franceschini M, Scaramellini G, Cantù G, Licitra L, Olmi

P, Tomatis S. Further improvement in outcomes of nasopharyngeal carcinoma with optimized radiotherapy and induction plus concomitant chemotherapy: an update of the Milan experience. *Int J Radiat Oncol Biol Phys* 2009; **74**: 774-780

S- Editor Yang XC **L- Editor** Webster JR **E- Editor** Yang XC

IL-6/IL-6R as a potential key signaling pathway in prostate cancer development

Andreia Azevedo, Virginia Cunha, Ana Luisa Teixeira, Rui Medeiros

Andreia Azevedo, Virginia Cunha, Ana Luisa Teixeira, Rui Medeiros, Molecular Oncology and Virology, Portuguese Institute of Oncology, 4200-072 Porto, Portugal

Andreia Azevedo, Rui Medeiros, Research Department of Portuguese League against Cancer (NRNorte), 4200-072 Porto, Portugal

Rui Medeiros, CEBIMED, Faculty of Health Sciences of Fernando Pessoa University, 4200-150 Porto, Portugal

Ana Luisa Teixeira, Rui Medeiros, ICBAS, Abel Salazar Institute for the Biomedical Sciences, University of Porto, 4200-072 Porto, Portugal

Author contributions: Azevedo A and Cunha V designed the structure of the review; Azevedo A wrote the initial draft and the final version of the manuscript; Cunha V, Teixeira AL and Medeiros R critically revised the manuscript for important intellectual content; Medeiros R supervised the study and approved the version to be published.

Supported by Calouste Gulbenkian Foundation (Oncology/2008/Project n 96736) and Science and Technology Foundation (FCT/PTDC/SAU-FCF/71552/2006)

Correspondence to: Rui Medeiros, Professor, Molecular Oncology and Virology, Portuguese Institute of Oncology, Porto, R. Dr. António Bernardino de Almeida, 4200-072 Porto, Portugal. ruimedei@ipoporto.min-saude.pt

Telephone: +351-22-5084000 Fax: +351-22-5084001

Received: September 9, 2011 Revised: November 8, 2011

Accepted: November 15, 2011

Published online: December 10, 2011

PC patients. In PC treatment, patients diagnosed with advanced stages are frequently submitted to hormonal castration, although most patients will eventually fail this therapy and die from recurrent castration-resistant prostate cancer (CRPC). Therefore, it is important to understand the mechanisms involved in CRPC. Several pathways have been proposed to be involved in CRPC development, and their understanding will improve the way to more effective therapies. In fact, the prostate is known to be dependent, not exclusively, on androgens, but also on growth factors and cytokines. The signaling pathway mediated by IL-6 may be an alternative pathway in the CRPC phenotype acquisition and cancer progression, under androgen deprivation conditions. The principal goal of this review is to evaluate the role of IL-6 pathway signaling in human PC development and progression and discuss the interaction of this pathway with the androgen receptor pathway. Furthermore, we intend to evaluate the inclusion of IL-6 and its receptor levels as a putative new class of tumor biomarkers. The IL-6/IL-6R signaling pathway may be included as a putative molecular marker for aggressiveness in PC and it may be able to maintain tumor growth through the AR pathway under androgen-deprivation conditions. The importance of the IL-6/IL-6R pathway in regulation of PC cells makes it a good candidate for targeted therapy.

© 2011 Baishideng. All rights reserved.

Abstract

Interleukin-6 (IL-6) is a pleiotropic cytokine involved in prostate regulation and in prostate cancer (PC) development/progression. IL-6 acts as a paracrine and autocrine growth stimulator in benign and tumor prostate cells. The levels of IL-6 and respective receptors are increased during prostate carcinogenesis and tumor progression. Several studies reported that increased serum and plasma IL-6 and soluble interleukin-6 receptor levels are associated with aggressiveness of the disease and are associated with a poor prognosis in

Key words: Androgen receptor; Castration-resistant prostate cancer; Hormonal castration; Interleukin-6; Interleukin-6 receptor; Prostate cancer; Tumor biomarker

Peer reviewers: Joan Carles, MD, PhD, Director, GU, CNS and Sarcoma Program, Department of Medical Oncology, Passeig Vall d'Hebron, 119-129, 08035 Barcelona, Spain; Shahrokh F Shariat, MD, PhD, Associate Professor, Department of Urology, New York Presbyterian Hospital, Weill Medical College of Cornell University, 525 East 68th Street, Starr 900,

New York, NY 10021, United States; Ke Zen, PhD, Chang-Jiang Professor, Vice Dean, School of Life Sciences, Nanjing University, Hankou Road 22, Nanjing 210093, Jiangsu Province, China

Azevedo A, Cunha V, Teixeira AL, Medeiros R. IL-6/IL-6R as a potential key signaling pathway in prostate cancer development. *World J Clin Oncol* 2011; 2(12): 384-396 Available from: URL: <http://www.wjgnet.com/2218-4333/full/v2/i12/384.htm> DOI: <http://dx.doi.org/10.5306/wjco.v2.i12.384>

INTRODUCTION

Prostate cancer (PC) is the most common cancer among men in Western populations^[1]. Ethnicity, advanced age and family history are well known risk factors for this disease^[1]. Furthermore, circulating androgen levels, chronic prostate inflammation and obesity are also risk factors frequently described in the literature^[2,3]. Currently, prostate-specific antigen (PSA) is the putative biomarker for PC screening. Two consecutive rises in PSA value over 0.5 ng/mL or one single value ≥ 4 ng/mL are indications for biopsy^[4]. However, although PSA testing has high sensibility, its specificity is rather low, causing clinicians to have doubts with regard to biopsying, since increased false-positive rates, overdiagnosis and overtreatment have been reported to be associated with PSA testing^[5-8]. Therefore, novel biomarkers are needed to improve identification of men at risk of having PC and to predict the natural behavior of the prostate tumor. The use of more sensitive and specific biomarkers will be an appropriate strategy for disease diagnosis, disease staging, disease prognosis, predicting and monitoring clinical response to therapy. In consequence of the high heterogeneity of PC, it is relevant to study molecular and cellular pathways involved in its development and progression to identify key genes and molecules implicated in different stages of the disease^[9].

In several diseases, a deregulation of cytokine levels can be observed. Numerous pro-inflammatory cytokines play an important role in the pathogenesis/carcinogenesis of many cancers. One of the most important cytokine associated with inflammation, interleukin-6 (IL-6), will be discussed in this manuscript.

Recent evidence suggests that the presence of inflammatory factors and cytokines at the tumor site results in tumor cell survival, proliferation, invasion and metastasis^[10]. The expression and function of pro-inflammatory cytokines in PC have been extensively investigated because of their role in the regulation of proliferation, apoptosis, migration, invasion, and angiogenesis^[11]. Recent studies have focused on the role of cytokines, including IL-6, in the etiology and progression of PC^[12-16]. Elevated levels of IL-6 in men with local PC and advanced disease made IL-6 a candidate biomarker for PC development and progression^[17].

This review is a summary of several studies outlining the potential role of the IL-6 signaling pathway in human

PC development and progression, an interesting area of scientific and clinical research.

IL-6 AND RECEPTORS IN CANCER

IL-6 is involved in the regulation of various cellular functions, among them proliferation, apoptosis, angiogenesis, differentiation and regulation of immune response^[12]. This protein is a pleiotropic cytokine synthesized by different cell types, such as B and T-cells, macrophages, monocytes, fibroblasts, endothelial and mesothelial cells, keratinocytes, mast cells, stromal cells, some nerve cells and certain tumor cells^[18]. Adipose tissue is another main source of IL-6^[19].

The biological activity of IL-6 is initiated when the cytokine binds to a receptor complex: an 80-kDa component receptor, non-signaling α -receptor subunit (IL-6R or 80gp) and two signal-transduction components of 130 kDa (gp130) (Figure 1)^[20,21]. IL-6R is expressed only by hepatocytes, neutrophils, monocytes/macrophages and some lymphocytes, while gp130 is expressed by all body cells^[22]. Dimerization of IL-6/IL-6R/gp130 lead to the initiation of intracellular signaling, through Janus Kinase-Signal Transducer and Activator of Transcription (JAK-STAT), Mitogen-activated protein kinase (MAPK) and phosphatidylinositol 3-kinase/Akt kinase (PI3-K/AKT) pathways^[23] and consequently activate the expression of different genes with crucial roles in inflammation and cancer development. This mechanism is the classical signaling pathway^[24]. When IL-6 binds to the receptor, STAT3 is activated in a JAK-dependent manner that leads to increased receptor activator of nuclear factor κ B ligand (RANKL) expression. IL-6 may also activate AKT *via* increased JAK-dependent PI3K activity and results in cell survival and anti-apoptosis signaling. Concomitantly, increased MAPK activity downstream of JAK activation can lead to up-regulated cell growth, proliferation, and mitosis (Figure 1)^[25].

Some cells express lower levels of IL-6 transmembrane receptor, in this case IL-6 can bind to a soluble form of the IL-6R (sIL-6R). Then, through a process denominated trans-signaling, the IL-6/sIL-6R complex binds to gp130 and subsequently signal transduction pathways are activated (Figure 1)^[24,26]. Due to the fact that the sIL-6R lacks a membrane signaling domain, there appears to be significant differences in the intracellular signaling pathways. While IL-6 trans-signaling also leads to phosphorylation and activation of STAT3, increased cell survival, proliferation, and mitosis occurs in an AKT- and MAPK-independent manner. The exact mechanisms for IL-6 trans-signaling leading to increased cell survival, proliferation, and mitosis are not yet known (Figure 1)^[25].

In humans, sIL-6R can be generated by proteolytic cleavage (90%) of the transmembrane receptor mediated by metalloproteinases such as a disintegrin and metalloproteinase 10 and 17^[27,28] or by an alternative mRNA splicing (10%)^[29].

A soluble form of gp130 (sgp130) is present at con-

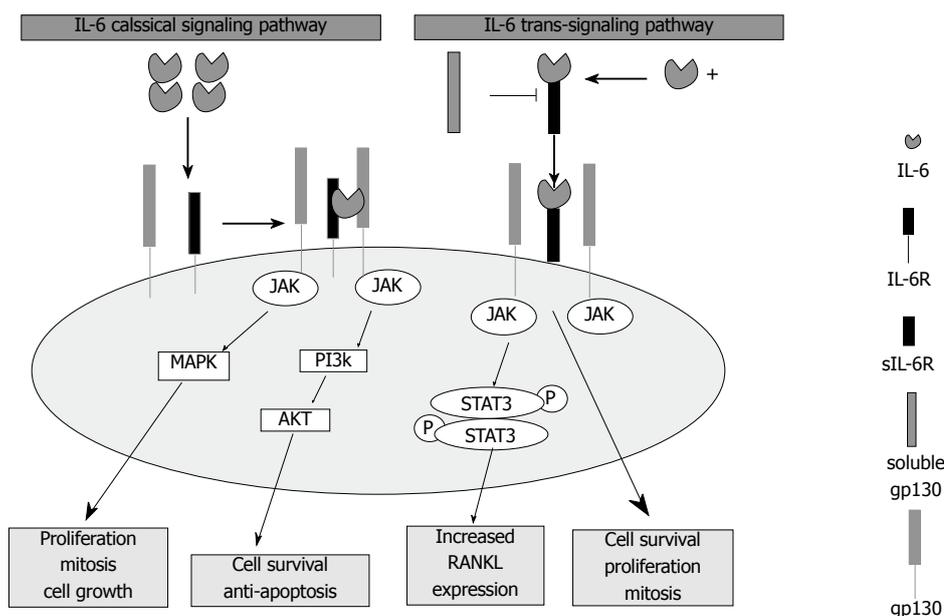


Figure 1 Schematic of IL-6 classical and trans-signaling pathway. IL-6: Interleukin 6; JAK: Janus kinase; MAPK: Mitogen-activated protein kinase; PI3k: Phosphatidylinositol 3-kinase; RANKL: Receptor activator of nuclear factor κ B ligand; sIL-6R: Soluble IL-6 receptor; STAT3: Signal transducer and activator of transcription 3.

centrations between 100-400 ng/mL (1-4 nmol/L) in healthy human serum^[30]. Jostock *et al*^[31] reported that IL-6 alone does not interact with sgp130. Thus, signaling *via* the membrane-bound IL-6R is not inhibited by sgp130^[31]. Based on their results, it was suggested that endogenous sgp130 may be a natural antagonist of the IL-6/sIL-6R complex *in vivo*, probably to prevent systemic IL-6 trans-signaling during inflammatory diseases^[31]. Sgp130 inhibits the activity of the IL-6/sIL-6R complex and is in competition with gp130 for this complex without interfering with the classical IL-6 signaling pathway (Figure 1)^[31-33]. A schematic of IL-6 classical and trans-signaling pathway is shown in Figure 1. In classical signaling, IL-6 binds first to the membrane-bound non-signaling IL-6R. After recruitment of two gp130 molecules the complex is formed and signal transduction is induced; in trans-signaling, IL-6 binds to the sIL-6R, which is generated by ectodomain shedding of the surface receptor or alternative splicing. The IL-6/sIL-6R can bind to both membrane-bound or sgp130 and a molar excess of sgp130 leads to competitive inhibition of the IL-6/sIL-6R response. Sgp130 has no access to the IL-6/IL-6R complex of the classical signaling pathway. Figure 1 was adapted from^[25,34,35].

IL-6 may have a crucial role in the growth and differentiation of malignant tumors. IL-6 has multiple effects on tumor progression, some are the result of autocrine activity on tumor cells and others are a consequence of paracrine action on normal cells in the tumor microenvironment, particularly osteoblasts, osteoclasts, endothelial cells and immune cells^[36]. For example, tumor cells from prostate, breast and colon cancer produce large amounts of IL-6 and express its receptors, IL-6R (gp80) and gp130, allowing them to respond in an autocrine manner to IL-6^[28]. Moreover, multiple myeloma and neuroblastoma cells do not produce IL-6R but express IL-6. These

cells respond in a paracrine manner to IL-6 present in the tumor microenvironment^[36].

IL-6 can also modulate the metastatic process. High IL-6 expression in specific organs such as lung, liver or brain may attract circulating tumor cells to these organs, promoting the development of metastases^[28]. Recently, it was demonstrated that the production of IL-6 and IL-8 in the primary tumor was responsible for the recruitment of circulating tumor cells to the primary site, resulting in a process called self-seeding which would lead to rapid tumor growth, angiogenesis, as well as stromal cell recruitment^[29].

An elevated serum IL-6 level has been correlated with adverse prognosis in patients with several different types of cancer, such as multiple myeloma^[37], lymphoma^[38], ovarian cancer^[39-41], PC^[42], metastatic renal carcinoma^[43], lung cancer^[44], and breast cancer^[45,46]. Hence, the interest in using serum IL-6 as a specific prognostic factor for PC and breast cancer has increased^[45,47].

On the other hand, trans-signaling through the IL-6/sIL-6R complex has been shown to have an important role in inflammatory diseases and in the development and progression of various malignant tumors^[34].

IL-6 trans-signaling has become a new area of research, and it was shown that sIL-6R is produced by various cancer cells, the serum concentration of sIL-6R is associated with decreased survival and increased aggressiveness of metastases in breast, prostate and colorectal cancers^[48-50]. The source of sIL-6R is not known, but it is shed by inflammatory cells including neutrophils, monocytes/macrophages and T cells^[51,52]. On the other hand, several tumor cells can shed IL-6R or produce it as a result of alternative splicing^[53]. Some data suggest that IL-6 trans-signaling causes various effects that promote cancer metastases including, increased detachment, proliferation

and migration^[54].

IL-6 has emerged as an important cytokine in the tumor microenvironment, which may contribute to the development and progression of various malignant tumors. Currently, little is known about the IL-6 trans-signaling pathway, and we suggest more studies to improve knowledge on the sIL-6R as a potential therapeutic target.

IL-6 PATHWAY IN PROSTATE CANCER

IL-6 serum levels and its receptors in prostate cancer patients

Involvement of IL-6 in PC development and progression has been suggested by several studies through increased IL-6 and sIL-6R levels during PC carcinogenesis and progression^[12-14]. Baillargeon *et al*^[15] suggested that serum and plasma IL-6 and sIL-6R levels are associated with progression and poor prognostic in PC patients.

Clinical observations have demonstrated increased IL-6 levels in plasma and serum from patients with CRPC^[55,56], metastatic PC^[42,48,57-59], biochemical recurrence^[42] and poorer overall survival^[15,60] compared to patients with earlier stages of the disease and healthy individuals.

Other studies support these findings and have shown that IL-6 can be correlated with the extension of disease, tumor size and bone metastases in PC^[48,61].

Akimoto *et al*^[57] found that serum levels of IL-6 were significantly higher in PC patients with bone metastasis than in PC patients without bone metastasis. These results suggested that serum levels of IL-6 were closely related to the metastatic burden of osseous tissue in PC patients^[57]. Adler *et al*^[58] reported that patients with metastatic PC had significantly elevated IL-6 levels when compared with those in other PC groups as well as controls. Of the 9 patients with distant metastatic disease (M1), 7 had elevated serum IL-6 levels. However, mean serum IL-6 was similar in patients with organ confined cancer (pT2) and in those with nodal metastases (N1). Eight of the 12 patients with N1 disease had decreased serum IL-6 compared to the pT2 group.

In another study, patients with metastatic PC were compared to patients with localized PC, and significant differences in IL-6 levels among the groups were observed. IL-6 levels were significantly higher in patients with metastatic disease. Patients with lymph node metastases or bone metastases had similar IL-6 levels. Serum IL-6 was significantly elevated in patients with Gleason score > 6^[59]. Shariat *et al*^[48] observed that the preoperative IL-6 and IL-6sR levels were elevated in patients with a final Gleason sum of 7 or greater. They reported that neither IL-6 nor IL-6sR were predictors of organ-confined disease. The mean preoperative IL-6 and sIL-6R plasma levels were higher in patients with aggressive disease than in those with non aggressive phenotype. IL-6 and sIL-6R plasma levels in patients with PC metastatic to bone were higher than those in patients with metastases in regional lymph nodes and these, in turn, were higher than those in prostatectomy patients and healthy subjects. However,

the IL-6 and IL-6sR levels were not different between the prostatectomy patients and the healthy subjects.

Regarding patients with CRPC, Drachenberg *et al*^[55] found that serum levels of IL-6 were significantly elevated in patients with clinically evident CRPC compared to normal controls, patients with BPH, prostatitis, and localized or recurrent disease. Wise *et al*^[56] observed elevated levels of the anti-inflammatory cytokine, IL-6, in CRPC patients when compared to patients with CSPC.

The results obtained by George *et al*^[60] showed that IL-6 levels were higher in patients with metastatic disease and these patients had worse survival when compared with patients who had low IL-6 levels. An analysis by Nakashima *et al*^[15] verified a significantly shorter survival in patients with elevated IL-6 levels, PSA serum levels and aggressive disease.

Additionally, Shariat's^[48,62,63] group demonstrated that preoperative plasma levels of IL-6, sIL-6R and transforming growth factor-beta1 (TGF- β 1) predicted biochemical recurrence after surgery or radical prostatectomy, suggesting an association with occult metastatic disease at the time of radical prostatectomy. The results of Alcover *et al*^[64] are in agreement with Shariat's group, as they observed that IL-6 predicts biochemical recurrence following radical prostatectomy. More recently, a predictive model was proposed that included only sIL-6R^[65]. Kattan *et al*^[66] developed and validated a prognostic model that added plasma sIL-6R and TGF- β 1 as standard clinical predictors for biochemical recurrence. However, Baillargeon *et al*^[6] found that serum IL-6 levels were not associated with PC. According to Finley *et al*^[67] the differences among various studies are a consequence of systemic cytokine levels detected in peripheral blood that may not reflect local concentrations in the tumor microenvironment.

IL-6 levels can also be elevated in obese individuals^[68]. The relationship between obesity, circulating IL-6 and PC may help to understand the role of this molecule and how it contributes to the molecular basis for the association between obesity and PC^[3,69-71]. Adipose tissue is a highly active endocrine tissue that secretes numerous factors, including growth factors, cytokines, hormone-like molecules and many other molecules^[72]. Several studies have demonstrated that the abundant number of growth factors such as vascular endothelial growth factor (VEGF), inflammatory cytokines [IL-6, tumor necrosis factor-alpha, interleukin-8 (IL-8)] and adipokines (adiponectin, leptin) released from adipose tissue can exert a substantial impact on the progression and outcome of many human diseases, including PC^[3,16,71]. These molecules have a crucial role in obesity^[73] and cell proliferation^[74]. For instance, adipose tissue surrounding the prostate, i.e., periprostatic adipose tissue, is frequently invaded by prostate tumor cells, although its contribution to the tumor microenvironment is largely unknown. A recent study demonstrated that periprostatic adipose tissue produced local IL-6 at levels significantly higher than those in the circulation^[67]. Other studies support a role for IL-6

production, which is up-regulated in obese patients, in PC cell migration and invasiveness^[67,71].

Recently, Shariat *et al.*^[75] in a study of 423 preoperative and 206 postoperative blood samples from patients treated with radical prostatectomy for clinically localized PC investigated the association between sgp130 levels and PC prognosis. In the group of patients treated with radical prostatectomy, higher preoperative plasma sgp130 was significantly associated with higher pathological Gleason, extraprostatic extension, seminal vesicle invasion, lymph node metastasis and biochemical recurrence. These authors concluded that higher sgp130 plasma levels were associated with features of biologically aggressive PC. In a subset of 206 patients, postoperative sgp130 levels were 18% lower than preoperative levels. These decreased levels suggest that the higher blood levels of sgp130 are produced by tumor cells^[75].

These clinical data support the biological role of the IL-6 pathway in PC, suggesting the inclusion of IL-6, sIL-6R and gp130 levels as new tumor biomarkers in PC patients^[76]. Improving prediction accuracy by using more prognostic factors supports an early detection of any changes in the progression of the disease^[25].

IL-6 expression in prostate cancer tissues and cell lines

The expression of IL-6 and its receptors has been investigated by several authors in benign prostate cells, PC tissues and in prostate cell lines (LNCaP, DU145, PC3). Hobisch *et al.*^[14] through immunohistochemical studies revealed that IL-6 expression was localized predominantly in basal cells of benign prostatic epithelium and in glandular cells of PC tissues. Another study observed that in normal prostate, IL-6 was immunolocalized in basal cells of the epithelium and gp130 was detected only in stromal cells^[77]. However, Hobish *et al.*^[14] showed that cultured stromal cells secreted IL-6, but the rate of IL-6 was so low that there was only a minimal amount contained in the cells, and thus it may not be detectable by immunohistochemical methods. In benign prostate cells, gp130 was confined to the epithelium and stroma, and IL-6 was immunolocalized preferentially in epithelium^[77]. These data are in agreement with Degeorges *et al.*^[78] who demonstrates that IL-6 is secreted by cultured benign prostate cells. In PC tissues, gp130 was detected in stroma and epithelium and the expression increased with Gleason Grade^[77]. On the other hand, IL-6 was localized in all cell types and immunostaining increased with Gleason Grade^[77]. These results are in agreement with the reported secretion of IL-6 by PC cells^[79] and with increased IL-6 levels in PC patients with poor prognosis^[42,48]. Palmer *et al.*^[80] observed that IL-6 and IL-6 receptors are expressed in PC cells. They showed that in LNCaP, DU145 and PC3 cell lines, this cytokine and its receptors are widely expressed, but not in normal prostate epithelial PZ-HPV-7 cells.

The role of the pro-inflammatory cytokine, IL-6, in PC lesions has not yet been clarified but may represent an interesting area of investigation. In order to delineate

their specific functions during prostate tumor development and progression, several authors used human PC cell lines (LNCaP, DU145 and PC3)^[17,81-88].

Significance of IL-6 pathway in cell growth: In vitro results

The castration-sensitive cell line, LNCaP, is one the most frequently used in PC studies. In order to improve our understanding of cellular events which may be relevant to PC patients with higher IL-6 levels, several authors treated LNCaP cells with IL-6^[81,82].

Okamoto *et al.*^[82] found that growth of the LNCaP cell line was stimulated by the administration of IL-6, but not by their conditioned medium. Conditioned medium containing biologically active components (e.g., growth factors, cytokines) was obtained from previously cultured cells or tissues released into the media, substances that affect certain cell functions. These authors observed that DU145 and PC3 (castration-resistant) cell lines proliferate in response to stimulation with IL-6 and in response to its conditioned medium. The authors concluded that IL-6 acts as an autocrine and/or paracrine proliferative factor in PC cell lines^[82]. It was demonstrated that IL-6 acts as a paracrine inducer of growth in the LNCaP cell line^[17,79,82,83]. On the other hand, IL-6 functions as an autocrine growth-inducer in the DU145 and PC3 cell lines^[82,84]. Chung *et al.*^[85] obtained similar results, finding that IL-6 acts as a growth inducer in PC3 and DU145 cell lines through an autocrine and paracrine action. However, the effect of IL-6 in LNCaP cells is still controversial. Some studies reported that IL-6 acts as an inducer of growth in this cell line^[82,83,86], while others showed that IL-6 acts as an inhibitor^[81,85,87]. The reason for these differences between the various studies is that IL-6 may have different functions in human PC cell line proliferation, according to their phenotypic characteristics.

Hobisch *et al.*^[81] generated a cell line (LNCaP-IL-6+) by exposing these cells to continuous administration of IL-6 and observed changes in their responsiveness and signal transduction. Initially, growth of LNCaP was inhibited by IL-6. After long-term treatment, the LNCaP-IL-6+ cell line began to secrete IL-6 and a higher basal proliferation rate was observed. In this situation, IL-6 induces cell growth.

Recently, there is evidence to suggest that IL-6 switches from a paracrine growth inhibitor to an autocrine growth stimulator^[12,81,88]. Similar changes in responsiveness to IL-6 were observed in melanoma cells^[89]. The behavior of the human prostate carcinoma LNCaP cell line may be dependent on the microenvironment of the culture system^[12].

A recent study by Shariat *et al.*^[75] showed that sgp130 promotes PC invasiveness *in vitro*. In this study, continuous exposure of PC cells to sgp130 led to an increase in their invasiveness^[75]. These authors suggest more studies are required with regard to the role of IL-6 and sgp130 in the PC biological behavior *in vitro* and *in vivo* for a better understanding of their role in this disease^[75].

HORMONAL CASTRATION IN ADVANCED PROSTATE CANCER PATIENTS: ANDROGEN RECEPTOR AND IL-6 PATHWAYS

Initial treatment for organ-confined PC is usually radical prostatectomy and/or radiation therapy^[90]. Although, most patients have advanced disease and are submitted to hormonal castration (HC)^[90]. The initial response rate is excellent, but the majority of PC patients relapse into CRPC. CRPC is a common lethal form of PC that typically metastasizes to bone and visceral organs, frequently resulting in patient death^[91,92]. Progression to metastatic disease is slow and can be accompanied by increased PSA levels^[93]. The mechanisms implicated in CRPC progression are unknown. These findings suggest that CRPC progression remains the major obstacle to effective control and cure of advanced phenotype disease. Consequently, novel therapeutic strategies that target the molecular mechanism involved in CRPC are needed^[94].

PC cells are androgen-dependent, in particular, testosterone is necessary for their growth and survival^[91]. Thus, the blockade of testosterone initially causes a stop in PC cell growth^[95]. However, not all tumor cells need testosterone for development. After HC, many tumors begin to exhibit a testosterone blockade resistance behavior. In this situation, CSPC cells undergo apoptosis and there is a selective advantage for CRPC cells with consequent proliferation^[96].

The androgen receptor (AR) is an important protein involved in the normal maintenance, development and growth of prostate epithelial cells. AR is a nuclear ligand-activated transcription factor in the prostate gland and mediates the biological response of androgens with a crucial function in a molecular mechanism responsible for the transition from CSPC to CRPC progression^[97-99]. The AR is expressed in the normal prostate and during various stages of prostate carcinogenesis (PIN, organ-confined tumors, metastatic tumors and before or after hormonal treatment)^[100,101].

The AR uses di-hydrotestosterone (DHT) and testosterone as its natural ligands for phosphorylation, and the ligand-receptor complex translocates into the nucleus where it binds to a DNA sequence in the regulatory regions of AR target genes^[99]. These complex interactions facilitate the activation or repression of the expression of several genes involved in the development, differentiation and proliferation of target cells. Some of these genes include *PSA* and *human glandular kallikrein 2 (hk2)*^[102].

During CRPC development, PC cells can develop alternative mechanisms which can influence their microenvironment and, consequently, their survival in an androgen-poor microenvironment^[103]. Several models have been proposed to explain the development of this phenotype: hypersensitive pathways, promiscuous receptor, coactivators and corepressors, bypass pathway, PC stem cells and outlaw pathways^[91,97,104-107].

In the hypersensitive pathways, the cells acquire the ability to use very low levels of androgen during HC^[97]. There are three mechanisms that may or probably are involved in this pathway: (1) AR amplification, PC cells have increased expression of AR^[103]. The increased AR expression allows higher ligand binding. This hypothesis is supported by results obtained with CRPC cell lines that show an increased expression of AR compared with CSCP^[97]; (2) increased AR sensitivity, consequently, tumor cells are hypersensitive to the growth promoting effects of DHT^[108]; and (3) increased 5-alpha-reductase activity that increases the conversion of testosterone to DHT^[91].

The promiscuous pathway involves acquisition of mutations in the AR protein, consequently, the AR can be activated by nonandrogenic steroid molecules that are present in the circulation^[91,97,105,106,109]. There are a large number of coactivators and corepressors involved in regulation of the AR^[110]. These molecules are intermediates between the AR signaling pathway and transcriptional machinery^[103]. In the bypass pathway, PC cells acquire a phenotype that allow them to survive and escape apoptosis in an androgen-depleted environment^[106]. Another bypass mechanism is correlated with neuroendocrine differentiation of PC cells^[106]. These cells are more represented in CRPC cells and are associated with a low rate of proliferation^[106]. Neuroendocrine cells have the capacity to increase the proliferation of surrounding cells, thus progression of the PC cells occurs in a androgen-poor microenvironment^[106].

The PC stem cells model showed that only a rare subset of cells is tumorigenic^[111]. Collis *et al.*^[112] reported that a population of cells comprising 0.1% of prostate tumors (CD44+/a2h1/CD133+) without AR expression may be prostate cancer stem or progenitor cells. There is also another possible mechanism for cell survival in an androgen-poor microenvironment, where the presence of prostate cancer stem cells continually resupply the tumor cell population and are not affected by HC. These cells are capable of differentiating into androgen-dependent and -independent cells, leading to the development of a heterogeneous androgen receptor phenotype. This phenotype is typical in patients with CRPC^[103,104].

Another potential hypothesis (outlaw pathway), is that during HC the AR can be activated by other nonsteroidal molecules synthesized and secreted by tumor cells, such as cytokines and growth factors [e.g., Keratinocyte Growth Factor, Epidermal Growth Factor, Insulin-like Growth Factor-1 and IL-6]^[113-115], leading to CRPC development and progression. Even if androgens are principally responsible for activating the AR, it is known that in the absence or presence of very low concentrations of androgens, the AR can be activated by growth factors and cytokines^[113]. In prostate tumor, the microenvironment is secreting growth factors and cytokines which may directly manipulate paracrine and autocrine pathways involved in PC development and promote CRPC in patients treated with hormonal therapy^[116].

Some studies have examined the IL-6 pathway in

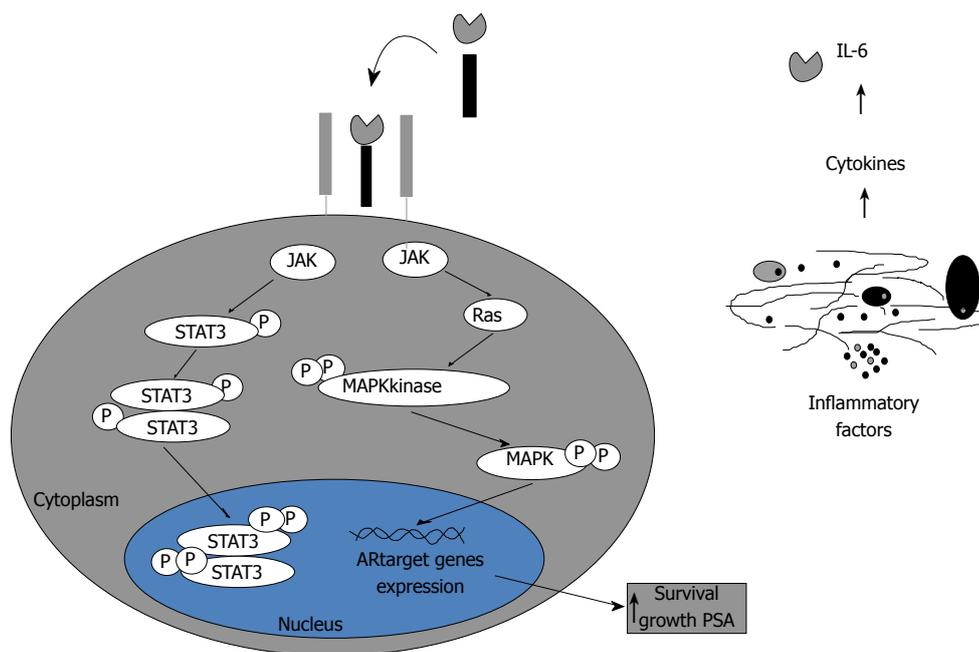


Figure 2 Hypothetical representation of AR pathway regulation by IL-6 in prostate cancer cells. IL-6: Interleukin 6; MAPK: Mitogen-activated protein kinase; STAT3: Signal transducer and activator of transcription 3; AR: Androgen receptor; PSA: Prostate-specific antigen.

CRPC progression and have revealed a possible involvement in regulation of the AR^[117-119]. In the absence of androgen, IL-6 causes activation of the AR which is approximately 50% of the maximal activity induced by androgen. In low concentration conditions, androgen is potentiated by IL-6, leading to synergistic activation of the AR^[93]. These observations demonstrate a cross-talk between the IL-6 pathway and AR. Due to the presence of increased IL-6 levels in patients with locally advanced or metastatic disease, the signaling pathway mediated by this cytokine could be an alternative pathway in the RCH phenotype acquisition and progression. Figure 2 shows that IL-6 activates the AR through a mechanism that is dependent on the MAPK and STAT3 signaling pathway in PC cells. This is a hypothetical representation of the AR pathway regulation by IL-6 in PC cells. The IL-6 pathway may be an alternative pathway for growth and proliferation of PC cells, under androgen deprivation conditions. IL-6 induces up-regulation of AR target gene expression such as PSA by STAT3 and MAPK signal transduction pathways. Figure 2 was adapted from^[99,116].

We consider IL-6, other cytokines and growth factors as important regulators of PC growth with a significant role in the AR pathway. It has been proposed that the combined blockage of key molecules in proliferation signaling pathways (e.g., TGF, IL-6) could be one of the most promising strategies for CRPC^[120]. Consequently, several authors have been investigating IL-6 activity in PC cells *in vitro* and *in vivo*. *In vitro* research on the regulation of PC cell growth and transcriptional activation of the AR by IL-6 have been focused on cell lines (LNCaP, DU145, PC3) and different results were obtained^[117,82-84].

***In vitro* and *in vivo* studies on androgen receptor regulation by IL-6**

The interaction between IL-6 and the AR may be more important in advanced PC patients who have elevated serum levels of IL-6 and its receptors. Several *in vitro* studies identified alternative pathways that influence or modify the activity of the AR signaling pathway in PC cells. Cross-talk between IL-6 and the AR was investigated in PC cells which transiently express the AR (DU-145) and in LNCaP cells which have a promiscuous mutated AR. Results on IL-6 induction, AR activation and tumor proliferation are contradictory. This is probably due to controversial results on the induction or stimulation effect of IL-6 on PC cell growth^[121].

In LNCaP and DU-145 cell lines, IL-6 activates the AR in a ligand-independent and synergistic manner in low androgen concentrations^[119]. This fact might seem paradoxical because proliferation of LNCaP was inhibited by IL-6. However, it has been suggested that IL-6 is an important molecule in the maintenance and differentiation of PC cells and this cytokine is involved in enhanced PSA mRNA regulation and in the regulation of its protein^[93]. It was observed that long-term treatment with IL-6 in LNCaP cells increased expression and activity of the AR^[117,119] and overexpression of IL-6 protects LNCaP cells from apoptosis induced by HC^[93].

In MDA PC 2b cells (androgen-sensitive cell line), IL-6 promotes growth and this effect was dependent of the AR^[76]. It is known that AR signaling is complex and there are several AR-associated proteins with a crucial role in the modulation of response in a cell type-dependent manner^[122]. For this reason, IL-6 up-regulation has been investigated in PC. Additionally, authors hypoth-

esized that IL-6 may promote tumor growth through AR activation *in vivo*. In their work, MDA PC 2b cells were xenografted into nude mice. They observed that growth of MDA PC 2b xenografts in castrated animals treated with IL-6 was similar to that in non-castrated animals. In addition, tumors did not significantly grow in castrated mice and mice treated with IL-6 and bicalutamide (oral non-steroidal anti-androgen used in the treatment of PC). Bicalutamide showed an inhibitory effect on IL-6-regulated growth *in vivo*^[76].

This evidence demonstrated that IL-6 has a crucial role in androgen-responsive gene expression and, consequently, is an important regulator in the growth of CRPC cells *in vitro* and *in vivo*. Another recent study evaluated the effects of IL-6 in the LNCaP cell line on phenotype changes before and after androgen deprivation^[94]. *In vitro* observations indicate that the growth of LNCaP/IL-6 (IL-6-transfected LNCaP) was significantly lower than LNCaP/Co (not IL-6-transfected LNCaP) under androgen-deprivation conditions. Furthermore, LNCaP/IL-6 tumors in nude mice rapidly regressed after castration. However, LNCaP/Co tumor growth was transiently inhibited after castration and then continuously accelerated^[94]. Gene microarray analyses showed that androgen deprivation resulted in the differential expression of genes involved in growth, apoptosis and carcinogenesis between LNCaP/Co and LNCaP/IL-6^[94]. The principal conclusion of this study is that IL-6 produced by LNCaP may have a suppressive role on growth and a crucial function in the androgen-resistance phenotype under an androgen-poor microenvironment^[94]. These and other future investigations will clarify the molecular mechanism involved in changes of phenotype in LNCaP cells that express higher IL-6 levels.

Efficient activation of the AR by IL-6 involves multiple signaling pathways such as STAT3, PI3K and MAPK. At present, little is known about these mechanisms. Some studies have used inhibitors of these signaling pathways. Inhibitors of the JAK, MAPK and PI3K signaling pathways resulted in a down-regulation of IL-6 on the AR^[119]. Lin *et al*^[117] showed that MAPK is important for AR activity through IL-6. These authors demonstrated that an inhibitor of MAPK abrogated IL-6 activation of the PSA promoter. In contrast, an inhibitor of the PI3K pathway had no effect on IL-6 regulation of AR activity^[117]. Chen *et al*^[118] also showed that JAK inhibition stopped AR activation by IL-6. This study demonstrated that the JAK-STAT pathway is implicated in AR activation by IL-6 in the LNCaP cell line. On the other hand, Zelivianski *et al*^[123] demonstrated that IL-6 up-regulated AR protein levels in low passage LNCaP cells, while in high passage LNCaP cells the opposite effect was observed.

Recent research in prostate carcinoma has focused on the role of sIL-6R. The trans-signaling pathway mediated by complex IL-6/sIL-6R seems to have an important role in the pathophysiology of certain inflammatory, nervous system and cardiovascular diseases and some cancer types^[54]. The IL-6/sIL-6R complex can have an antiapop-

otic role and is therefore considered a possible cause of certain cancers such as colon cancer and melanoma^[26,34,52,124-127]. To the best of our knowledge, the classical and trans-signaling pathways in specific PC events are not well known. Furthermore, sIL-6R showed a stronger association with disease progression than IL-6^[54], suggesting a role of complex IL-6/sIL-6R in the spread of metastases. Santer *et al*^[54] conducted a study with the principal aim of determining the effect of complex IL-6/sIL-6R on PC cell proliferation and in metastasis formation. These authors focused their studies on sIL-6R, because it is the first element in the IL-6 signaling pathway. They found that activation of sIL-6R resulted in increased PC cell motility and migration^[54]. Thereby, it is believed that sIL-6R may be important in the metastatic process through down-regulation of the tumor suppressor, maspin, by sIL-6R. In contrast, the IL-6 trans-signaling pathway reduces PC cell proliferation^[54].

It was suggested that targeting sIL-6R may be an alternative method of improving anti-IL-6 therapies used in PC treatment^[54]. Understanding how the IL-6 pathway affects cellular events in the PC cell microenvironment and its interaction with the AR pathway will allow the development of preventive and therapeutic strategies for PC patients in the future.

IL-6 TARGETING

The importance of the IL-6/IL-6R pathway in the regulation of PC cells and the potential involvement of this signaling pathway in androgen-resistant growth of PC cells makes it a good candidate for targeted therapy. In this perspective, the use of IL-6-neutralization antibodies, antisense oligonucleotides and antagonists should be the subject of study^[88]. The principal goal is to identify new therapies to target tumor cells and/or microenvironment and consequently increase the chances of survival for PC patients with aggressive phenotypes and those who develop resistance to hormonal therapy.

It has been reported that IL-6R blockade by IL-6R antagonists might reduce tumor cell growth and consequently disease progression^[128]. The IL-6/IL-6R signaling pathway involves numerous proteins and a large number of phosphorylation cascade pathways. Downstream molecules of these proteins and pathways may be crucial targets for specific therapies. For example, inhibition of STAT3 suppresses PC progression^[129] and reduces STAT3 target gene expression, such as VEGF, Bcl-X and cyclin D1 and leads to apoptosis^[130].

Other investigations are involved in the study of the chimeric monoclonal anti-IL-6 antibody, siltuximab (CNT0 328). Steiner *et al*^[131] showed that tumor growth in nude mice inoculated with LNCaP-IL-6+ cells after CNT0 328 treatment was reduced. Other studies obtained analogous results when PC3 and LuCaP 35 xenografts were treated with CNT0 328^[132,133]. Another study reported that CNT0 328 can inhibit PC cell growth *in vitro* and improve survival by reducing the level of ca-

Table 1 Summary of several studies on IL-6, sIL-6R and sgp130 levels in prostate cancer patients and targeted therapies for the IL-6 signaling pathway

	Ref.	Conclusions
Prognostic implications	[15,16]	L-6 level is a significant prognostic factor for PC. A significantly shorter survival in PC patients was associated with elevated IL-6 levels, serum PSA levels and aggressive disease
	[48,57-60]	The serum levels of IL-6 were significantly higher in PC patients with metastatic disease
	[48,62-66]	The levels of IL-6, sIL-6R and TGF-β1 predicted biochemical recurrence after surgery or radical prostatectomy
	[75]	In patients treated with radical prostatectomy higher preoperative plasma sgp130 was significantly associated with higher pathological Gleason, extraprostatic extension, seminal vesicle invasion, lymph node metastasis and biochemical recurrence. The postoperative sgp130 levels were 18% lower than preoperative levels
Therapeutic implications	[131-133]	These studies involved the chimeric monoclonal anti-IL-6 antibody, siltuximab (CNT0 328) It was shown that tumor growth in nude mice inoculated with LNCaP-IL-6+ cells after CNT0 328 treatment was reduced. Analogous results were obtained when PC3 and LuCaP 35 xenografts were treated with CNT0 328
	[135]	The administration of siltuximab in a group of patients who had already received docetaxel therapy had no clinical efficacy
	[136]	PC cells can develop resistance to docetaxel and STAT1 is increasingly expressed in docetaxel-resistant PC cells
	[84]	The treatment of the PC3 cell line with Sant7 inhibits cell growth more efficiently than other anti-IL-6 antibodies
	[134]	CNT0 328 can inhibit PC cell growth <i>in vitro</i> and improve survival by reducing the level of cachexia in an animal model of PC
	[133]	In mice, CNT0 328 inhibited the conversion of CSPC into more aggressive disease
	[137]	STAT3 and MAPK activity is suppressed in patients taking siltuximab, which may inhibit IL-6-mediated drug resistance

PC: Prostate cancer; CSPC: Castration-sensitive prostate cancer; TGF-β1: Transforming growth factor-beta1; IL-6: Interleukin-6; sIL-6R: Soluble interleukin-6 receptor; STAT3: Signal transducer and activator of transcription 3; MAPK: Mitogen-activated protein kinase.

chexia in an animal model of PC^[134]. In addition, CNT0 328 has been shown in mice to inhibit the conversion of CSPC into more aggressive disease, bone metastasis, and difficult to treat CRCP^[133].

In a clinical trial, it was shown that the administration of siltuximab in a group of patients who had already received docetaxel therapy had biological, but not clinical efficacy^[135]. PC cells can develop resistance to docetaxel, and Patterson *et al.*^[136] reported that STAT1 is increasingly expressed in docetaxel-resistant PC cells. The high heterogeneity of prostate tumors can explain this resistance to docetaxel in PC cells^[11]. Recent studies indicate that STAT3 and MAPK activity is suppressed in patients taking siltuximab, which may inhibit IL-6-mediated drug

resistance^[137]. However, in a study that involved patients with CRPC, where the disease had progressed beyond docetaxel therapy, siltuximab had a minimal clinical effect, despite positive biological IL-6 inhibition^[135].

Lou *et al.*^[83] reported that targeting IL-6 may have multiple advantages in patients that receive limited therapeutic and survival benefit from conventional therapies. In a previous study, Borsellino *et al.*^[84] found that treatment of the PC3 cell line with Sant7, a modified interleukin-6 which binds with high affinity to IL-6R but does not bind with gp130, inhibits cell growth more efficiently than other anti-IL-6 antibodies.

We believe that the development and availability of IL-6 inhibitors is fundamental for the treatment of IL-6-dependent cancers, where the IL-6 signaling pathway is deregulated. This review demonstrates the role of IL-6 and the levels of its receptors as prognostic factors in PC patients. These deregulated levels could be important in anti-IL-6 therapy development. Table 1 presents a summary of several studies on IL-6, sIL-6R and sgp130 levels in PC patients and targeted therapies for the IL-6 signaling pathway. However, more studies and appropriate clinical trials are needed to determine the effectiveness of anti-IL-6 therapies in cancer patients.

CONCLUSION

The information presented in this review suggests that the IL-6 signaling pathway plays an important role in PC development/progression, and IL-6 is able to maintain tumor growth through the AR pathway in androgen-deprived conditions. Further studies are suggested to assess the functionality of the IL-6/sIL-6R complex in PC. Understanding how IL-6 affects cellular events in the PC cell microenvironment and its interaction with the AR pathway will allow the development of preventive and therapeutic strategies for PC patients in the future. However, it is also important to study and characterize other signaling pathways involved in CRPC progression and the cross-talk among them, allowing the design of new and more adequate targeted therapies. Additionally, diverse studies reported that serum and plasma levels of IL-6 and sIL-6R are increased in patients with aggressive disease and a poor prognosis, suggesting the inclusion of IL-6 and the levels of its receptors as putative new tumor biomarkers. In addition, changes in serum IL-6 levels could help direct additional treatment strategies in the future, however, clinical studies are needed to assess this potential.

In conclusion, IL-6 is a good candidate for the development of targeted therapies in PC, but more studies and appropriate clinical trials need to be carried out to ascertain the effectiveness of anti-IL-6 therapies in PC patients.

REFERENCES

- 1 Hsing AW, Chokkalingam AP. Prostate cancer epidemiology. *Front Biosci* 2006; **11**: 1388-1413

- 2 **Platz EA**, De Marzo AM. Epidemiology of inflammation and prostate cancer. *J Urol* 2004; **171**: S36-S40
- 3 **Ribeiro R**, Lopes C, Medeiros R. The link between obesity and prostate cancer: the leptin pathway and therapeutic perspectives. *Prostate Cancer Prostatic Dis* 2006; **9**: 19-24
- 4 **Mistry K**, Cable G. Meta-analysis of prostate-specific antigen and digital rectal examination as screening tests for prostate carcinoma. *J Am Board Fam Pract* 2003; **16**: 95-101
- 5 **Nash AF**, Melezinek I. The role of prostate specific antigen measurement in the detection and management of prostate cancer. *Endocr Relat Cancer* 2000; **7**: 37-51
- 6 **Draisma G**, Etzioni R, Tsodikov A, Mariotto A, Wever E, Gulati R, Feuer E, de Koning H. Lead time and overdiagnosis in prostate-specific antigen screening: importance of methods and context. *J Natl Cancer Inst* 2009; **101**: 374-383
- 7 **Schröder FH**, Hugosson J, Roobol MJ, Tammela TL, Ciatto S, Nelen V, Kwiatkowski M, Lujan M, Lilja H, Zappa M, Denis LJ, Recker F, Berenguer A, Mänttinen L, Bangma CH, Aus G, Villers A, Biehlard X, van der Kwast T, Blijenberg BG, Moss SM, de Koning HJ, Auvinen A. Screening and prostate-cancer mortality in a randomized European study. *N Engl J Med* 2009; **360**: 1320-1328
- 8 **Welch HG**, Albertsen PC. Prostate cancer diagnosis and treatment after the introduction of prostate-specific antigen screening: 1986-2005. *J Natl Cancer Inst* 2009; **101**: 1325-1329
- 9 **Ferlay J**, Shin HR, Bray F, Forman D, Mathers C, Parkin DM. Estimates of worldwide burden of cancer in 2008: GLOBOCAN 2008. *Int J Cancer* 2010; Jun 17 [Epub ahead of print]
- 10 **Germano G**, Allavena P, Mantovani A. Cytokines as a key component of cancer-related inflammation. *Cytokine* 2008; **43**: 374-379
- 11 **Culig Z**, Puhm M. Interleukin-6: A multifunctional targetable cytokine in human prostate cancer. *Mol Cell Endocrinol* 2011; Jun 1 [Epub ahead of print]
- 12 **Culig Z**, Steiner H, Bartsch G, Hobisch A. Interleukin-6 regulation of prostate cancer cell growth. *J Cell Biochem* 2005; **95**: 497-505
- 13 **Hong DS**, Angelo LS, Kurzrock R. Interleukin-6 and its receptor in cancer: implications for translational therapeutics. *Cancer* 2007; **110**: 1911-1928
- 14 **Hobisch A**, Rogatsch H, Hittmair A, Fuchs D, Bartsch G, Klocker H, Bartsch G, Culig Z. Immunohistochemical localization of interleukin-6 and its receptor in benign, pre-malignant and malignant prostate tissue. *J Pathol* 2000; **191**: 239-244
- 15 **Nakashima J**, Tachibana M, Horiguchi Y, Oya M, Ohigashi T, Asakura H, Murai M. Serum interleukin 6 as a prognostic factor in patients with prostate cancer. *Clin Cancer Res* 2000; **6**: 2702-2706
- 16 **Baillargeon J**, Platz EA, Rose DP, Pollock BH, Ankerst DP, Haffner S, Higgins B, Lokshin A, Troyer D, Hernandez J, Lynch S, Leach RJ, Thompson IM. Obesity, adipokines, and prostate cancer in a prospective population-based study. *Cancer Epidemiol Biomarkers Prev* 2006; **15**: 1331-1335
- 17 **Lee SO**, Lou W, Hou M, de Miguel F, Gerber L, Gao AC. Interleukin-6 promotes androgen-independent growth in LNCaP human prostate cancer cells. *Clin Cancer Res* 2003; **9**: 370-376
- 18 **Kishimoto T**, Akira S, Narazaki M, Taga T. Interleukin-6 family of cytokines and gp130. *Blood* 1995; **86**: 1243-1254
- 19 **Fain JN**, Madan AK, Hiler ML, Cheema P, Bahouth SW. Comparison of the release of adipokines by adipose tissue, adipose tissue matrix, and adipocytes from visceral and subcutaneous abdominal adipose tissues of obese humans. *Endocrinology* 2004; **145**: 2273-2282
- 20 **Yamasaki K**, Taga T, Hirata Y, Yawata H, Kawanishi Y, Seed B, Taniguchi T, Hirano T, Kishimoto T. Cloning and expression of the human interleukin-6 (BSF-2/IFN beta 2) receptor. *Science* 1988; **241**: 825-828
- 21 **Hibi M**, Murakami M, Saito M, Hirano T, Taga T, Kishimoto T. Molecular cloning and expression of an IL-6 signal transducer, gp130. *Cell* 1990; **63**: 1149-1157
- 22 **Peters M**, Müller AM, Rose-John S. Interleukin-6 and soluble interleukin-6 receptor: direct stimulation of gp130 and hematopoiesis. *Blood* 1998; **92**: 3495-3504
- 23 **Rose-John S**. Coordination of interleukin-6 biology by membrane bound and soluble receptors. *Adv Exp Med Biol* 2001; **495**: 145-151
- 24 **Rose-John S**, Scheller J, Elson G, Jones SA. Interleukin-6 biology is coordinated by membrane-bound and soluble receptors: role in inflammation and cancer. *J Leukoc Biol* 2006; **80**: 227-236
- 25 **Tawara K**, Oxford JT, Jorcyk CL. Clinical significance of interleukin (IL)-6 in cancer metastasis to bone: potential of anti-IL-6 therapies. *Cancer Manag Res* 2011; **3**: 177-189
- 26 **Kallen KJ**. The role of transsignalling via the agonistic soluble IL-6 receptor in human diseases. *Biochim Biophys Acta* 2002; **1592**: 323-343
- 27 **Mullberg J**, Schooltink H, Stoyan T, Gunther M, Graeve L, Buse G, Mackiewicz A, Heinrich PC, Rose-John S. The soluble interleukin-6 receptor is generated by shedding. *Eur J Immunol* 1993; **23**: 473-480
- 28 **Knüpfner H**, Preiss R. sIL-6R: more than an agonist? *Immunol Cell Biol* 2008; **86**: 87-91
- 29 **Lust JA**, Donovan KA, Kline MP, Greipp PR, Kyle RA, Maihle NJ. Isolation of an mRNA encoding a soluble form of the human interleukin-6 receptor. *Cytokine* 1992; **4**: 96-100
- 30 **Gaillard JP**, Bataille R, Brailly H, Zuber C, Yasukawa K, Attal M, Maruo N, Taga T, Kishimoto T, Klein B. Increased and highly stable levels of functional soluble interleukin-6 receptor in sera of patients with monoclonal gammopathy. *Eur J Immunol* 1993; **23**: 820-824
- 31 **Jostock T**, Müllberg J, Ozbek S, Atreya R, Blinn G, Voltz N, Fischer M, Neurath MF, Rose-John S. Soluble gp130 is the natural inhibitor of soluble interleukin-6 receptor transsignaling responses. *Eur J Biochem* 2001; **268**: 160-167
- 32 **Müller-Newen G**, Küster A, Hemmann U, Keul R, Horsten U, Martens A, Graeve L, Wijdenes J, Heinrich PC. Soluble IL-6 receptor potentiates the antagonistic activity of soluble gp130 on IL-6 responses. *J Immunol* 1998; **161**: 6347-6355
- 33 **Atreya R**, Mudter J, Finotto S, Müllberg J, Jostock T, Wirtz S, Schütz M, Bartsch B, Holtmann M, Becker C, Strand D, Czaja J, Schlaak JF, Lehr HA, Autschbach F, Schürmann G, Nishimoto N, Yoshizaki K, Ito H, Kishimoto T, Galle PR, Rose-John S, Neurath MF. Blockade of interleukin 6 trans signaling suppresses T-cell resistance against apoptosis in chronic intestinal inflammation: evidence in crohn disease and experimental colitis in vivo. *Nat Med* 2000; **6**: 583-588
- 34 **Mitsuyama K**, Sata M, Rose-John S. Interleukin-6 transsignaling in inflammatory bowel disease. *Cytokine Growth Factor Rev* 2006; **17**: 451-461
- 35 **Chalaris A**, Garbers C, Rabe B, Rose-John S, Scheller J. The soluble Interleukin 6 receptor: generation and role in inflammation and cancer. *Eur J Cell Biol* 2011; **90**: 484-494
- 36 **Ara T**, Declerck YA. Interleukin-6 in bone metastasis and cancer progression. *Eur J Cancer* 2010; **46**: 1223-1231
- 37 **Ludwig H**, Nachbaur DM, Fritz E, Krainer M, Huber H. Interleukin-6 is a prognostic factor in multiple myeloma. *Blood* 1991; **77**: 2794-2795
- 38 **Seymour JF**, Talpaz M, Cabanillas F, Wetzler M, Kurzrock R. Serum interleukin-6 levels correlate with prognosis in diffuse large-cell lymphoma. *J Clin Oncol* 1995; **13**: 575-582
- 39 **Berek JS**, Chung C, Kaldi K, Watson JM, Knox RM, Martínez-Maza O. Serum interleukin-6 levels correlate with disease status in patients with epithelial ovarian cancer. *Am J Obstet Gynecol* 1991; **164**: 1038-1042; discussion 1038-1042
- 40 **Plante M**, Rubin SC, Wong GY, Federici MG, Finstad CL, Gastl GA. Interleukin-6 level in serum and ascites as a prognostic factor in patients with epithelial ovarian cancer. *Cancer* 1994; **73**: 1882-1888

- 41 **Hefler LA**, Grimm C, Ackermann S, Malur S, Radjabi-Rahat AR, Leodolter S, Beckmann MW, Zeillinger R, Koelbl H, Tempfer CB. An interleukin-6 gene promoter polymorphism influences the biological phenotype of ovarian cancer. *Cancer Res* 2003; **63**: 3066-3068
- 42 **Twillie DA**, Eisenberger MA, Carducci MA, Hseih WS, Kim WY, Simons JW. Interleukin-6: a candidate mediator of human prostate cancer morbidity. *Urology* 1995; **45**: 542-549
- 43 **Blay JY**, Negrier S, Combaret V, Attali S, Goillot E, Merrouche Y, Mercatello A, Ravault A, Tourani JM, Moskovtchenko JF. Serum level of interleukin 6 as a prognosis factor in metastatic renal cell carcinoma. *Cancer Res* 1992; **52**: 3317-3322
- 44 **De Vita F**, Orditura M, Auriemma A, Infusino S, Catalano G. Serum concentrations of proinflammatory cytokines in advanced non small cell lung cancer patients. *J Exp Clin Cancer Res* 1998; **17**: 413-417
- 45 **Knüpfer H**, Preiss R. Significance of interleukin-6 (IL-6) in breast cancer (review). *Breast Cancer Res Treat* 2007; **102**: 129-135
- 46 **Salgado R**, Junius S, Benoy I, Van Dam P, Vermeulen P, Van Marck E, Huget P, Dirix LY. Circulating interleukin-6 predicts survival in patients with metastatic breast cancer. *Int J Cancer* 2003; **103**: 642-646
- 47 **Kuroda K**, Nakashima J, Kanao K, Kikuchi E, Miyajima A, Horiguchi Y, Nakagawa K, Oya M, Ohigashi T, Murai M. Interleukin 6 is associated with cachexia in patients with prostate cancer. *Urology* 2007; **69**: 113-117
- 48 **Shariat SF**, Andrews B, Kattan MW, Kim J, Wheeler TM, Slawin KM. Plasma levels of interleukin-6 and its soluble receptor are associated with prostate cancer progression and metastasis. *Urology* 2001; **58**: 1008-1015
- 49 **Atreya R**, Neurath MF. Signaling molecules: the pathogenic role of the IL-6/STAT-3 trans signaling pathway in intestinal inflammation and in colonic cancer. *Curr Drug Targets* 2008; **9**: 369-374
- 50 **Knüpfer H**, Preiss R. Lack of Knowledge: Breast Cancer and the Soluble Interleukin-6 Receptor. *Breast Care (Basel)* 2010; **5**: 177-180
- 51 **Chalaris A**, Rabe B, Paliga K, Lange H, Laskay T, Fielding CA, Jones SA, Rose-John S, Scheller J. Apoptosis is a natural stimulus of IL6R shedding and contributes to the proinflammatory trans-signaling function of neutrophils. *Blood* 2007; **110**: 1748-1755
- 52 **Jones SA**, Horiuchi S, Topley N, Yamamoto N, Fuller GM. The soluble interleukin 6 receptor: mechanisms of production and implications in disease. *FASEB J* 2001; **15**: 43-58
- 53 **Scheller J**, Ohnesorge N, Rose-John S. Interleukin-6 transsignalling in chronic inflammation and cancer. *Scand J Immunol* 2006; **63**: 321-329
- 54 **Santer FR**, Malinowska K, Culig Z, Cavarretta IT. Interleukin-6 trans-signalling differentially regulates proliferation, migration, adhesion and maspin expression in human prostate cancer cells. *Endocr Relat Cancer* 2010; **17**: 241-253
- 55 **Drachenberg DE**, Elgamal AA, Rowbotham R, Peterson M, Murphy GP. Circulating levels of interleukin-6 in patients with hormone refractory prostate cancer. *Prostate* 1999; **41**: 127-133
- 56 **Wise GJ**, Marella VK, Talluri G, Shirazian D. Cytokine variations in patients with hormone treated prostate cancer. *J Urol* 2000; **164**: 722-725
- 57 **Akimoto S**, Okumura A, Fuse H. Relationship between serum levels of interleukin-6, tumor necrosis factor-alpha and bone turnover markers in prostate cancer patients. *Endocr J* 1998; **45**: 183-189
- 58 **Adler HL**, McCurdy MA, Kattan MW, Timme TL, Scardino PT, Thompson TC. Elevated levels of circulating interleukin-6 and transforming growth factor-beta1 in patients with metastatic prostatic carcinoma. *J Urol* 1999; **161**: 182-187
- 59 **Michalaki V**, Syrigos K, Charles P, Waxman J. Serum levels of IL-6 and TNF-alpha correlate with clinicopathological features and patient survival in patients with prostate cancer. *Br J Cancer* 2004; **90**: 2312-2316
- 60 **George DJ**, Halabi S, Shepard TF, Sanford B, Vogelzang NJ, Small EJ, Kantoff PW. The prognostic significance of plasma interleukin-6 levels in patients with metastatic hormone-refractory prostate cancer: results from cancer and leukemia group B 9480. *Clin Cancer Res* 2005; **11**: 1815-1820
- 61 **Shariat SF**, Kattan MW, Traxel E, Andrews B, Zhu K, Wheeler TM, Slawin KM. Association of pre- and postoperative plasma levels of transforming growth factor beta(1) and interleukin 6 and its soluble receptor with prostate cancer progression. *Clin Cancer Res* 2004; **10**: 1992-1999
- 62 **Shariat SF**, Karam JA, Walz J, Roehrborn CG, Montorsi F, Margulis V, Saad F, Slawin KM, Karakiewicz PI. Improved prediction of disease relapse after radical prostatectomy through a panel of preoperative blood-based biomarkers. *Clin Cancer Res* 2008; **14**: 3785-3791
- 63 **Shariat SF**, Walz J, Roehrborn CG, Zlotta AR, Perrotte P, Suardi N, Saad F, Karakiewicz PI. External validation of a biomarker-based preoperative nomogram predicts biochemical recurrence after radical prostatectomy. *J Clin Oncol* 2008; **26**: 1526-1531
- 64 **Alcover J**, Filella X, Luqué P, Molina R, Izquierdo L, Augé JM, Alcaraz A. Prognostic value of IL-6 in localized prostatic cancer. *Anticancer Res* 2010; **30**: 4369-4372
- 65 **Svatek RS**, Jeldres C, Karakiewicz PI, Suardi N, Walz J, Roehrborn CG, Montorsi F, Slawin KM, Shariat SF. Pre-treatment biomarker levels improve the accuracy of post-prostatectomy nomogram for prediction of biochemical recurrence. *Prostate* 2009; **69**: 886-894
- 66 **Kattan MW**, Shariat SF, Andrews B, Zhu K, Canto E, Matsumoto K, Muramoto M, Scardino PT, Ohori M, Wheeler TM, Slawin KM. The addition of interleukin-6 soluble receptor and transforming growth factor beta1 improves a preoperative nomogram for predicting biochemical progression in patients with clinically localized prostate cancer. *J Clin Oncol* 2003; **21**: 3573-3579
- 67 **Finley DS**, Calvert VS, Inokuchi J, Lau A, Narula N, Petricoin EF, Zaldivar F, Santos R, Tyson DR, Ornstein DK. Periprostatic adipose tissue as a modulator of prostate cancer aggressiveness. *J Urol* 2009; **182**: 1621-1627
- 68 **Rötter V**, Nagaev I, Smith U. Interleukin-6 (IL-6) induces insulin resistance in 3T3-L1 adipocytes and is, like IL-8 and tumor necrosis factor-alpha, overexpressed in human fat cells from insulin-resistant subjects. *J Biol Chem* 2003; **278**: 45777-45784
- 69 **Sadagurski M**, Norquay L, Farhang J, D'Aquino K, Copps K, White MF. Human IL6 enhances leptin action in mice. *Diabetologia* 2010; **53**: 525-535
- 70 **Akira S**. IL-6-regulated transcription factors. *Int J Biochem Cell Biol* 1997; **29**: 1401-1418
- 71 **Mistry T**, Digby JE, Desai KM, Randeva HS. Obesity and prostate cancer: a role for adipokines. *Eur Urol* 2007; **52**: 46-53
- 72 **Ahima RS**, Flier JS. Adipose tissue as an endocrine organ. *Trends Endocrinol Metab* 2000; **11**: 327-332
- 73 **Frühbeck G**, Gómez-Ambrosi J, Muruzábal FJ, Burrell MA. The adipocyte: a model for integration of endocrine and metabolic signaling in energy metabolism regulation. *Am J Physiol Endocrinol Metab* 2001; **280**: E827-E847
- 74 **Onuma M**, Bub JD, Rummel TL, Iwamoto Y. Prostate cancer cell-adipocyte interaction: leptin mediates androgen-independent prostate cancer cell proliferation through c-Jun NH2-terminal kinase. *J Biol Chem* 2003; **278**: 42660-42667
- 75 **Shariat SF**, Chromecki TF, Hoefler J, Barbieri CE, Scherr DS, Karakiewicz PI, Roehrborn CG, Montorsi F, Culig Z, Cavarretta IT. Soluble gp130 Regulates Prostate Cancer Invasion and Progression in an Interleukin-6 Dependent and Independent Manner. *J Urol* 2011; **186**: 2107-2114
- 76 **Malinowska K**, Neuwirt H, Cavarretta IT, Bektic J, Steiner H,

- Dietrich H, Moser PL, Fuchs D, Hobisch A, Culig Z. Interleukin-6 stimulation of growth of prostate cancer in vitro and in vivo through activation of the androgen receptor. *Endocr Relat Cancer* 2009; **16**: 155-169
- 77 **Royuela M**, Ricote M, Parsons MS, García-Tuñón I, Paniagua R, de Miguel MP. Immunohistochemical analysis of the IL-6 family of cytokines and their receptors in benign, hyperplastic, and malignant human prostate. *J Pathol* 2004; **202**: 41-49
- 78 **Degeorges A**, Tatoud R, Fauvel-Lafeve F, Podgorniak MP, Millot G, de Cremoux P, Calvo F. Stromal cells from human benign prostate hyperplasia produce a growth-inhibitory factor for LNCaP prostate cancer cells, identified as interleukin-6. *Int J Cancer* 1996; **68**: 207-214
- 79 **Giri D**, Ozen M, Ittmann M. Interleukin-6 is an autocrine growth factor in human prostate cancer. *Am J Pathol* 2001; **159**: 2159-2165
- 80 **Palmer J**, Hertzog PJ, Hammacher A. Differential expression and effects of gp130 cytokines and receptors in prostate cancer cells. *Int J Biochem Cell Biol* 2004; **36**: 2258-2269
- 81 **Hobisch A**, Ramoner R, Fuchs D, Godoy-Tundidor S, Bartsch G, Klocker H, Culig Z. Prostate cancer cells (LNCaP) generated after long-term interleukin 6 (IL-6) treatment express IL-6 and acquire an IL-6 partially resistant phenotype. *Clin Cancer Res* 2001; **7**: 2941-2948
- 82 **Okamoto M**, Lee C, Oyasu R. Interleukin-6 as a paracrine and autocrine growth factor in human prostatic carcinoma cells in vitro. *Cancer Res* 1997; **57**: 141-146
- 83 **Lou W**, Ni Z, Dyer K, Tweardy DJ, Gao AC. Interleukin-6 induces prostate cancer cell growth accompanied by activation of stat3 signaling pathway. *Prostate* 2000; **42**: 239-242
- 84 **Borsellino N**, Belledgrun A, Bonavida B. Endogenous interleukin 6 is a resistance factor for cis-diamminedichloroplatinum and etoposide-mediated cytotoxicity of human prostate carcinoma cell lines. *Cancer Res* 1995; **55**: 4633-4639
- 85 **Chung TD**, Yu JJ, Spiotto MT, Bartkowski M, Simons JW. Characterization of the role of IL-6 in the progression of prostate cancer. *Prostate* 1999; **38**: 199-207
- 86 **Qiu Y**, Robinson D, Pretlow TG, Kung HJ. Etk/Bmx, a tyrosine kinase with a pleckstrin-homology domain, is an effector of phosphatidylinositol 3'-kinase and is involved in interleukin 6-induced neuroendocrine differentiation of prostate cancer cells. *Proc Natl Acad Sci U S A* 1998; **95**: 3644-3649
- 87 **Mori S**, Murakami-Mori K, Bonavida B. Dexamethasone enhances expression of membrane and soluble interleukin-6 receptors by prostate carcinoma cell lines. *Anticancer Res* 1998; **18**: 4403-4408
- 88 **Culig Z**, Bartsch G, Hobisch A. Interleukin-6 regulates androgen receptor activity and prostate cancer cell growth. *Mol Cell Endocrinol* 2002; **197**: 231-238
- 89 **Böhm M**, Schulte U, Funk JO, Raghunath M, Behrmann I, Kortylewski M, Heinrich PC, Kues T, Luger TA, Schwarz T. Interleukin-6-resistant melanoma cells exhibit reduced activation of STAT3 and lack of inhibition of cyclin E-associated kinase activity. *J Invest Dermatol* 2001; **117**: 132-140
- 90 **Pronzato P**, Rondini M. Hormonotherapy of advanced prostate cancer. *Ann Oncol* 2005; **16** Suppl 4: iv80-iv84
- 91 **Feldman BJ**, Feldman D. The development of androgen-independent prostate cancer. *Nat Rev Cancer* 2001; **1**: 34-45
- 92 **Rosenberg J**, Small EJ. Prostate cancer update. *Curr Opin Oncol* 2003; **15**: 217-221
- 93 **Culig Z**. Role of the androgen receptor axis in prostate cancer. *Urology* 2003; **62**: 21-26
- 94 **Terakawa T**, Miyake H, Furukawa J, Ettinger SL, Gleave ME, Fujisawa M. Enhanced sensitivity to androgen withdrawal due to overexpression of interleukin-6 in androgen-dependent human prostate cancer LNCaP cells. *Br J Cancer* 2009; **101**: 1731-1739
- 95 **Robson M**, Dawson N. How is androgen-dependent metastatic prostate cancer best treated? *Hematol Oncol Clin North Am* 1996; **10**: 727-747
- 96 **Thompson IM**, Goodman PJ, Tangen CM, Lucia MS, Miller GJ, Ford LG, Lieber MM, Cespedes RD, Atkins JN, Lippman SM, Carlin SM, Ryan A, Szczepanek CM, Crowley JJ, Coltman CA. The influence of finasteride on the development of prostate cancer. *N Engl J Med* 2003; **349**: 215-224
- 97 **Chen CD**, Welsbie DS, Tran C, Baek SH, Chen R, Vessella R, Rosenfeld MG, Sawyers CL. Molecular determinants of resistance to antiandrogen therapy. *Nat Med* 2004; **10**: 33-39
- 98 **Zegarra-Moro OL**, Schmidt LJ, Huang H, Tindall DJ. Disruption of androgen receptor function inhibits proliferation of androgen-refractory prostate cancer cells. *Cancer Res* 2002; **62**: 1008-1013
- 99 **Suzuki H**, Ueda T, Ichikawa T, Ito H. Androgen receptor involvement in the progression of prostate cancer. *Endocr Relat Cancer* 2003; **10**: 209-216
- 100 **Sadi MV**, Walsh PC, Barrack ER. Immunohistochemical study of androgen receptors in metastatic prostate cancer. Comparison of receptor content and response to hormonal therapy. *Cancer* 1991; **67**: 3057-3064
- 101 **Tilley WD**, Lim-Tio SS, Horsfall DJ, Aspinnall JO, Marshall VR, Skinner JM. Detection of discrete androgen receptor epitopes in prostate cancer by immunostaining: measurement by color video image analysis. *Cancer Res* 1994; **54**: 4096-4102
- 102 **Trapman J**, Cleutjens KB. Androgen-regulated gene expression in prostate cancer. *Semin Cancer Biol* 1997; **8**: 29-36
- 103 **Pienta KJ**, Bradley D. Mechanisms underlying the development of androgen-independent prostate cancer. *Clin Cancer Res* 2006; **12**: 1665-1671
- 104 **Shah RB**, Mehra R, Chinnaiyan AM, Shen R, Ghosh D, Zhou M, Macvicar GR, Varambally S, Harwood J, Bismar TA, Kim R, Rubin MA, Pienta KJ. Androgen-independent prostate cancer is a heterogeneous group of diseases: lessons from a rapid autopsy program. *Cancer Res* 2004; **64**: 9209-9216
- 105 **Nelson WG**, De Marzo AM, Isaacs WB. Prostate cancer. *N Engl J Med* 2003; **349**: 366-381
- 106 **Debes JD**, Tindall DJ. Mechanisms of androgen-refractory prostate cancer. *N Engl J Med* 2004; **351**: 1488-1490
- 107 **Tindall D**, Horne FM, Hruszkewycz A, Mohla S, Shuman M, Wang Z, Kantoff P. Symposium on androgen action in prostate cancer. *Cancer Res* 2004; **64**: 7178-7180
- 108 **Gregory CW**, Johnson RT, Mohler JL, French FS, Wilson EM. Androgen receptor stabilization in recurrent prostate cancer is associated with hypersensitivity to low androgen. *Cancer Res* 2001; **61**: 2892-2898
- 109 **Culig Z**, Steiner H, Bartsch G, Hobisch A. Mechanisms of endocrine therapy-responsive and -unresponsive prostate tumours. *Endocr Relat Cancer* 2005; **12**: 229-244
- 110 **Jänne OA**, Moilanen AM, Poukka H, Rouleau N, Karvonen U, Kotaja N, Häkli M, Palvimo JJ. Androgen-receptor-interacting nuclear proteins. *Biochem Soc Trans* 2000; **28**: 401-405
- 111 **Colombel M**, Symmans F, Gil S, O'Toole KM, Chopin D, Benson M, Olsson CA, Korsmeyer S, Buttyan R. Detection of the apoptosis-suppressing oncoprotein bc1-2 in hormone-refractory human prostate cancers. *Am J Pathol* 1993; **143**: 390-400
- 112 **Collins AT**, Berry PA, Hyde C, Stower MJ, Maitland NJ. Prospective identification of tumorigenic prostate cancer stem cells. *Cancer Res* 2005; **65**: 10946-10951
- 113 **Culig Z**. Androgen receptor cross-talk with cell signalling pathways. *Growth Factors* 2004; **22**: 179-184
- 114 **Culig Z**, Hobisch A, Cronauer MV, Radmayr C, Trapman J, Hittmair A, Bartsch G, Klocker H. Androgen receptor activation in prostatic tumor cell lines by insulin-like growth factor-I, keratinocyte growth factor, and epidermal growth factor. *Cancer Res* 1994; **54**: 5474-5478
- 115 **Wang G**, Wang J, Sadar MD. Crosstalk between the androgen receptor and beta-catenin in castrate-resistant prostate cancer. *Cancer Res* 2008; **68**: 9918-9927
- 116 **Reebye V**, Frilling A, Habib NA, Mintz PJ. Intracellular

- adaptor molecules and AR signalling in the tumour micro-environment. *Cell Signal* 2011; **23**: 1017-1021
- 117 **Lin DL**, Whitney MC, Yao Z, Keller ET. Interleukin-6 induces androgen responsiveness in prostate cancer cells through up-regulation of androgen receptor expression. *Clin Cancer Res* 2001; **7**: 1773-1781
- 118 **Chen T**, Wang LH, Farrar WL. Interleukin 6 activates androgen receptor-mediated gene expression through a signal transducer and activator of transcription 3-dependent pathway in LNCaP prostate cancer cells. *Cancer Res* 2000; **60**: 2132-2135
- 119 **Hobisch A**, Eder IE, Putz T, Horninger W, Bartsch G, Klocker H, Culig Z. Interleukin-6 regulates prostate-specific protein expression in prostate carcinoma cells by activation of the androgen receptor. *Cancer Res* 1998; **58**: 4640-4645
- 120 **Teixeira AL**, Ribeiro R, Morais A, Lobo F, Fraga A, Pina F, Calais-da-Silva FM, Calais-da-Silva FE, Medeiros R. Combined analysis of EGF+61G& gt; A and TGFBI+869T& gt; C functional polymorphisms in the time to androgen independence and prostate cancer susceptibility. *Pharmacogenomics J* 2009; **9**: 341-346
- 121 **Paule B**, Terry S, Kheuang L, Soyeux P, Vacherot F, de la Taille A. The NF-kappaB/IL-6 pathway in metastatic androgen-independent prostate cancer: new therapeutic approaches? *World J Urol* 2007; **25**: 477-489
- 122 **Culig Z**. Cytokine disbalance in common human cancers. *Biochim Biophys Acta* 2011; **1813**: 308-314
- 123 **Zelivianski S**, Verni M, Moore C, Kondrikov D, Taylor R, Lin MF. Multipathways for transdifferentiation of human prostate cancer cells into neuroendocrine-like phenotype. *Biochim Biophys Acta* 2001; **1539**: 28-43
- 124 **Becker C**, Fantini MC, Schramm C, Lehr HA, Wirtz S, Nikolaev A, Burg J, Strand S, Kiesslich R, Huber S, Ito H, Nishimoto N, Yoshizaki K, Kishimoto T, Galle PR, Blessing M, Rose-John S, Neurath MF. TGF-beta suppresses tumor progression in colon cancer by inhibition of IL-6 trans-signaling. *Immunity* 2004; **21**: 491-501
- 125 **Kang KW**, Wagley Y, Kim HW, Pokharel YR, Chung YY, Chang IY, Kim JJ, Moon JS, Kim YK, Nah SY, Kang HS, Oh JW. Novel role of IL-6/SIL-6R signaling in the expression of inducible nitric oxide synthase (iNOS) in murine B16, metastatic melanoma clone F10.9, cells. *Free Radic Biol Med* 2007; **42**: 215-227
- 126 **Wagley Y**, Yoo YC, Seo HG, Rhee MH, Kim TH, Kang KW, Nah SY, Oh JW. The IL-6/sIL-6R treatment of a malignant melanoma cell line enhances susceptibility to TNF-alpha-induced apoptosis. *Biochem Biophys Res Commun* 2007; **354**: 985-991
- 127 **Nowell MA**, Williams AS, Carty SA, Scheller J, Hayes AJ, Jones GW, Richards PJ, Slinn S, Ernst M, Jenkins BJ, Topley N, Rose-John S, Jones SA. Therapeutic targeting of IL-6 trans signaling counteracts STAT3 control of experimental inflammatory arthritis. *J Immunol* 2009; **182**: 613-622
- 128 **Economides AN**, Carpenter LR, Rudge JS, Wong V, Koehler-Stec EM, Hartnett C, Pyles EA, Xu X, Daly TJ, Young MR, Fandl JP, Lee F, Carver S, McNay J, Bailey K, Ramakanth S, Hutabarat R, Huang TT, Radziejewski C, Yancopoulos GD, Stahl N. Cytokine traps: multi-component, high-affinity blockers of cytokine action. *Nat Med* 2003; **9**: 47-52
- 129 **Ni Z**, Lou W, Leman ES, Gao AC. Inhibition of constitutively activated Stat3 signaling pathway suppresses growth of prostate cancer cells. *Cancer Res* 2000; **60**: 1225-1228
- 130 **Xi S**, Gooding WE, Grandis JR. In vivo antitumor efficacy of STAT3 blockade using a transcription factor decoy approach: implications for cancer therapy. *Oncogene* 2005; **24**: 970-979
- 131 **Steiner H**, Cavarretta IT, Moser PL, Berger AP, Bektic J, Dietrich H, Zaki MH, Nakada M, Hobisch A, Nemeth JA, Culig Z. Regulation of growth of prostate cancer cells selected in the presence of interleukin-6 by the anti-interleukin-6 antibody CNTO 328. *Prostate* 2006; **66**: 1744-1752
- 132 **Smith PC**, Keller ET. Anti-interleukin-6 monoclonal antibody induces regression of human prostate cancer xenografts in nude mice. *Prostate* 2001; **48**: 47-53
- 133 **Wallner L**, Dai J, Escara-Wilke J, Zhang J, Yao Z, Lu Y, Trikha M, Nemeth JA, Zaki MH, Keller ET. Inhibition of interleukin-6 with CNTO328, an anti-interleukin-6 monoclonal antibody, inhibits conversion of androgen-dependent prostate cancer to an androgen-independent phenotype in orchiectomized mice. *Cancer Res* 2006; **66**: 3087-3095
- 134 **Zaki MH**, Nemeth JA, Trikha M. CNTO 328, a monoclonal antibody to IL-6, inhibits human tumor-induced cachexia in nude mice. *Int J Cancer* 2004; **111**: 592-595
- 135 **Dorff TB**, Goldman B, Pinski JK, Mack PC, Lara PN, Van Veldhuizen PJ, Quinn DI, Vogelzang NJ, Thompson IM, Hussain MH. Clinical and correlative results of SWOG S0354: a phase II trial of CNTO328 (siltuximab), a monoclonal antibody against interleukin-6, in chemotherapy-pretreated patients with castration-resistant prostate cancer. *Clin Cancer Res* 2010; **16**: 3028-3034
- 136 **Patterson SG**, Wei S, Chen X, Sallman DA, Gilvary DL, Zhong B, Pow-Sang J, Yeatman T, Djeu JY. Novel role of Stat1 in the development of docetaxel resistance in prostate tumor cells. *Oncogene* 2006; **25**: 6113-6122
- 137 **Karkera J**, Steiner H, Li W, Skradski V, Moser PL, Riethdorf S, Reddy M, Puchalski T, Safer K, Prabhakar U, Pantel K, Qi M, Culig Z. The anti-interleukin-6 antibody siltuximab down-regulates genes implicated in tumorigenesis in prostate cancer patients from a phase I study. *Prostate* 2011; **71**: 1455-1465

S- Editor Yang XC L- Editor Webster JR E- Editor Yang XC

Clinical utility of cerebrovascular reactivity mapping in patients with low grade gliomas

Jay J Pillai, Domenico Zacá

Jay J Pillai, Domenico Zacá, Neuroradiology Division, Russell H Morgan Department of Radiology and Radiological Science, The Johns Hopkins University, School of Medicine and The Johns Hopkins Hospital, 600 N. Wolfe Street, Phipps B-100, Baltimore, MD 21287, United States

Author contributions: Pillai JJ and Zacá D contributed equally to this work; Pillai JJ and Zacá D performed the research; Pillai JJ and Zacá D wrote the paper.

Supported by A grant from Siemens Medical Solutions, Inc., which only partially funded this work without any involvement in the actual research

Correspondence to: Jay J Pillai, MD, Neuroradiology Division, Russell H Morgan Department of Radiology and Radiological Science, The Johns Hopkins University, School of Medicine and The Johns Hopkins Hospital, 600 N. Wolfe Street, Phipps B-100, Baltimore, MD 21287, United States. jpillai1@jhmi.edu

Telephone: +410-955-2353 Fax: +410-614-1213
Received: September 8, 2011 Revised: November 24, 2011

Accepted: December 1, 2011

Published online: December 10, 2011

Abstract

AIM: To evaluate neurovascular uncoupling (NVU) associated with low grade gliomas (LGG) using blood oxygen level dependent (BOLD) cerebrovascular reactivity mapping.

METHODS: Seven patients with low grade gliomas referred by neurosurgeons for presurgical mapping were included in this pilot study. Cerebrovascular reactivity (CVR) mapping was performed by acquiring BOLD images while patients performed a block-design breath-hold (BH) hypercapnia task. CVR mapping was expressed as BOLD percentage signal change (PSC) from baseline associated with performance of the BH hypercapnia task. Standard T2* Dynamic Susceptibility Contrast perfusion imaging was performed and relative cerebral blood volume (rCBV) and relative cerebral blood flow (rCBF) maps were generated. Structural T1 weighted MR images were also acquired. A correlation

analysis between intratumoral normalized (*via* ratio with contralateral homologous regions) BOLD BH PSC [referred to as (n_{CVR})] and intratumoral normalized resting state rCBV (rCBF) values (i.e., n_{CBV} and n_{CBF} , respectively) was performed.

RESULTS: No significant correlation was seen between the normalized BOLD BH PSC (i.e., n_{CVR}) and n_{CBV} or n_{CBF} . However, the average n_{CVR} (median = 0.50, $z = -2.28$, $P = 0.01$) was significantly less than 1.0, indicating abnormally reduced vascular responses in the tumor regions relative to normal contralesional homologous regions, whereas the average n_{CBV} (median = 0.94, $z = -0.92$, $P = 0.375$) and n_{CBF} (median = 0.93, $z = -1.16$, $P = 0.25$) were not significantly higher or lower than 1.0, indicating iso-perfusion in the tumor regions relative to normal contralesional homologous regions. These findings suggest that in LGG, hyperperfusion that is seen in high grade gliomas is not present, but, nevertheless, abnormally decreased regional CVR is present within and adjacent to LGG. Since the patients all demonstrated at least some residual function attributable to the cortical regions of impaired CVR, but were incapable of producing a BOLD response in these regions regardless of the tasks performed, such regionally decreased CVR is indicative of NVU. The low n_{CVR} ratios indicate high prevalence of NVU in this LGG cohort, which is an important consideration in the interpretation of clinical presurgical mapping with functional magnetic resonance (MR) imaging.

CONCLUSION: Our preliminary study shows that BH CVR mapping is clinically feasible and demonstrates an unexpectedly high prevalence of NVU in patients with LGG.

© 2011 Baishideng. All rights reserved.

Key words: Blood oxygen level dependent; Brain tumor; Cerebrovascular reactivity; Functional MRI; Neurovascular uncoupling; Presurgical mapping

Peer reviewer: Ali Syed Arbab, MD, PhD, Associate Scientist and Director, Cellular and Molecular Imaging Laboratory, Department of Radiology, Henry Ford Hospital, 1 Ford Place, 2F, PO Box 82, Detroit, MI 48202, United States

Pillai JJ, Zacá D. Clinical utility of cerebrovascular reactivity mapping in patients with low grade gliomas. *World J Clin Oncol* 2011; 2(12): 397-403 Available from: URL: <http://www.wjgnet.com/2218-4333/full/v2/i12/397.htm> DOI: <http://dx.doi.org/10.5306/wjco.v2.i12.397>

INTRODUCTION

Presurgical localization of brain sensorimotor, visual and language regions in patients with brain tumors or epilepsy who are candidates for surgical resection currently represents the most mature clinical application of blood oxygen level dependent (BOLD) functional MR imaging (fMRI)^[1-3]. Presurgical mapping with fMRI can assist neurosurgeons by providing useful information for: (1) preoperative risk assessment; (2) planning the safest surgical trajectory; (3) selection of patients for asleep *vs* awake craniotomy; and most importantly; and (4) optimization of efficiency, exposure and technique of intraoperative mapping. However, despite its many advantages in presurgical planning that have resulted in widespread clinical utilization over the last decade, some limitations of clinical fMRI do exist^[4-9]. One such limitation is the frequent inability to distinguish essential from nonessential participatory activated (i.e., “eloquent”) cortex involved in performance of a particular cognitive, sensorimotor or visual task, thus leading to lower than ideal specificity of activation maps^[10].

Another limitation is the problem of decreased sensitivity for detection of actual electrically active eloquent cortex in areas of impaired cerebrovascular reactivity (CVR); this phenomenon is referred to as neurovascular uncoupling (NVU)^[11,12]. NVU can result in false negative BOLD activation, which constitutes a major hazard with respect to interpretation of clinical fMRI examinations; such false negatives within or adjacent to tumor boundaries can result in undesirable resection (in the absence of intraoperative electrophysiologic confirmation) of essential electrically active eloquent cortex that is incapable of demonstrating a BOLD response due to impaired CVR. Such eloquent cortical resection may lead to serious permanent postoperative neurological deficits. Thus, the phenomenon of NVU is not merely a theoretical issue of scientific interest, but is rather an issue of considerable clinical relevance and importance. NVU has been documented in the immediate vicinity of high grade gliomas (HGG), mostly due to tumor angiogenesis, which is associated with abnormal vascular permeability, abnormal hyperperfusion (elevated relative cerebral blood volume [rCBV]) related to increased vascular density, and impaired regional CVR^[13,14]. However, it is not clear how high the prevalence of impaired CVR (and resultant NVU) is in low grade gliomas (LGG), in which

Table 1 Age, sex, tumor location and histology for seven brain tumor cases included in the study

Age	Sex	Tumor location	Histology/tumor grade
25	F	Left frontal lobe	Oligoastrocytoma grade II
27	M	Right cingulate gyrus	Oligodendroglioma grade II
42	M	Left temporal lobe	Astrocytoma grade II
25	M	Right temporal lobe	Diffuse astrocytoma grade II
54	M	Left frontal	Oligodendroglioma grade II
27	F	Left hemispheric (primarily insular and inferior frontal)	Pilocytic astrocytoma grade I
41	M	Left insular	Oligoastrocytoma grade II

F: Female; M: Male.

hyperperfusion is unusual. In this study, we investigated regional CVR, using a BOLD breath-hold (BH) hypercapnia task, within LGG, which are known to infiltrate, rather than destroy or displace, eloquent cortex, in order to determine whether the same NVU potential exists in these tumors as in HGG. In this study we report our initial experience using BH CVR mapping in 7 patients with LGG (6 patients with grade II gliomas and 1 patient with grade I glioma) as a quality control tool for detecting risk of NVU, and we compare these results to those of T2* DSC perfusion imaging that was also performed during the same scan sessions as part of a comprehensive clinical presurgical mapping protocol. The findings of this study are discussed in the context of current literature pertaining to brain tumor-related NVU.

MATERIALS AND METHODS

Seven patients (mean age 34 ± 11 year, 5M/2F) with histopathologically proved grade I and II intra-axial primary brain tumors (Table 1) underwent our institutional clinical BOLD fMRI protocol for presurgical planning which included multiple T2* BOLD fMRI sequences during performance of motor, language or visual tasks and a BH task, a T2* Dynamic Susceptibility Contrast perfusion sequence and a structural T1-weighted 3D MPRAGE sequence after Gadolinium injection. Details of these three sequences are reported in Table 2. Images were acquired on a 3T MRI scanner (Magnetom Trio, Siemens Medical Solutions, Erlangen, Germany). The block design BH task consisted of four cycles of 40 s each of normal breathing (baseline) alternating with blocks of 4 s of inhalation and 16 s of breath-holding^[15]. Instructions for task performance were delivered visually using Prism Acquire Software (Prism Clinical Imaging, Elm Grove, WI, United States).

The study was approved by our Institutional Review Board. Images for each patient were first transferred to an external workstation. Perfusion and raw BOLD BH images were coregistered to the T1 MPRAGE images using DynaSuiteNeuro software (DynaSuiteNeuro, InVivo Corporation, Pewaukee, WI, United States). Perfusion image analysis included the generation of rCBV and rCBF maps. rCBV was calculated by adding a correction

Table 2 Main parameters for the sequences of interest in the study

Sequence	TR (ms)	TE (ms)	FA	FOV (cm ²)	Acquisition matrix	Slice thickness (mm)
T1 MPRAGE	7	3.5	9°	24 × 24	256 × 256	1
T2* DSC	2450	45	90°	24 × 24	128 × 128	4
T2* BOLD	2000	30	90°	24 × 24	64 × 64	4

DSC: Dynamic susceptibility contrast; BOLD: blood oxygen level dependent; FOV: Field of view.

factor to take into account the contrast leakage through the disrupted blood-brain barrier^[16]. BOLD BH data analysis was carried out using AFNI software (afni.nimh.nih.gov) and included slice timing correction, realignment, spatial smoothing followed by generation of PSC maps^[17]. Subsequently, a Region of Interest (ROI) analysis was carried out using MIPAV (mipav.cit.nih.gov) (Medical Image Processing, Analysis and Visualization) software. For each patient, two independent raters [JJP (a board-certified neuroradiologist with 14 years of neuroimaging experience) and DZ (an imaging scientist with a PhD in functional imaging and 3 years of postdoctoral neuroimaging experience)] selected on the high resolution T1 MPRAGE images a ROI that included the tumor entirely, defined as the entire hypointense component. This ROI encompassing the entire lesion shall be referred to as the “ipsilesional ROI.” This T1 hypointense region corresponded exactly to the areas of tumor T2/FLAIR hyperintensity seen on other sequences acquired as part of the overall clinical fMRI examination, but T1 MPRAGE images were selected for ROI delineation because of their higher resolution compared to the standard FSE T2 and T2 FLAIR sequences. A mirror homologous contralateral hemispheric (referred to as “contralesional”) ROI was generated in a semi-automated fashion, with particular attention paid to trying (to the greatest extent possible) to ensure a similar degree of contribution from gray and white matter structures in the contralesional ROI as in the ipsilesional ROI, considering the degree of anatomic distortion resulting from the tumor. The following metrics were then calculated: a normalized rCBV (n_{CBV}), expressed as the ratio between the mean rCBV value (of all included voxels) in the ipsilesional ROI and the mean rCBV value in the contralesional ROI; a similarly computed normalized rCBF (n_{CBF}), defined as the ratio between the mean rCBF value in the ipsilesional ROI and the mean rCBF value in the contralesional ROI; a normalized PSC (n_{CVR}) expressed as the ratio between the mean BOLD PSC in the ipsilesional ROI and the mean BOLD PSC in the contralesional ROI.

Correlation analysis was performed between n_{CBV} and n_{CVR} as well as between n_{CBF} and n_{CVR} . Mean values among the raters were used. A one sample Wilcoxon test was also performed to assess whether the PSC normalized ratio was significantly lower than 1.0. An identical statistical test was also performed on n_{CBV} and n_{CBF} to determine whether there were any significant differences in perfusion metrics

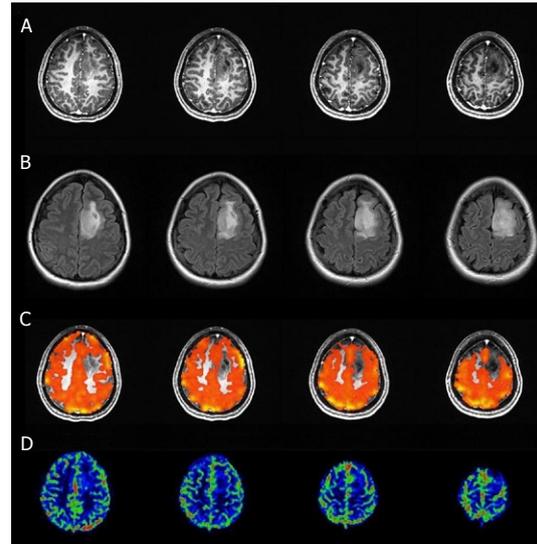


Figure 1 Axial T1 postcontrast 3D MPRAGE and T2 FLAIR images (A and B) breath-hold cerebrovascular reactivity maps (C) and relative cerebral blood flow maps (D) in a patient presenting with a left frontal lobe lesion, classified as grade II oligoastrocytoma after surgical resection, are displayed. Breath-hold cerebrovascular reactivity (BH CVR) maps were fused with axial T1 post Gadolinium images, and the threshold was set to 0.35 for blood oxygen level dependent PSC. Decreased CVR is present within and at the anterosuperolateral margin of the lesion (involving the infiltrated cortex of the left superior frontal gyrus) relative to contralateral hemispheric normal tissue. This represents an area of tumor-induced neurovascular uncoupling. Perfusion imaging did not provide equivalent information because the same area does not demonstrate any definite regional perfusion abnormality.

between the ipsilesional and contralesional ROIs. Statistical analysis was performed using OriginPro 8.0 software.

RESULTS

T1 MPRAGE and T2 FLAIR images, perfusion maps and BH PSC maps for two cases are shown in Figures 1 and 2. Reduced PSC is clearly visible in the ipsilesional ROI compared to the contralesional ROI, whereas in both cases the lesion appears iso-perfused relative to the contralesional ROIs. Intraclass Correlation Coefficients (ICC) among the raters were excellent (0.88 for n_{CVR} , 0.98 for n_{CBV} , 0.98 for n_{CBF}). In this group of 7 patients, none of the cases demonstrated a n_{CVR} that was greater than 1.0 (Figure 3A). The fact that all 7 cases demonstrated n_{CVR} values less than 1.0 indicated that every one of the LGG cases demonstrated abnormally decreased CVR in the ipsilesional ROI compared to the contralesional ROI, suggesting a high risk of NVU in all cases in areas of cortex where regionally decreased CVR were noted. Correlation with the patients’ clinical status confirmed that these areas of regionally decreased cortical CVR corresponded to areas of actual NVU, since preservation of residual motor and language function was noted clinically despite absent expected task-based activation in these regions of decreased CVR on task-based fMRI activation maps obtained as part of the concurrent clinical fMRI examinations. Based on group

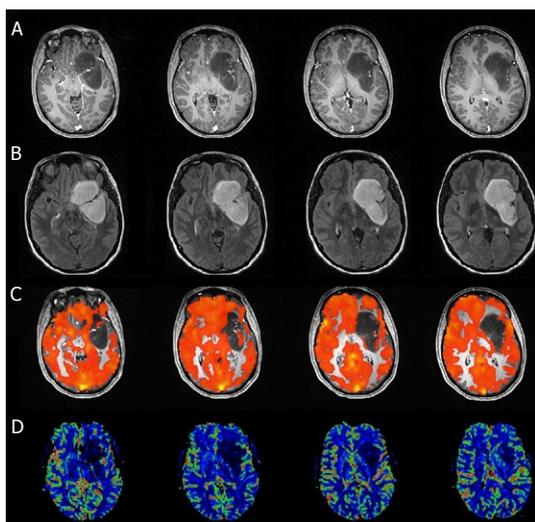


Figure 2 Axial T1 postcontrast 3D MPRAGE and T2 FLAIR images (A and B), breath-hold cerebrovascular reactivity maps (C) and relative cerebral blood flow maps (D) in a patient presenting with a left hemispheric lesion, classified as pilocytic grade I astrocytoma after surgical resection, are displayed. Breath-hold cerebrovascular reactivity (BH CVR) maps were fused with axial T1 post Gadolinium images, and the threshold was set to 0.45 for blood oxygen level dependent (BOLD) signal. Decreased CVR is present within the lateral margin of the lesion (involving the adjacent cortex of the left superior temporal gyrus) relative to contralateral hemispheric normal tissue. This represents an area of tumor-induced neurovascular uncoupling. Perfusion imaging did not provide equivalent information because the same area does not demonstrate regional perfusion abnormality.

analysis, the overall distribution of n_{CVR} in this cohort of patients was statistically significantly lower than 1.0 (median = 0.50, $z = -2.28$, $P = 0.01$). n_{CBV} (median = 0.94, $z = -0.92$, $P = 0.375$) and n_{CBF} (median = 0.93, $z = -1.16$, $P = 0.25$) were not significantly higher or lower than 1.0 at a group level, indicating the absence of any substantial hyperperfusion or hypoperfusion in the ipsilesional ROI compared to the contralesional ROI (Figure 3B and C). We did not find any significant correlation between the perfusion and CVR metrics (Figure 4A and B), indicating that perfusion imaging by itself is not a valid predictor of vascular reactivity, and therefore an indicator of NVU in this particular cohort of LGG patients.

The use of a semi-automated approach to contralesional ROI placement helped to ensure that similar contributions to the contralesional ROI from gray and white matter structures were obtained as in the ipsilesional ROIs. This avoided spuriously high contralateral perfusion and CVR values related to greater contributions from normal gray matter in the contralesional ROIs. This was especially important considering that all of the lesions were gliomas, and as such involved mostly white matter rather than cortical gray matter. None of the 7 LGGs demonstrated any appreciable contrast enhancement on Gadolinium-enhanced T1 weighted images, and none demonstrated any internal areas of necrosis by histopathology or imaging features, although some very small regions of internal cystic change were noted in some of the oligodendrogliomas in this group. It is im-

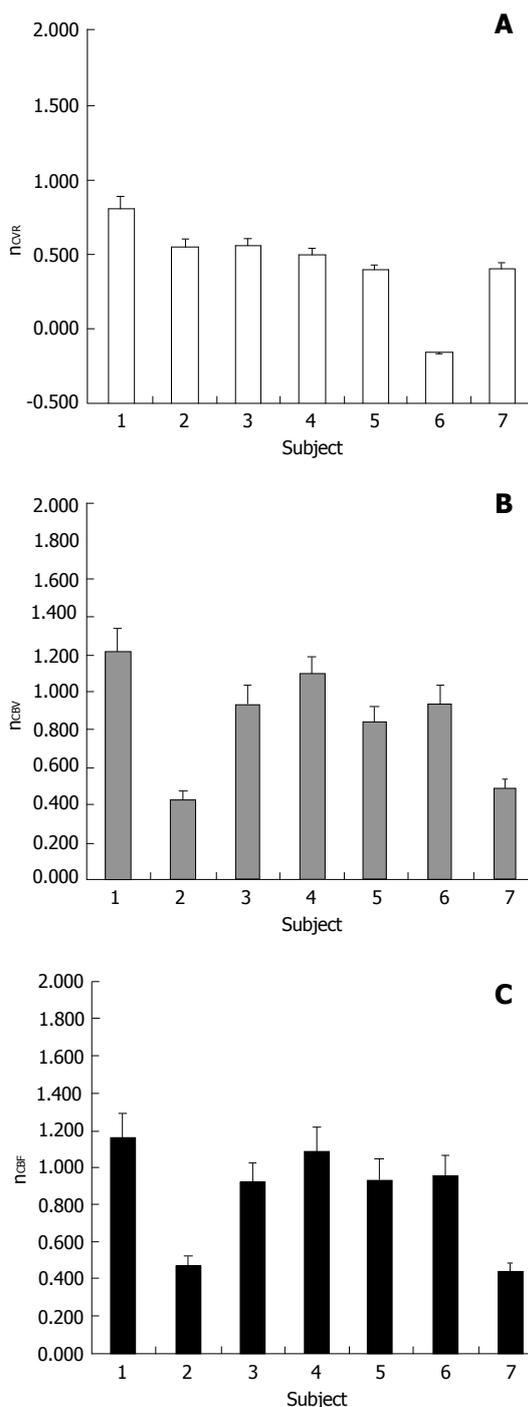


Figure 3 Cerebrovascular reactivity ratio (n_{CVR}) (A) cerebral blood volume ratio (n_{CBV}) (B) and cerebral blood flow ratio (n_{CBF}) (C) distribution for the group of seven patients with classified grade I and grade II tumors included in this study. In all cases n_{CVR} is less than 1.0 and the median value is significantly less than 1.0 according to the one sample Wilcoxon test. n_{CBV} and n_{CBF} median values are not significantly lower or higher than 1.0.

portant to note that no internal necrosis or enhancement was present, since such features may result in spuriously decreased or increased mean perfusion, respectively, in ipsilesional ROIs. In the patients with oligodendrogliomas in our cohort, the known propensity toward relatively higher tumor perfusion than comparable low grade astrocytomas is balanced by propensity for internal cystic

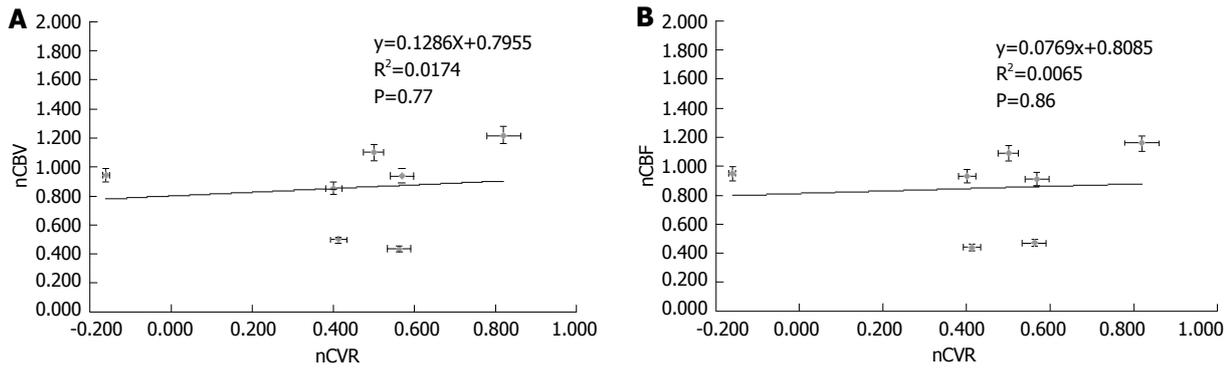


Figure 4 Normalized relative cerebral blood volume ratio (n_{CBV}) vs normalized cerebrovascular reactivity ratio (n_{CVR}) (A) and normalized relative cerebral blood flow ratio (n_{CBF}) vs normalized CVR ratio (n_{CVR}) (B) in our cohort of low grade gliomas. As described in the Methods section, these normalized ratios were calculated by dividing the mean voxel values obtained in ipsilesional regions of interest (ROIs) by those obtained in mirror contralateral ROIs. Neither n_{CBF} nor n_{CBV} demonstrated significant inverse correlation with n_{CVR} in this LGG group. LGG: Low grade gliomas; CVR: Cerebrovascular reactivity

change, thus not resulting in overall mean voxel hyperperfusion within the ipsilesional ROIs. By using overall mean perfusion metrics computed from all voxels within the ROI rather than simply voxels with maximal perfusion metrics within the ROI, we avoided the risk of spuriously high perfusion values contributing to artifactually high perfusion ratios.

DISCUSSION

All of the LGG cases in our study demonstrated reduced CVR in the tumor (i.e., ipsilesional) ROI compared to the normal contralateral hemispheric mirror (i.e., contralateral) ROI. Such regionally abnormally reduced CVR, despite the presence of clinically intact, albeit impaired, motor or language function in all of these patients is direct evidence of tumor-related NVU. The fact that no substantial corresponding regional perfusion abnormality was present in any of these cases is reflected in the absence of significant correlation between the normalized (i.e., ipsilesional to contralateral) perfusion ratios and normalized CVR ratios. The absence of tumor hyperperfusion is expected in this cohort of LGG, since such tumors, unlike HGG, are not associated with angiogenesis^[18,19]. Although reports of NVU associated with hyperperfusion in HGG exist in the literature^[20,21], few reports of NVU related to LGG exist^[22,23]. We have shown in our study that the phenomenon of NVU, as detected by regionally decreased CVR, in LGG is much more prevalent than previously thought^[24]. We have also shown the clinical feasibility of the performance of such BH CVR mapping in such a patient population. Our results suggest that BH CVR mapping should be considered in all brain tumor patients regardless of tumor grade.

The coupling mechanism between neuronal firing and blood flow changes results from a complex sequence (which we can consider as the NVU cascade) of cellular, metabolic and vascular processes involving neurons, glial/astrocytic components, neurotransmitters, chemical mediators and eventually vascular smooth muscle cells. The currently accepted explanation is that during neuro-

nal activity, synaptic release of neurotransmitters, such as glutamate, that bind to receptors on other neurons, may trigger the secondary release of vasodilatory mediators such as nitric oxide, which in turn increase CBF and CBV. These neurotransmitters such as glutamate can also act on astrocytes through different receptors, thus resulting in the release of compounds such as arachidonic acid and prostaglandin E2, which in turn result in vasoconstriction or vasodilatation, respectively, by acting on arteriolar smooth muscle^[25]. It is possible that while in HGG, aberrant neovascularity with abnormal permeability and vasoactivity may be primarily responsible for the NVU, in LGG, abnormal astrocytic function or dysfunction involving other elements of the NVU cascade may be responsible. However, little is known about the pathophysiologic mechanisms underlying such NVU in LGG.

The need to detect NVU, when present, is critical in the interpretation of clinical fMRI examinations because regional cortical NVU will result in an inability to elicit BOLD activation in the affected cortex regardless of the nature of the particular fMRI task performed. Thus, false negative activation in these cortical regions may result during performance of sensorimotor, visual, language or other cognitive tasks that are expected to activate such regions based on a priori knowledge of functional anatomy. Such false negatives may not only result in incorrect language lateralization, but also incorrect localization or underestimation of the true extent of localization of eloquent cortex, as well as possibly incorrect inferences regarding tumor-induced cortical functional reorganization^[11]. Such erroneous interpretations of task-related fMRI activation maps can result in unexpected and tragic postoperative neurological deficits related to inadvertent resection of eloquent cortex that is “BOLD-silent” directly due to NVU. The added value of BH CVR mapping in this setting lies in the additional confidence in assignment of function to areas of activation on fMRI maps in cases where no regional CVR abnormality results, as well as in proper exercising of caution in cases where functional activation is expected on a particular task in a particular cortical region where absent activation

is seen on task-based fMRI with corresponding abnormally decreased regional CVR. In the latter case, such as in this LGG cohort, one needs to acknowledge the limitation of clinical fMRI and needs to inform the referring neurosurgeon that complementary intraoperative electrophysiologic mapping will be necessary to exclude eloquent cortex in these regions of impaired CVR adjacent to or within the LGG. Such knowledge is very useful, in our clinical experience, in neurosurgical planning as well as in counseling of patients regarding the potential risks of tumor resection.

Thus, in conclusion, although our results are preliminary and based on a fairly small sample size, they suggest that BH CVR mapping in patients with LGG is both clinically feasible and capable of detecting NVU, which is a critical limitation of clinical fMRI. We furthermore note an unexpectedly high prevalence of NVU in LGG, suggesting that NVU is a commonly encountered phenomenon in brain tumors of all grades, and not just in HGG as previously thought.

COMMENTS

Background

Cerebrovascular reactivity (CVR) mapping using a breath-hold (BH) technique is a method of evaluating how responsive the microvasculature in the brain is to external stimuli. Although the mechanism for BH CVR mapping is related to transient mild increases in pCO₂ (i.e., hypercapnia), resulting in vasodilatation, this can be applied to the evaluation of standard clinical blood oxygen level dependent (BOLD) functional MRI (fMRI) examinations, where sensorimotor, visual or language/cognitive stimuli result in transient blood flow changes in brain microvasculature adjacent to activated neurons. If CVR is impaired for any reason, such as aberrant tumor neovascularity or astrocytic dysfunction due to tumor infiltration, then no BOLD response is possible on standard clinical fMRI activation studies because the BOLD response relies on intact CVR.

Research frontiers

It has been determined that CVR is impaired within or adjacent to low grade gliomas (LGG), thus compromising our ability to accurately map eloquent cortex for surgical planning using fMRI. It has already been established that such impaired CVR is present in high grade gliomas (HGG) due to tumor angiogenesis as reflected in hyperperfusion on MR perfusion imaging.

Innovations and breakthroughs

Very little is currently known about CVR in LGG. Most of the work to date relating to CVR mapping in brain tumor patients relates to applications in HGG. It is clear that tumor hyperperfusion, as detected on T2* DSC (standard clinical) MR perfusion imaging, is related to tumor angiogenesis, but in LGG, angiogenesis typically does not occur. Astrocytic dysfunction, however, is known to occur due to tumor infiltration and primary astrocytic and/or oligodendrocyte involvement by all LGGs.

Applications

The findings of a high prevalence (100% in our cohort) of abnormal CVR in LGG is of immense clinical value, because this is a potentially serious limitation of standard clinical BOLD fMRI examinations that may result in false negatives which adversely impact surgical planning. The recognition of this pitfall of fMRI is critical for proper surgical planning and counseling of patients prior to surgical resection of LGG. Further studies with larger sample sizes will need to be performed to evaluate the true prevalence of such findings in LGG of different histologic subtypes.

Terminology

Cerebrovascular reactivity mapping and BOLD fMRI have been described in the Background section above. BOLD fMRI is a method of noninvasively evaluating sensorimotor, visual, language and other cognitive functions and mapping eloquent cortical regions prior to neurosurgical intervention, particularly in brain tumor patients and patients with other conditions such as epilepsy.

Peer review

The manuscript is well written and authors are reporting a valuable research. Methods and material section is well described and results are defined well.

REFERENCES

- 1 **Ogawa S**, Lee TM, Kay AR, Tank DW. Brain magnetic resonance imaging with contrast dependent on blood oxygenation. *Proc Natl Acad Sci U S A* 1990; **87**: 9868-9872
- 2 **Norris DG**. Principles of magnetic resonance assessment of brain function. *J Magn Reson Imaging* 2006; **23**: 794-807
- 3 **Sunaert S**. Presurgical planning for tumor resectioning. *J Magn Reson Imaging* 2006; **23**: 887-905
- 4 **Pillai JJ**, Zaca D, Choudhri A. Clinical impact of integrated physiologic brain tumor imaging. *Technol Cancer Res Treat* 2010; **9**: 359-380
- 5 **Petrella JR**, Shah LM, Harris KM, Friedman AH, George TM, Sampson JH, Pekala JS, Voyvodic JT. Preoperative functional MR imaging localization of language and motor areas: effect on therapeutic decision making in patients with potentially resectable brain tumors. *Radiology* 2006; **240**: 793-802
- 6 **Håberg A**, Kvistad KA, Unsgård G, Haraldseth O. Preoperative blood oxygen level-dependent functional magnetic resonance imaging in patients with primary brain tumors: clinical application and outcome. *Neurosurgery* 2004; **54**: 902-914; discussion 914-915
- 7 **Vlieger EJ**, Majoie CB, Leenstra S, Den Heeten GJ. Functional magnetic resonance imaging for neurosurgical planning in neurooncology. *Eur Radiol* 2004; **14**: 1143-1153
- 8 **Medina LS**, Bernal B, Dunoyer C, Cervantes L, Rodriguez M, Pacheco E, Jayakar P, Morrison G, Ragheb J, Altman NR. Seizure disorders: functional MR imaging for diagnostic evaluation and surgical treatment—prospective study. *Radiology* 2005; **236**: 247-253
- 9 **Wengenroth M**, Blatow M, Guenther J, Akbar M, Tronnier VM, Stippich C. Diagnostic benefits of presurgical fMRI in patients with brain tumours in the primary sensorimotor cortex. *Eur Radiol* 2011; **21**: 1517-1525
- 10 **Giussani C**, Roux FE, Ojemann J, Sganzerla EP, Pirillo D, Papagno C. Is preoperative functional magnetic resonance imaging reliable for language areas mapping in brain tumor surgery? Review of language functional magnetic resonance imaging and direct cortical stimulation correlation studies. *Neurosurgery* 2010; **66**: 113-120
- 11 **Ulmer JL**, Krouwer HG, Mueller WM, Ugurel MS, Kocak M, Mark LP. Pseudo-reorganization of language cortical function at fMR imaging: a consequence of tumor-induced neurovascular uncoupling. *AJNR Am J Neuroradiol* 2003; **24**: 213-217
- 12 **Zaca D**, Hua J, Pillai JJ. Cerebrovascular reactivity mapping for brain tumor presurgical planning. *World J Clin Oncol* 2011; **2**: 289-298
- 13 **Holodny AI**, Schulder M, Liu WC, Wolko J, Maldjian JA, Kalnin AJ. The effect of brain tumors on BOLD functional MR imaging activation in the adjacent motor cortex: implications for image-guided neurosurgery. *AJNR Am J Neuroradiol* 2000; **21**: 1415-1422
- 14 **Chen CM**, Hou BL, Holodny AI. Effect of age and tumor grade on BOLD functional MR imaging in preoperative assessment of patients with glioma. *Radiology* 2008; **248**: 971-978
- 15 **Magon S**, Basso G, Farace P, Ricciardi GK, Beltramello A, Sbarbati A. Reproducibility of BOLD signal change induced by breath holding. *Neuroimage* 2009; **45**: 702-712
- 16 **Boxerman JL**, Schmainda KM, Weisskoff RM. Relative cerebral blood volume maps corrected for contrast agent extravasation significantly correlate with glioma tumor grade, whereas uncorrected maps do not. *AJNR Am J Neuroradiol* 2006; **27**: 859-867

- 17 **Birn RM**, Smith MA, Jones TB, Bandettini PA. The respiration response function: the temporal dynamics of fMRI signal fluctuations related to changes in respiration. *Neuroimage* 2008; **40**: 644-654
- 18 **Provenzale JM**, Wang GR, Brenner T, Petrella JR, Sorensen AG. Comparison of permeability in high-grade and low-grade brain tumors using dynamic susceptibility contrast MR imaging. *AJR Am J Roentgenol* 2002; **178**: 711-716
- 19 **Law M**, Yang S, Babb JS, Knopp EA, Golfinos JG, Zagzag D, Johnson G. Comparison of cerebral blood volume and vascular permeability from dynamic susceptibility contrast-enhanced perfusion MR imaging with glioma grade. *AJNR Am J Neuroradiol* 2004; **25**: 746-755
- 20 **Hou BL**, Bradbury M, Peck KK, Petrovich NM, Gutin PH, Holodny AI. Effect of brain tumor neovasculature defined by rCBV on BOLD fMRI activation volume in the primary motor cortex. *Neuroimage* 2006; **32**: 489-497
- 21 **Lüdemann L**, Förchler A, Grieger W, Zimmer C. BOLD signal in the motor cortex shows a correlation with the blood volume of brain tumors. *J Magn Reson Imaging* 2006; **23**: 435-443
- 22 **Schreiber A**, Hubbe U, Ziyeh S, Hennig J. The influence of gliomas and nonglial space-occupying lesions on blood-oxygen-level-dependent contrast enhancement. *AJNR Am J Neuroradiol* 2000; **21**: 1055-1063
- 23 **Ulmer JL**, Hacein-Bey L, Mathews VP, Mueller WM, DeYoe EA, Prost RW, Meyer GA, Krouwer HG, Schmainda KM. Lesion-induced pseudo-dominance at functional magnetic resonance imaging: implications for preoperative assessments. *Neurosurgery* 2004; **55**: 569-579; discussion 580-581
- 24 **Jiang Z**, Krainik A, David O, Salon C, Troprès I, Hoffmann D, Pannetier N, Barbier EL, Bombin ER, Warnking J, Pasteris C, Chabardes S, Berger F, Grand S, Segebarth C, Gay E, Le Bas JF. Impaired fMRI activation in patients with primary brain tumors. *Neuroimage* 2010; **52**: 538-548
- 25 **Attwell D**, Buchan AM, Charpak S, Lauritzen M, MacVicar BA, Newman EA. Glial and neuronal control of brain blood flow. *Nature* 2010; **468**: 232-243

S- Editor Yang XC L- Editor Webster JR E- Editor Yang XC

Acknowledgments to reviewers of *World Journal of Clinical Oncology*

Many reviewers have contributed their expertise and time to the peer review, a critical process to ensure the quality of *World Journal of Clinical Oncology*. The editors and authors of the articles submitted to the journal are grateful to the following reviewers for evaluating the articles (including those published in this issue and those rejected for this issue) during the last editing time period.

Ali Syed Arbab, MD, PhD, Associate Scientist and Director, Cellular and Molecular Imaging Laboratory, Department of Radiology, Henry Ford Hospital, 1 Ford Place, 2F, PO Box 82, Detroit, MI 48202, United States

Joan Carles, MD, PhD, Director, GU, CNS and Sarcoma

Program, Department of Medical Oncology, Passeig Vall d'Hebron, 119-129, 08035 Barcelona, Spain

Robert Mandic, MD, Associate Professor, Head of Research Laboratory, Department of Otolaryngology, Head and Neck Surgery, University Hospital Giessen and Marburg, Campus Marburg, Deutschhausstrasse 3, D-35037 Marburg, Germany

Shahrokh F Shariat, MD, PhD, Associate Professor, Department of Urology, New York Presbyterian Hospital, Weill Medical College of Cornell University, 525 East 68th Street, Starr 900, New York, NY 10021, United States

Ke Zen, PhD, Chang-Jiang Professor, Vice Dean, School of Life Sciences, Nanjing University, Hankou Road 22, Nanjing 210093, Jiangsu Province, China

Events Calendar 2011

January 13-14, 2011

3rd Breast-Gynecology International Cancer Conference BGICC, Cairo, Egypt

January 15-16, 2011

Melanoma 2011: 21st Annual Cutaneous Malignancy Update, San Diego, CA, United States

January 15, 2011

Current Trends in Breast Cancer: Updates From the 2010 San Antonio Breast Cancer Symposium, Dallas, TX, United States

January 20-22, 2011

Gastrointestinal Cancers Symposium 2011, San Francisco, CA, United States

January 21-23, 2011

8th Meeting of the EAU Section of Oncological Urology, London, England, United Kingdom

January 27-28, 2011

2nd National Conference: Recent Advances in Renal and Bladder Cancer, London, United Kingdom

January 27-28, 2011

8th Annual Cancer Drugs Research & Development, San Diego, CA, United States

February 10-12, 2011

17th Annual NOCR Meeting, Las Vegas, NV, United States

February 19-22, 2011

Scripps Cancer Center's 31st Annual Conference: Clinical

Hematology and Oncology, San Diego, CA, United States

February 24-26, 2011

European Multidisciplinary Conference in Thoracic Oncology (Lung 2011-EMCTO), Lugano, Switzerland

February 25-27, 2011

7th European Congress on Hematologic Malignancies: From Clinical Science to Clinical Practice, Budapest, Hungary

March 02-05, 2011

64th Society of Surgical Oncology Annual Cancer Symposium 2011, San Antonio, TX, United States

March 04-06, 2011

8th Annual Oncology Nursing Advanced Practice: Innovation through Practice, San Diego, CA, United States

March 07-09, 2011

9th International Symposium on Targeted Anticancer Therapies, Paris, France

March 09-13, 2011

16th National Comprehensive Cancer Network Annual Conference (NCCN 2011), Hollywood, FL, United States

March 11-12, 2011

12th European Congress: Perspectives in Lung Cancer, Torino, Italy

March 14-18, 2011

Oncology Imaging Update in Costa Rica, Guanacaste, Costa Rica

March 17-19, 2011

International Cancer Prevention Update Symposium, New York, United States

March 18-22, 2011

Vienna, Austria 26th Annual EAU Congress

April 02-06, 2011

AACR 102nd Annual Meeting, Orlando, FL, United States

April 08-10, 2011

Asian Oncology Summit 2011, Hong Kong, China

April 20-23, 2011

9th International Gastric Cancer Congress, Seoul, South Korea

April 29-30, 2011

Cancer Survivorship Conference, Minneapolis, MN, United States

May 23-24, 2011

4th International Conference on Ovarian Cancer Screening, London, United Kingdom

June 03-07, 2011

47th American Society of Clinical Oncology Annual Meeting, Chicago, IL, United States

June 20-23, 2011

7th EADO Congress European Association of Dermato-Oncology, Nantes, France

June 22-25, 2011

ESMO Conference: 13th World Congress on Gastrointestinal Cancer, Barcelona, Spain

June 23-25, 2011

"MASCC/ISOO 2011 International Symposium, Athens, Greece

July 03-07, 2011

14th World Conference on Lung Cancer, Amsterdam, Netherlands

July 14-17, 2011

3rd World Congress of the International Academy of Oral Oncology 2011, Singapore, Singapore

August 15-17, 2011

International Conference and Exhibition on Cancer Science & Therapy, Las Vegas, Nevada, United States

September 1-3, 2011

Tri-Society Head and Neck Oncology, Singapore, Singapore

September 7-10, 2011

Hallmarks and Horizons of Cancer, Lausanne, Switzerland

September 23-27, 2011

Joint 16th ECCO and 36th ESMO Multidisciplinary Cancer Congress, Stockholm, Sweden

October 06-07, 2011

Current Status and Future of Anti-Cancer Targeted Therapies, Buenos Aires, Argentina

November 30-December 03, 2011

AORTIC 2011-Entering the 21st Century for Cancer Control in Africa, Cairo, Egypt

November 6-9, 2011

NCRI Cancer Conference, Liverpool, United Kingdom

November 10-12, 2011

21st Asia Pacific Cancer Conference 2011, Kuala Lumpur, Wilayah Persekutuan, Malaysia

GENERAL INFORMATION

World Journal of Clinical Oncology (*World J Clin Oncol*, *WJCO*, online ISSN 2218-4333, DOI: 10.5306) is a monthly peer-reviewed, online, open-access (OA), journal supported by an editorial board consisting of 316 experts in oncology from 33 countries.

The biggest advantage of the OA model is that it provides free, full-text articles in PDF and other formats for experts and the public without registration, which eliminates the obstacle that traditional journals possess and usually delays the speed of the propagation and communication of scientific research results. The open access model has been proven to be a true approach that may achieve the ultimate goal of the journals, i.e. the maximization of the value to the readers, authors and society.

Maximization of personal benefits

The role of academic journals is to exhibit the scientific levels of a country, a university, a center, a department, and even a scientist, and build an important bridge for communication between scientists and the public. As we all know, the significance of the publication of scientific articles lies not only in disseminating and communicating innovative scientific achievements and academic views, as well as promoting the application of scientific achievements, but also in formally recognizing the "priority" and "copyright" of innovative achievements published, as well as evaluating research performance and academic levels. So, to realize these desired attributes of *WJCO* and create a well-recognized journal, the following four types of personal benefits should be maximized. The maximization of personal benefits refers to the pursuit of the maximum personal benefits in a well-considered optimal manner without violation of the laws, ethical rules and the benefits of others. (1) Maximization of the benefits of editorial board members: The primary task of editorial board members is to give a peer review of an unpublished scientific article *via* online office system to evaluate its innovativeness, scientific and practical values and determine whether it should be published or not. During peer review, editorial board members can also obtain cutting-edge information in that field at first hand. As leaders in their field, they have priority to be invited to write articles and publish commentary articles. We will put peer reviewers' names and affiliations along with the article they reviewed in the journal to acknowledge their contribution; (2) Maximization of the benefits of authors: Since *WJCO* is an open-access journal, readers around the world can immediately download and read, free of charge, high-quality, peer-reviewed articles from *WJCO* official website, thereby realizing the goals and significance of the communication between authors and peers as well as public reading; (3) Maximization of the benefits of readers: Readers can read or use, free of charge, high-quality peer-reviewed articles without any limits, and cite the arguments, viewpoints, concepts, theories, methods, results, conclusion or facts and data of pertinent literature so as to validate the innovativeness, scientific and practical values of their own research achievements, thus ensuring that their articles have novel arguments or viewpoints, solid evidence and correct conclusion; and (4) Maximization of the benefits of employees: It is an iron law that a first-class journal is unable to exist without first-class editors, and only first-class editors can create a first-class academic journal. We insist on strengthening our team cultivation and construction so that every employee, in an open, fair and transparent environment, could contribute their wisdom to edit and publish high-quality articles, thereby realizing the maximization of the personal benefits of editorial board members,

authors and readers, and yielding the greatest social and economic benefits.

Aims and scope

The aim of *WJCO* is to report rapidly new theories, methods and techniques for prevention, diagnosis, treatment, rehabilitation and nursing in the field of oncology. *WJCO* covers etiology, epidemiology, evidence-based medicine, informatics, diagnostic imaging, endoscopy, tumor recurrence and metastasis, tumor stem cells, radiotherapy, chemotherapy, interventional radiology, palliative therapy, clinical chemotherapy, biological therapy, minimally invasive therapy, physiotherapy, psycho-oncology, comprehensive therapy, oncology-related traditional medicine, integrated Chinese and Western medicine, and nursing. *WJCO* covers tumors in various organs/tissues, including the female reproductive system, bone and soft tissue, respiratory system, urinary system, endocrine system, skin, breast, nervous system, head and neck, digestive system, and hematologic and lymphatic system. The journal also publishes original articles and reviews that report the results of applied and basic research in fields related to oncology, such as immunology, physiopathology, cell biology, pharmacology, medical genetics, and pharmacology of Chinese herbs.

Columns

The columns in the issues of *WJCO* will include: (1) Editorial: To introduce and comment on major advances and developments in the field; (2) Frontier: To review representative achievements, comment on the state of current research, and propose directions for future research; (3) Topic Highlight: This column consists of three formats, including (A) 10 invited review articles on a hot topic, (B) a commentary on common issues of this hot topic, and (C) a commentary on the 10 individual articles; (4) Observation: To update the development of old and new questions, highlight unsolved problems, and provide strategies on how to solve the questions; (5) Guidelines for Basic Research: To provide Guidelines for basic research; (6) Guidelines for Clinical Practice: To provide guidelines for clinical diagnosis and treatment; (7) Review: To review systemically progress and unresolved problems in the field, comment on the state of current research, and make suggestions for future work; (8) Original Articles: To report innovative and original findings in oncology; (9) Brief Articles: To briefly report the novel and innovative findings in oncology; (10) Case Report: To report a rare or typical case; (11) Letters to the Editor: To discuss and make reply to the contributions published in *WJCO*, or to introduce and comment on a controversial issue of general interest; (12) Book Reviews: To introduce and comment on quality monographs of oncology; and (13) Guidelines: To introduce consensus and guidelines reached by international and national academic authorities worldwide on the research oncology.

Name of journal

World Journal of Clinical Oncology

ISSN

ISSN 2218-4333 (online)

Indexed and Abstracted in

PubMed Central, PubMed, Digital Object Identifier, and Directory of Open Access Journals.

Published by

Baishideng Publishing Group Co., Limited

SPECIAL STATEMENT

All articles published in this journal represent the viewpoints of the authors except where indicated otherwise.

Biostatistical editing

Statistical review is performed after peer review. We invite an expert in Biomedical Statistics from to evaluate the statistical method used in the paper, including *t*-test (group or paired comparisons), chi-squared test, Ridit, probit, logit, regression (linear, curvilinear, or stepwise), correlation, analysis of variance, analysis of covariance, *etc.* The reviewing points include: (1) Statistical methods should be described when they are used to verify the results; (2) Whether the statistical techniques are suitable or correct; (3) Only homogeneous data can be averaged. Standard deviations are preferred to standard errors. Give the number of observations and subjects (*n*). Losses in observations, such as drop-outs from the study should be reported; (4) Values such as ED50, LD50, IC50 should have their 95% confidence limits calculated and compared by weighted probit analysis (Bliss and Finney); and (5) The word 'significantly' should be replaced by its synonyms (if it indicates extent) or the *P* value (if it indicates statistical significance).

Conflict-of-interest statement

In the interests of transparency and to help reviewers assess any potential bias, *WJCO* requires authors of all papers to declare any competing commercial, personal, political, intellectual, or religious interests in relation to the submitted work. Referees are also asked to indicate any potential conflict they might have reviewing a particular paper. Before submitting, authors are suggested to read "Uniform Requirements for Manuscripts Submitted to Biomedical Journals: Ethical Considerations in the Conduct and Reporting of Research: Conflicts of Interest" from International Committee of Medical Journal Editors (ICMJE), which is available at: http://www.icmje.org/ethical_4conflicts.html.

Sample wording: [Name of individual] has received fees for serving as a speaker, a consultant and an advisory board member for [names of organizations], and has received research funding from [names of organization]. [Name of individual] is an employee of [name of organization]. [Name of individual] owns stocks and shares in [name of organization]. [Name of individual] owns patent [patent identification and brief description].

Statement of informed consent

Manuscripts should contain a statement to the effect that all human studies have been reviewed by the appropriate ethics committee or it should be stated clearly in the text that all persons gave their informed consent prior to their inclusion in the study. Details that might disclose the identity of the subjects under study should be omitted. Authors should also draw attention to the Code of Ethics of the World Medical Association (Declaration of Helsinki, 1964, as revised in 2004).

Statement of human and animal rights

When reporting the results from experiments, authors should follow the highest standards and the trial should conform to Good Clinical Practice (for example, US Food and Drug Administration Good Clinical Practice in FDA-Regulated Clinical Trials; UK Medicines Research Council Guidelines for Good Clinical Practice in Clinical Trials) and/or the World Medical Association Declaration of Helsinki. Generally, we suggest authors follow the lead investigator's national standard. If doubt exists whether the research was conducted in accordance with the above standards, the authors must explain the rationale for their approach and demonstrate that the institutional review body explicitly approved the doubtful aspects of the study.

Before submitting, authors should make their study approved by the relevant research ethics committee or institutional review board. If human participants were involved, manuscripts must be accompanied by a statement that the experiments were undertaken with the understanding and appropriate informed consent of each. Any personal item or information will not be published without explicit consents from the involved patients. If experimental animals were used, the materials and methods (experimental procedures) section must

clearly indicate that appropriate measures were taken to minimize pain or discomfort, and details of animal care should be provided.

SUBMISSION OF MANUSCRIPTS

Manuscripts should be typed in 1.5 line spacing and 12 pt. Book Antiqua with ample margins. Number all pages consecutively, and start each of the following sections on a new page: Title Page, Abstract, Introduction, Materials and Methods, Results, Discussion, Acknowledgements, References, Tables, Figures, and Figure Legends. Neither the editors nor the publisher are responsible for the opinions expressed by contributors. Manuscripts formally accepted for publication become the permanent property of Baishideng Publishing Group Co., Limited, and may not be reproduced by any means, in whole or in part, without the written permission of both the authors and the publisher. We reserve the right to copy-edit and put onto our website accepted manuscripts. Authors should follow the relevant guidelines for the care and use of laboratory animals of their institution or national animal welfare committee. For the sake of transparency in regard to the performance and reporting of clinical trials, we endorse the policy of the ICMJE to refuse to publish papers on clinical trial results if the trial was not recorded in a publicly-accessible registry at its outset. The only register now available, to our knowledge, is <http://www.clinicaltrials.gov> sponsored by the United States National Library of Medicine and we encourage all potential contributors to register with it. However, in the case that other registers become available you will be duly notified. A letter of recommendation from each author's organization should be provided with the contributed article to ensure the privacy and secrecy of research is protected.

Authors should retain one copy of the text, tables, photographs and illustrations because rejected manuscripts will not be returned to the author(s) and the editors will not be responsible for loss or damage to photographs and illustrations sustained during mailing.

Online submissions

Manuscripts should be submitted through the Online Submission System at: <http://www.wjgnet.com/2218-4333/office>. Authors are highly recommended to consult the ONLINE INSTRUCTIONS TO AUTHORS (http://www.wjgnet.com/2218-4333/g_info_20100722172206.htm) before attempting to submit online. For assistance, authors encountering problems with the Online Submission System may send an email describing the problem to wjco@wjgnet.com, or by telephone: +86-10-85381892. If you submit your manuscript online, do not make a postal contribution. Repeated online submission for the same manuscript is strictly prohibited.

MANUSCRIPT PREPARATION

All contributions should be written in English. All articles must be submitted using word-processing software. All submissions must be typed in 1.5 line spacing and 12 pt. Book Antiqua with ample margins. Style should conform to our house format. Required information for each of the manuscript sections is as follows:

Title page

Title: Title should be less than 12 words.

Running title: A short running title of less than 6 words should be provided.

Authorship: Authorship credit should be in accordance with the standard proposed by International Committee of Medical Journal Editors, based on (1) substantial contributions to conception and design, acquisition of data, or analysis and interpretation of data; (2) drafting the article or revising it critically for important intellectual content; and (3) final approval of the version to be published. Authors should meet conditions 1, 2, and 3.

Institution: Author names should be given first, then the complete name of institution, city, province and postcode. For example, Xu-Chen Zhang, Li-Xin Mei, Department of Pathology, Chengde Medical College, Chengde 067000, Hebei Province, China. One author may be represented from two institutions, for example, George

Sgourakis, Department of General, Visceral, and Transplantation Surgery, Essen 45122, Germany; George Sgourakis, 2nd Surgical Department, Korgialenio-Benakio Red Cross Hospital, Athens 15451, Greece

Author contributions: The format of this section should be: Author contributions: Wang CL and Liang L contributed equally to this work; Wang CL, Liang L, Fu JF, Zou CC, Hong F and Wu XM designed the research; Wang CL, Zou CC, Hong F and Wu XM performed the research; Xue JZ and Lu JR contributed new reagents/analytic tools; Wang CL, Liang L and Fu JF analyzed the data; and Wang CL, Liang L and Fu JF wrote the paper.

Supportive foundations: The complete name and number of supportive foundations should be provided, e.g. Supported by National Natural Science Foundation of China, No. 30224801

Correspondence to: Only one corresponding address should be provided. Author names should be given first, then author title, affiliation, the complete name of institution, city, postcode, province, country, and email. All the letters in the email should be in lower case. A space interval should be inserted between country name and email address. For example, Montgomery Bissell, MD, Professor of Medicine, Chief, Liver Center, Gastroenterology Division, University of California, Box 0538, San Francisco, CA 94143, United States. montgomery.bissell@ucsf.edu

Telephone and fax: Telephone and fax should consist of +, country number, district number and telephone or fax number, e.g. Telephone: +86-10-59080039 Fax: +86-10-85381893

Peer reviewers: All articles received are subject to peer review. Normally, three experts are invited for each article. Decision for acceptance is made only when at least two experts recommend an article for publication. Reviewers for accepted manuscripts are acknowledged in each manuscript, and reviewers of articles which were not accepted will be acknowledged at the end of each issue. To ensure the quality of the articles published in *WJCO*, reviewers of accepted manuscripts will be announced by publishing the name, title/position and institution of the reviewer in the footnote accompanying the printed article. For example, reviewers: Professor Jing-Yuan Fang, Shanghai Institute of Digestive Disease, Shanghai, Affiliated Renji Hospital, Medical Faculty, Shanghai Jiaotong University, Shanghai, China; Professor Xin-Wei Han, Department of Radiology, The First Affiliated Hospital, Zhengzhou University, Zhengzhou, Henan Province, China; and Professor Anren Kuang, Department of Nuclear Medicine, Huaxi Hospital, Sichuan University, Chengdu, Sichuan Province, China.

Abstract

There are unstructured abstracts (no more than 256 words) and structured abstracts (no more than 480). The specific requirements for structured abstracts are as follows:

An informative, structured abstracts of no more than 480 words should accompany each manuscript. Abstracts for original contributions should be structured into the following sections. AIM (no more than 20 words): Only the purpose should be included. Please write the aim as the form of "To investigate/study/..."; MATERIALS AND METHODS (no more than 140 words); RESULTS (no more than 294 words): You should present *P* values where appropriate and must provide relevant data to illustrate how they were obtained, e.g. 6.92 ± 3.86 vs 3.61 ± 1.67 , $P < 0.001$; CONCLUSION (no more than 26 words).

Key words

Please list 5-10 key words, selected mainly from *Index Medicus*, which reflect the content of the study.

Text

For articles of these sections, original articles and brief articles, the main text should be structured into the following sections: INTRO-

DUCTION, MATERIALS AND METHODS, RESULTS and DISCUSSION, and should include appropriate Figures and Tables. Data should be presented in the main text or in Figures and Tables, but not in both. The main text format of these sections, editorial, topic highlight, case report, letters to the editors, can be found at: http://www.wjgnet.com/2218-4333/g_info_list.htm.

Illustrations

Figures should be numbered as 1, 2, 3, *etc.*, and mentioned clearly in the main text. Provide a brief title for each figure on a separate page. Detailed legends should not be provided under the figures. This part should be added into the text where the figures are applicable. Figures should be either Photoshop or Illustrator files (in tiff, eps, jpeg formats) at high-resolution. Examples can be found at: <http://www.wjgnet.com/1007-9327/13/4520.pdf>; <http://www.wjgnet.com/1007-9327/13/4554.pdf>; <http://www.wjgnet.com/1007-9327/13/4891.pdf>; <http://www.wjgnet.com/1007-9327/13/4986.pdf>; <http://www.wjgnet.com/1007-9327/13/4498.pdf>. Keeping all elements compiled is necessary in line-art image. Scale bars should be used rather than magnification factors, with the length of the bar defined in the legend rather than on the bar itself. File names should identify the figure and panel. Avoid layering type directly over shaded or textured areas. Please use uniform legends for the same subjects. For example: Figure 1 Pathological changes in atrophic gastritis after treatment. A: ...; B: ...; C: ...; D: ...; E: ...; F: ...; G: ... *etc.* It is our principle to publish high resolution-figures for the printed and E-versions.

Tables

Three-line tables should be numbered 1, 2, 3, *etc.*, and mentioned clearly in the main text. Provide a brief title for each table. Detailed legends should not be included under tables, but rather added into the text where applicable. The information should complement, but not duplicate the text. Use one horizontal line under the title, a second under column heads, and a third below the Table, above any footnotes. Vertical and italic lines should be omitted.

Notes in tables and illustrations

Data that are not statistically significant should not be noted. ^a*P* < 0.05, ^b*P* < 0.01 should be noted (*P* > 0.05 should not be noted). If there are other series of *P* values, ^c*P* < 0.05 and ^d*P* < 0.01 are used. A third series of *P* values can be expressed as ^e*P* < 0.05 and ^f*P* < 0.01. Other notes in tables or under illustrations should be expressed as ¹F, ²F, ³F; or sometimes as other symbols with a superscript (Arabic numerals) in the upper left corner. In a multi-curve illustration, each curve should be labeled with ●, ○, ■, □, ▲, △, *etc.*, in a certain sequence.

Acknowledgments

Brief acknowledgments of persons who have made genuine contributions to the manuscript and who endorse the data and conclusions should be included. Authors are responsible for obtaining written permission to use any copyrighted text and/or illustrations.

REFERENCES

Coding system

The author should number the references in Arabic numerals according to the citation order in the text. Put reference numbers in square brackets in superscript at the end of citation content or after the cited author's name. For citation content which is part of the narration, the coding number and square brackets should be typeset normally. For example, "Crohn's disease (CD) is associated with increased intestinal permeability^{1,2}". If references are cited directly in the text, they should be put together within the text, for example, "From references^{19,22-24}, we know that..."

When the authors write the references, please ensure that the order in text is the same as in the references section, and also ensure the spelling accuracy of the first author's name. Do not list the same citation twice.

Instructions to authors

PMID and DOI

Pleased provide PubMed citation numbers to the reference list, e.g. PMID and DOI, which can be found at <http://www.ncbi.nlm.nih.gov/sites/entrez?db=pubmed> and <http://www.crossref.org/SimpleTextQuery/>, respectively. The numbers will be used in E-version of this journal.

Style for journal references

Authors: the name of the first author should be typed in bold-faced letters. The family name of all authors should be typed with the initial letter capitalized, followed by their abbreviated first and middle initials. (For example, Lian-Sheng Ma is abbreviated as Ma LS, Bo-Rong Pan as Pan BR). The title of the cited article and italicized journal title (journal title should be in its abbreviated form as shown in PubMed), publication date, volume number (in black), start page, and end page [PMID: 11819634 DOI: 10.3748/wjg.13.5396].

Style for book references

Authors: the name of the first author should be typed in bold-faced letters. The surname of all authors should be typed with the initial letter capitalized, followed by their abbreviated middle and first initials. (For example, Lian-Sheng Ma is abbreviated as Ma LS, Bo-Rong Pan as Pan BR) Book title. Publication number. Publication place: Publication press, Year: start page and end page.

Format

Journals

English journal article (list all authors and include the PMID where applicable)

- 1 **Jung EM**, Clevert DA, Schreyer AG, Schmitt S, Rennert J, Kubale R, Feuerbach S, Jung F. Evaluation of quantitative contrast harmonic imaging to assess malignancy of liver tumors: A prospective controlled two-center study. *World J Gastroenterol* 2007; **13**: 6356-6364 [PMID: 18081224 DOI: 10.3748/wjg.13.6356]

Chinese journal article (list all authors and include the PMID where applicable)

- 2 **Lin GZ**, Wang XZ, Wang P, Lin J, Yang FD. Immunologic effect of Jianpi Yishen decoction in treatment of Pixu-diarhoea. *Shijie Huaren Xiaohua Zazhi* 1999; **7**: 285-287

In press

- 3 **Tian D**, Araki H, Stahl E, Bergelson J, Kreitman M. Signature of balancing selection in Arabidopsis. *Proc Natl Acad Sci USA* 2006; In press

Organization as author

- 4 **Diabetes Prevention Program Research Group**. Hypertension, insulin, and proinsulin in participants with impaired glucose tolerance. *Hypertension* 2002; **40**: 679-686 [PMID: 12411462 PMID:2516377 DOI:10.1161/01.HYP.0000035706.28494.09]

Both personal authors and an organization as author

- 5 **Vallancien G**, Emberton M, Harving N, van Moorselaar RJ; Alf-One Study Group. Sexual dysfunction in 1, 274 European men suffering from lower urinary tract symptoms. *J Urol* 2003; **169**: 2257-2261 [PMID: 12771764 DOI:10.1097/01.ju.0000067940.76090.73]

No author given

- 6 21st century heart solution may have a sting in the tail. *BMJ* 2002; **325**: 184 [PMID: 12142303 DOI:10.1136/bmj.325.7357.184]

Volume with supplement

- 7 **Geraud G**, Spierings EL, Keywood C. Tolerability and safety of frovatriptan with short- and long-term use for treatment of migraine and in comparison with sumatriptan. *Headache* 2002; **42** Suppl 2: S93-99 [PMID: 12028325 DOI:10.1046/j.1526-4610.42.s2.7.x]

Issue with no volume

- 8 **Banit DM**, Kaufer H, Hartford JM. Intraoperative frozen section analysis in revision total joint arthroplasty. *Clin Orthop Relat Res* 2002; (**401**): 230-238 [PMID: 12151900 DOI:10.1097/00003086-200208000-00026]

No volume or issue

- 9 Outreach: Bringing HIV-positive individuals into care. *HRS-A Careaction* 2002; 1-6 [PMID: 12154804]

Books

Personal author(s)

- 10 **Sherlock S**, Dooley J. Diseases of the liver and biliary system. 9th ed. Oxford: Blackwell Sci Pub, 1993: 258-296

Chapter in a book (list all authors)

- 11 **Lam SK**. Academic investigator's perspectives of medical treatment for peptic ulcer. In: Swabb EA, Azabo S. Ulcer disease: investigation and basis for therapy. New York: Marcel Dekker, 1991: 431-450

Author(s) and editor(s)

- 12 **Breedlove GK**, Schorfheide AM. Adolescent pregnancy. 2nd ed. Wiczorek RR, editor. White Plains (NY): March of Dimes Education Services, 2001: 20-34

Conference proceedings

- 13 **Harnden P**, Joffe JK, Jones WG, editors. Germ cell tumours V. Proceedings of the 5th Germ cell tumours Conference; 2001 Sep 13-15; Leeds, UK. New York: Springer, 2002: 30-56

Conference paper

- 14 **Christensen S**, Oppacher F. An analysis of Koza's computational effort statistic for genetic programming. In: Foster JA, Lutton E, Miller J, Ryan C, Tettamanzi AG, editors. Genetic programming. EuroGP 2002: Proceedings of the 5th European Conference on Genetic Programming; 2002 Apr 3-5; Kinsdale, Ireland. Berlin: Springer, 2002: 182-191

Electronic journal (list all authors)

- 15 Morse SS. Factors in the emergence of infectious diseases. *Emerg Infect Dis* serial online, 1995-01-03, cited 1996-06-05; 1(1): 24 screens. Available from: URL: <http://www.cdc.gov/ncidod/eid/index.htm>

Patent (list all authors)

- 16 **Pagedas AC**, inventor; Ancel Surgical R&D Inc., assignee. Flexible endoscopic grasping and cutting device and positioning tool assembly. United States patent US 20020103498. 2002 Aug 1

Statistical data

Write as mean \pm SD or mean \pm SE.

Statistical expression

Express *t* test as *t* (in italics), *F* test as *F* (in italics), chi square test as χ^2 (in Greek), related coefficient as *r* (in italics), degree of freedom as *v* (in Greek), sample number as *n* (in italics), and probability as *P* (in italics).

Units

Use SI units. For example: body mass, *m* (B) = 78 kg; blood pressure, *p* (B) = 16.2/12.3 kPa; incubation time, *t* (incubation) = 96 h, blood glucose concentration, *c* (glucose) 6.4 \pm 2.1 mmol/L; blood CEA mass concentration, *p* (CEA) = 8.6 24.5 μ g/L; CO₂ volume fraction, 50 mL/L CO₂, not 5% CO₂; likewise for 40 g/L formaldehyde, not 10% formalin; and mass fraction, 8 ng/g, *etc.* Arabic numerals such as 23, 243, 641 should be read 23 243 641.

The format for how to accurately write common units and quantum numbers can be found at: http://www.wjgnet.com/2218-4333/g_info_20100723153305.htm.

Abbreviations

Standard abbreviations should be defined in the abstract and on first mention in the text. In general, terms should not be abbreviated unless they are used repeatedly and the abbreviation is helpful to the reader. Permissible abbreviations are listed in Units, Symbols and Abbreviations: A Guide for Biological and Medical Editors and Authors (Ed. Baron DN, 1988) published by The Royal Society of Medicine, London. Certain commonly used abbreviations, such as DNA, RNA, HIV, LD50, PCR, HBV, ECG, WBC, RBC, CT, ESR, CSF, IgG, ELISA, PBS, ATP, EDTA, mAb, can be used directly without further explanation.

Italics

Quantities: *t* time or temperature, *c* concentration, *A* area, *l* length, *m* mass, *V* volume.

Genotypes: *gyrA*, *arg 1*, *c myc*, *c fos*, etc.

Restriction enzymes: *EcoRI*, *HindIII*, *BamHI*, *Kho I*, *Kpn I*, etc.

Biology: *H. pylori*, *E. coli*, etc.

Examples for paper writing

Editorial: http://www.wjgnet.com/2218-4333/g_info_20100723140942.htm

Frontier: http://www.wjgnet.com/2218-4333/g_info_20100723141035.htm

Topic highlight: http://www.wjgnet.com/2218-4333/g_info_20100723141239.htm

Observation: http://www.wjgnet.com/2218-4333/g_info_20100723141532.htm

Guidelines for basic research: http://www.wjgnet.com/2218-4333/g_info_20100723142040.htm

Guidelines for clinical practice: http://www.wjgnet.com/2218-5836/g_info_20100723142248.htm

Review: http://www.wjgnet.com/2218-4333/g_info_20100723145519.htm

Original articles: http://www.wjgnet.com/2218-4333/g_info_20100723145856.htm

Brief articles: http://www.wjgnet.com/2218-4333/g_info_20100723150253.htm

Case report: http://www.wjgnet.com/2218-4333/g_info_20100723150420.htm

Letters to the editor: http://www.wjgnet.com/2218-4333/g_info_20100723150642.htm

Book reviews: http://www.wjgnet.com/2218-4333/g_info_20100723150839.htm

Guidelines: http://www.wjgnet.com/2218-4333/g_info_20100723150924.htm

SUBMISSION OF THE REVISED MANUSCRIPTS AFTER ACCEPTED

Please revise your article according to the revision policies of *WJCO*. The revised version including manuscript and high-resolution image figures (if any) should be copied on a floppy or compact disk. The author should send the revised manuscript, along with printed high-resolution color or black and white photos, copyright transfer letter, and responses to the reviewers by courier (such as EMS/DHL).

Editorial Office

World Journal of Clinical Oncology

Editorial Department: Room 903, Building D,

Ocean International Center,
No. 62 Dongsihuan Zhonglu,
Chaoyang District, Beijing 100025, China
E-mail: wjco@wjgnet.com
<http://www.wjgnet.com>
Telephone: +86-10-8538-1892
Fax: +86-10-8538-1893

Language evaluation

The language of a manuscript will be graded before it is sent for revision. (1) Grade A: priority publishing; (2) Grade B: minor language polishing; (3) Grade C: a great deal of language polishing needed; and (4) Grade D: rejected. Revised articles should reach Grade A or B.

Copyright assignment form

Please download a Copyright assignment form from http://www.wjgnet.com/2218-4333/g_info_20100723153117.htm.

Responses to reviewers

Please revise your article according to the comments/suggestions provided by the reviewers. The format for responses to the reviewers' comments can be found at: http://www.wjgnet.com/2218-4333/g_info_20100723152755.htm.

Proof of financial support

For paper supported by a foundation, authors should provide a copy of the document and serial number of the foundation.

Links to documents related to the manuscript

WJCO will be initiating a platform to promote dynamic interactions between the editors, peer reviewers, readers and authors. After a manuscript is published online, links to the PDF version of the submitted manuscript, the peer-reviewers' report and the revised manuscript will be put on-line. Readers can make comments on the peer reviewer's report, authors' responses to peer reviewers, and the revised manuscript. We hope that authors will benefit from this feedback and be able to revise the manuscript accordingly in a timely manner.

Science news releases

Authors of accepted manuscripts are suggested to write a science news item to promote their articles. The news will be released rapidly at EurekAlert/AAAS (<http://www.eurekalert.org>). The title for news items should be less than 90 characters; the summary should be less than 75 words; and main body less than 500 words. Science news items should be lawful, ethical, and strictly based on your original content with an attractive title and interesting pictures.

Publication fee

WJCO is an international, peer-reviewed, Open-Access, online journal. Articles published by this journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non commercial and is otherwise in compliance with the license. Authors of accepted articles must pay a publication fee. The related standards are as follows. Publication fee: 1300 USD per article. Editorial, topic highlights, book reviews and letters to the editor are published free of charge.