

World Journal of *Diabetes*

World J Diabetes 2015 November 10; 6(15): 1285-1308





Editorial Board

2011-2015

The *World Journal of Diabetes* Editorial Board now consists of 712 members, representing a team of worldwide experts in diabetes mellitus. They are from 56 countries, including Argentina (2), Australia (27), Austria (11), Belgium (5), Brazil (13), Canada (25), Chile (3), China (40), Cuba (1), Czech Republic (3), Denmark (16), Egypt (3), Finland (5), France (12), Germany (27), Greece (17), Hungary (4), India (28), Iran (8), Iraq (2), Ireland (3), Israel (10), Italy (56), Japan (30), Jordan (1), Kuwait (3), Lebanon (1), Malaysia (1), Malta (1), Mexico (4), Netherlands (9), New Zealand (3), Nigeria (2), Norway (2), Oman (3), Pakistan (2), Poland (7), Portugal (1), Qatar (1), Romania (2), Saudi Arabia (1), Singapore (4), Slovakia (1), South Africa (1), South Korea (15), Spain (24), Sweden (5), Switzerland (4), Thailand (4), Tunisia (1), Turkey (13), United Arab Emirates (3), United Kingdom (27), United States (213), Venezuela (1), and Yemen (1).

EDITOR-IN-CHIEF

Lu Qi, *Boston*
Jingbo Zhao, *Aalborg*

STRATEGY ASSOCIATE EDITOR-IN-CHIEF

Undurti Narasimha Das, *Shaker Heights*
Min Du, *Laramie*
Gregory I Liou, *Augusta*
Zhong-Cheng Luo, *Quebec*
Demosthenes B Panagiotakos, *Athens*

GUEST EDITORIAL BOARD MEMBERS

Juei-Tang Cheng, *Tainan*
Chih-Hsung Chu, *Kaohsiung*
Low-Tone (Larry) Ho, *Taipei*
Cheng-Cheng Hsiao, *Keelung*
Yung-Hsi Kao, *Taoyuan*
Chi Feng Liu, *Taipei*
Shing-Hwa Liu, *Taipei*
Wayne H-H Sheu, *Taichung*
Eing-Mei Tsai, *Kaohsiung*
Chin-Hsiao Tseng, *Taipei*
Yen Tzung-Hai, *Taipei*
Ching-Shuang Wu, *Kaohsiung*
Wei-Chung Vivian Yang, *Taipei*
Wen-Chin Yang, *Taipei*

MEMBERS OF THE EDITORIAL BOARD



Argentina

Justo P Castaño, *Cordoba*
Eduardo Spinedi, *La Plata*



Australia

Sof Andrikopoulos, *Heidelberg Heights*
Hugh Russell Barrett, *Perth*
Bernhard T Baune, *Townsville*
Grant Brinkworth, *Adelaide*
Louise Janet Maple Brown, *Casuarina*
Melinda Therese Coughlan, *Melbourne*
Josephine Maree Forbes, *Melbourne*
Paul A Fournier, *Perth*
Angela Gialamas, *Adelaide*
Mark Douglas Gorrell, *Newtown*
Graeme Hankey, *Perth*
Anandwardhan A Hardikar, *Melbourne*
Michael Horowitz, *Adelaide*
Karin Jandeleit-Dahm, *Melbourne*
Martha Lappas, *Victoria*
Peter J Little, *Melbourne*
Xin Liu, *Brisbane*
Dianna Josephine Magliano, *Caulfield*
Robyn McDermott, *Adelaide*
Beverly Sara Muhlhausler, *Adelaide*
Christopher Nolan, *Canberra*
Luciano Pirola, *Melbourne*
Maryam Rashidi, *Victoria*
Karly Calliopi Sourris, *Victoria*
Greg Tesch, *Clayton*
Jack Ronald Wall, *Penrith*
Owen Llewellyn Woodman, *Bundoora*



Austria

Christian Heinz Anderwald, *Vienna*
Helmuth Martin Borkenstein, *Graz*
Walter Hermann Hörl, *Vienna*
Alexandra Kautzky-Willer, *Vienna*

Friedrich Mittermayer, *Vienna*
Markus Paulmichl, *Salzburg*
Stefan Pilz, *Graz*
Guntram Schernthaner, *Vienna*
Harald Sourij, *Graz*
Thomas Michael Stulnig, *Vienna*
Ludwig Wagner, *Vienna*



Belgium

Giovanni Dapri, *Brussels*
Christophe De Block, *Antwerp*
Ekaterine Tskitishvili, *Liege*
F Andre Van Assche, *Leuven*
Luc F Van Gaal, *Antwerp*



Brazil

Monica Levy Andersen, *Vila Clementino*
Claudia RL Cardoso, *Rio de Janeiro*
Ricardo Vitor Cohen, *São Paulo*
Marcelo Correia, *Rio de Janeiro*
Cassyano Januario Correr, *Curitiba*
Matheus Roriz Cruz, *Porto Alegre*
Cintia Chaves Curioni, *Rio de Janeiro*
Freddy Goldberg Eliaschewitz, *Rua Goiás*
Rodrigo Jorge, *Ribeirão Preto*
Luciana Ansaneli Naves, *Asa Norte*
Júlio César Voltarelli, *Ribeirão Preto*
Bernardo L Wajchenberg, *Pinheiros*
Jacqueline Nelisis Zanoní, *Maringá*



Canada

Jean-Luc Ardilouze, *Sherbrooke*

Subrata Chakrabarti, *London*
 David Cherney, *Ontario*
 Mervyn Deitel, *Toronto*
 Jean-Pierre Després, *Quebec*
 David Joseph Hill, *London*
 Tian-Ru Jin, *Toronto*
 Arulmozhi D Kandasamy, *Edmonton*
 Jennifer L Kuk, *Toronto*
 Ismail Laher, *Vancouver*
 Roger S McIntyre, *Toronto*
 David Meyre, *Ontario*
 Joseph Fomusi Ndisang, *Saskatoon*
 Raj Padwal, *Alberta*
 Ciriaco A Piccirillo, *Montreal*
 Remi Rabasa-Lhoret, *Montreal*
 AM James Shapiro, *Edmonton*
 Guang Sun, *St. John's*
 Valerie Taylor, *Hamilton*
 Cory Toth, *Calgary*
 André Tremblay, *Montréal*
 Vincent C Woo, *Winnipeg*
 James Roscoe Wright, *Calgary*
 Xi-Long Zheng, *Calgary*



Chile

Sebastian San Martin, *Valparaiso*
 Armando Rojas-Rubio, *Talca*
 Luis Sobrevia, *Santiago*



China

Pang-Zeng Chang, *Qingdao*
 Jie Chen, *Nanjing*
 Bernard Man Yung Cheung, *Hong Kong*
 William Chi-shing Cho, *Hong Kong*
 Tian-Pei Hong, *Beijing*
 Qin Huang, *Shanghai*
 Po Sing Leung, *Hong Kong*
 Chao Liu, *Nanjing*
 Jian-Kang Liu, *Xi'an*
 Lie-Gang Liu, *Wuhan*
 Ronald Ching Wan Ma, *Hong Kong*
 Jin-Sheng Qi, *Shijiazhuang*
 Wing Yee So, *Hong Kong*
 Cheuk Chun Szeto, *Hong Kong*
 Kathryn Tan, *Hong Kong*
 Cheng-Ming Wang, *Yangzhou*
 Cong-Yi Wang, *Wuhan*
 Yu Wang, *Hong Kong*
 Guang-Da Xiang, *Wuhan*
 Bao-Feng Yang, *Harbin*
 Shu-Yu Yang, *Fujian*
 Xi-Lin Yang, *Hong Kong*
 Zai-Qing Yang, *Wuhan*
 Shan-Dong Ye, *Hefei*
 Shi-Sheng Zhou, *Dalian*
 Zhi-Guang Zhou, *Changsha*



Cuba

Luis Sarmiento-Pérez, *Havana*



Czech Republic

Martin Haluzik, *Prague*

Michal Krcma, *Plzen*
 Terezie Pelikanova, *Prague*



Denmark

Charlotte Brøns, *Gentofte*
 Jens Sandahl Christiansen, *Arhus*
 Flemming Dela, *Copenhagen*
 Kristine Færch, *Gentofte*
 Erik L Grove, *Aarhus*
 Louise Groth Grunnet, *Gentofte*
 R Scott Heller, *Gentofte*
 Kurt Højlund, *Odense C*
 Filip K Knop, *Hellerup*
 Helle Markholst, *Måløv*
 Jens D Mikkelsen, *Copenhagen*
 Ole Hartvig Mortensen, *Copenhagen*
 Oluf Pedersen, *Copenhagen*
 Esben Thyssen Vestergaard, *Aarhus*
 Milan Zdravkovic, *Søborg*



Egypt

Mamdouh Moawad Ali Hssan, *Cairo*
 Moshira Abdel Hakim Rateb, *Cairo*
 Mona Farag Schaalán, *Cairo*



Finland

Siamak Bidel, *Helsinki*
 Gang Hu, *Helsinki*
 Thomas Kietzmann, *Oulu*
 Qing Qiao, *Helsinki*
 Karoliina Wehkalampi, *Helsinki*



France

Jean Claude Ansquer, *Dijon*
 Bertrand Cariou, *Nantes*
 Sylvie Dejager, *Rueil-Malmaison*
 Naim Akhtar Khan, *Dijon*
 Jean-Philippe Lavigne, *Nîmes*
 Michel Marre, *Paris*
 Marie-Claude Morice, *Massy*
 Riccardo Perfetti, *Paris*
 Gérard Said, *Paris*
 Sophie Visvikis Siest, *Nancy*
 Dominique Simon, *Paris*
 Didier Vieau, *Villeneuve d'Ascq*



Germany

Ioanna Gouni Berthold, *Cologne*
 Christa Buechler, *Regensburg*
 Roland Büttner, *Heidelberg*
 Michael Froehner, *Dresden*
 Hammes Hans-Peter, *Mannheim*
 Nadj Herbach, *Munich*
 Andrea Icks, *Düsseldorf*
 Thomas Jax, *Neuss*
 Ulrich Arthur Julius, *Dresden*
 Michael Kluge, *Munich*
 Florian Lang, *Tuebingen*
 Matthias Laudes, *Köln*
 Ralf Lobmann, *Stuttgart*

Rafael T Mikolajczyk, *Bremen*
 Andreas Stefan Mueller, *Halle (Saale)*
 Karsten Müssig, *Tübingen*
 Nahid Parvizi, *Neustadt am Rübenberge*
 Thomas Peter Reinehr, *Datteln*
 Michael Ristow, *Jena*
 Sven Schinner, *Duesseldorf*
 Peter Egbert Hermann Schwarz, *Dresden*
 Konstantinos Stellos, *Tubingen*
 Ovidiu Alin Stirban, *Bad Oeynhausen*
 Diego J Walther, *Berlin*
 Silvia Anette Wein, *Kiel*
 Christian Wrede, *Berlin*
 Dan Ziegler, *Düsseldorf*



Greece

George P Chrousos, *Athens*
 Moses S Elisaf, *Ioannina*
 Panagiotis Georgoulis, *Larissa*
 Nikolaos Kadoglou, *Thessaloniki*
 Gerasimos E Krassas, *Krini*
 Spilios Manolakopoulos, *Attiki*
 Nikolaos Papanas, *Alexandroupolis*
 Dimitrios Papazoglou, *Alexandroupolis*
 Sokratis Pastromas, *Athens*
 Melpomeni Peppas, *Athens*
 Christina Piperi, *Goudi*
 Nicholas K Tentolouris, *Athens*
 Konstantinos A Toulis, *Salonika*
 Apostolos Tsapas, *Thessaloniki*
 Konstantinos Tziomalos, *Thessaloniki*
 Elias Zintzaras, *Thessaly*



Hungary

Mária Bagyánszki, *Szeged*
 György Jermendy, *Budapest*
 Karoly Racz, *Budapest*
 Gyula Soltesz, *Pécs*



India

Deepak Narayan Amrapurkar, *Mumbai*
 C V Anuradha, *Tamil Nadu*
 Sarika Arora, *New Delhi*
 Pitchai Balakumar, *Sivakasi*
 Muthuswamy Balasubramanyam, *Chennai*
 Subhabrata Chakrabarti, *Hyderabad*
 Abhay Sankar Chakraborti, *Kolkata*
 Tapan K Chaudhuri, *New Delhi*
 Kanwaljit Chopra, *Chandigarh*
 Malabika Datta, *Delhi*
 Debidas Ghosh, *West Bengal*
 Ravinder Goswami, *New Delhi*
 Pappachan M Joseph, *Kerala*
 Jothydev Kesavadev, *Kerala*
 KVS Hari Kumar, *Lucknow*
 Anoop Misra, *New Delhi*
 Analava Mitra, *Kharagpur*
 Viswanathan Mohan, *Chennai*
 S P Murthy, *Bangalore*
 Pallavi Panchu, *Guntur*
 Usharani Pingali, *Hyderabad*
 Ambady Ramachandran, *Egmore Chennai*
 Vadde Ramakrishna, *Kadapa*

Geetha Vani Rayasam, *Haryana*
Rajat Sandhir, *Chandigarh*
Manju Sharma, *New Delhi*
Suman Bala Sharma, *Delhi*
Tarun Sharma, *Chennai*



Iran

Mohammad Abdollahi, *Tehran*
Mohammad Kazemi Arababadi, *Rafsanjan*
Leila Azadbakht, *Isfahan*
Hamid Baradaran, *Tehran*
Behrooz Broumand, *Tehran*
Ahmad Esmailzadeh, *Isfahan*
Majid Ghayour-Mobarhan, *Mashhad*
Mohsen Janghorbani, *Isfahan*



Iraq

Saad Abdul-Rahman Hussain, *Baghdad*
Abbas Ali Mansour, *Basrah*



Ireland

Amar Agha, *Dublin*
Mark Philip Hehir, *Dublin*
Gerald H Tomkin, *Dublin*



Israel

Michael Aviram, *Haifa*
Gal Dubnov-Raz, *Tel Hashomer*
Shimon Efrat, *Tel Aviv*
Raymond Elias Farah, *Safed*
Oren Froy, *Rehovot*
Saher Hamed, *Haifa*
Arid Nakhoul, *Haifa*
Orit Pinhas-Hamiel, *Tel Hashomer*
Haim Werner, *Tel Aviv*
Marina Shargorodsky Zimlichman, *Holon*



Italy

Luigi Angrisani, *Napoli*
Moschetta Antonio, *Bari*
Antonio Aversa, *Rome*
Roberto Baldelli, *Rome*
Giuseppe Barbaro, *Rome*
Alessandro Bartolomucci, *Parma*
Giuseppina Basta, *Pisa*
Simona Bertoli, *Milano*
Federico Bilotta, *Rome*
Fabio Broglio, *Torino*
Francesco G Chiarelli, *Chieti*
Sergio Coccheri, *Bologna*
Massimo Collino, *Torino*
Marco Aristide Comaschi, *Genoa*
Renzo Cordera, *Genova*
Francesco Dotta, *Siena*
Gagliardini Elena, *Bergamo*
Stefano Fiorucci, *Perugia*
Maurizio Galderisi, *Naples*
Amalia Gastaldelli, *Pisa*

Ezio Ghigo, *Turin*
Carla Giordano, *Palermo*
Paolo Gisondi, *Verona*
Riccarda Granata, *Turin*
Giorgio Iervasi, *Pisa*
Claudia Kusmic, *Pisa*
Carmelo La Rosa, *Catania*
Francesco Landi, *Rome*
Monica Rosa Loizzo, *Arcavacata Rende*
Paolo Magni, *Milano*
Mariano Malaguarnera, *Catania*
Melania Manco, *Rome*
Piero Marchetti, *Pisa*
Massimo Massi-Benedetti, *Perugia*
Antonio Nicolucci, *Imbaro*
Lucia Pacifico, *Rome*
Stefano Palomba, *Catanzaro*
Giampaolo Papi, *Carpi*
Renato Pasquali, *Bologna*
Piermarco Piatti, *Milano*
Dario Pitocco, *Rome*
Antonio E Pontiroli, *Milano*
Giulio Marchesini Reggiani, *Bergamo*
Giuseppe Remuzzi, *Bergamo*
Manfredi Rizzo, *Palermo*
Raffaella Rosso, *Genoa*
Giuseppe Schillaci, *Perugia*
Leonardo A Sechi, *Sassari*
Imad Sheiban, *Torino*
Cesare R Sirtori, *Milano*
Giovanni Tarantino, *Naples*
Giovanni Targher, *Verona*
Donadon Valter, *Pordenone*
Alberto Verrotti, *Chieti*
Andrea Viggiano, *Napoli*
Gianvincenzo Zuccotti, *Milan*



Japan

Masato Asahina, *Chiba*
Takuya Awata, *Saitama*
Yuichiro Eguchi, *Saga*
Goji Hasegawa, *Kyoto*
Satoshi Inoue, *Tokyo*
Eiji Ishimura, *Osaka*
Masayuki Iwano, *Nara*
Takashi Kadowaki, *Tokyo*
Eisuke Kagawa, *Hiroshima*
Masahito Katahira, *Aichi*
Eiji Kawasaki, *Nagasaki*
Noriyuki Koibuchi, *Gunma*
Kazuhiko Kotani, *Tochigi*
Daisuke Koya, *Ishikawa*
Norikazu Maeda, *Osaka*
Takayuki Masaki, *Oita*
Yuji Matsuzawa, *Osaka*
Kazuaki Nishio, *Tokyo*
Kenji Okumura, *Nagoya*
Motoaki Saito, *Yonago*
Toshiyasu Sasaoka, *Toyama*
Michio Shimabukuro, *Okinawa*
Kohzo Takebayashi, *Saitama*
Hiroyuki Tamemoto, *Tochigi*
Takashi Togo, *Yokohama*
Jun Udagawa, *Izumo*
Yoshinari Uehara, *Fukuoka*
Takuya Watanabe, *Tokyo*
Toshihiko Yada, *Tochigi*

Tohru Yorifuji, *Osaka*



Jordan

Yousef S Khader, *Irbid*



Kuwait

Kamal AA Sulaiman Al-Shoumer, *Kuwait*
Ibrahim Fadel Benter, *Safat*
Abu Salim Mustafa, *Kuwait*



Lebanon

Ramzi F Sabra, *Beirut*



Malaysia

Mafauzy Mohamed, *Kota Bharu*



Malta

Charles Savona-Ventura, *Msida*



Mexico

Manuel González-Ortiz, *Guadalajara*
Fernando Guerrero-Romero, *Durango*
Jesus Alberto Olivares-Reyes, *Mexico City*
Rocío Salceda, *Mexico City*



Netherlands

Sander Kersten, *Wageningen*
Nanne Kleefstra, *Zwolle*
Edwin Mariman, *Maastricht*
Don Poldermans, *Rotterdam*
François Pouwer, *Tilburg*
Han Roelofsen, *Groningen*
Hendrik-Jan Schuurman, *Utrecht*
Suat Simsek, *Alkmaar*
Marcel Twickler, *Bergen op Zoom*



New Zealand

Paul Hofman, *Auckland*
Peter E Lobie, *Auckland*
Elaine Rush, *Auckland*



Nigeria

Adejuwon A Adeneye, *Lagos*
Anthonia Okeoghene Ogbera, *Lagos*



Norway

Akhtar Hussain, *Oslo*
Odd Erik Johansen, *Hovik*

**Oman**

Mohammed Al Shafae, *Muscat*
Jumana S Saleh, *Muscat*
Radha Shenoy, *Muscat*

**Pakistan**

Shahid Hameed, *Islamabad*
Jamil A Malik, *Islamabad*

**Poland**

Marcin Baranowski, *Bialystok*
Jerzy Beltowski, *Lublin*
Alicia Hubalewska Dydejczyk, *Krakow*
Maciej Owecki, *Poznań*
Ewa Pankowska, *Warsaw*
Agnieszka Piwowar, *Wroclaw*
Dorota Anna Zieba, *Krakow*

**Portugal**

M Graça Pereira, *Braga*

**Qatar**

Hong Ding, *Doha*

**Romania**

Elena Ganea, *Bucharest*
Adriana Georgescu, *Bucharest*

**Saudi Arabia**

J Fernando Arevalo, *Caracas*

**Singapore**

S Thameem Dheen, *Singapore*
Yung Seng Lee, *Singapore*
Daniel Ng, *Singapore*
Rob Martinus van Dam, *Singapore*

**Slovakia**

Katarína Šebeková, *Bratislava*

**South Africa**

Md Shahidul Islam, *Durban*

**South Korea**

Huneg-Sik Choi, *Gwangju*
Kyung Mook Choi, *Seoul*
Won Mi Hwang, *Seoul*
Eui-Bae Jeung, *Chungbuk*

Ju-Hee Kang, *Incheon*
Sin Gon Kim, *Seongbuk-Gu*
Sung-Jin Kim, *Seoul*
Young-Gyu Ko, *Seoul*
Kang-Beom Kwon, *Chonbuk*
Myung Gull Lee, *Bucheon*
Soo Lim, *Seongnam*
Byung-Hyun Park, *Jeonbuk*
Seungjoon Park, *Seoul*
Kun-Ho Yoon, *Seoul*
Jeesuk Yu, *Cheonan*

**Spain**

Vivencio Barrios, *Madrid*
M Lusia Bonet, *Palma de Mallorca*
Manuel Vazquez Carrera, *Barcelona*
Maria Luz Martinez Chantar, *Derio*
Manuel Aguilar Diosdado, *Cádiz*
Javier Espino, *Badajoz*
Ricardo V García-Mayor, *Vigo*
José Manuel Gómez-Sáez, *Barcelona*
Oreste Gualillo, *Santiago de Compostela*
J Alfredo Martínez Hernández, *Pamplona*
Emilio Herrera, *Madrid*
Amelia Marti, *Pamplona*
Merce Miranda, *Tarragona*
JF Navarro-González, *Santa Cruz de Tenerife*
Alberto Ortiz, *Madrid*
Maria Javier Ramirez, *Pamplona*
Eugenia Resmini, *Barcelona*
Pedro Romero-Aroca, *Reus*
Jordi Salas-Salvadó, *Reus*
Gines M Salido, *Caceres*
Victor Sanchez-Margalet, *Seville*
Helmut Schröder, *Barcelona*
Carmen Segundo, *Cádiz*
Rafael Simó, *Barcelona*

**Sweden**

Joanna Hlebowicz, *Malmö*
Kaj S Stenlöf, *Göteborg*
Ann-Britt Wirén, *Linköping*
Weili Xu, *Stockholm*
Shao-Nian Yang, *Stockholm*

**Switzerland**

Kaspar Berneis, *Zurich*
Pascal Bovet, *Lausanne*
Luc Tappy, *Lausanne*
Christian Toso, *Geneva*

**Thailand**

Narattaphol Charoenphandhu, *Bangkok*
Arthorn Riewpaiboon, *Bangkok*
Rawee Teanpaisan, *Hat-Yai*
Viroj Wiwanitkit, *Bangkok*

**Tunisia**

Khaled Hamden, *Sfax*

**Turkey**

Ugur Cavlak, *Denizli*
Teoman Dogru, *Ankara*
Ersin Fadillioğlu, *Ankara*
Abdurrahman Fatih Fidan, *Afyonkarahisar*
Muammer Karadeniz, *Bornova-Izmir*
Cevdet Kaya, *Istanbul*
Fahrettin Kelestimur, *Kayseri*
Altan Onat, *Istanbul*
Semir Ozdemir, *Antalya*
Mustafa Şahin, *Ankara*
Ilker Tasci, *Ankara*
Belma Turan, *Ankara*
Serap Yalin, *Mersin*

**United Arab Emirates**

Ernest Akingunola Adeghate, *Al Ain*
Mukesh M Agarwal, *Al Ain*
Samir M Awadallah, *Sharjah*

**United Kingdom**

Nisreen Alwan, *Leeds*
Ambika P Ashraf, *Birmingham*
Chen Bing, *Liverpool*
Fay Crawford, *Edinburgh*
Tim M Curtis, *Belfast*
Umesh Dashora, *Hastings*
Gareth Davison, *Belfast*
Peter Raymond Flatt, *Coleraine*
Kathleen M Gillespie, *Bristol*
Peter John Grant, *Leeds*
Lorna W Harries, *Exeter*
Nigel Hoggard, *Aberdeen*
Nigel Irwin, *Coleraine*
Edward Jude, *Lancashire*
Andreas F Kolb, *Aberdeen*
Stefan Marciniak, *Cambridge*
Moffat Joha Nyirenda, *Edinburgh*
Jeetesh Patel, *Birmingham*
Snorri Bjorn Rafnsson, *Edinburgh*
Thozhukat Sathyapalan, *Yorkshire*
Latika Sibal, *Newcastle*
Rajagopalan Sriraman, *Lincoln*
Ramasamyiyer Swaminathan, *London*
Abd A Tahrani, *Birmingham*
G Neil Thomas, *Birmingham*
Cecil Thompson, *London*
Paul Henry Whiting, *Leicester*

**United States**

Varun Agrawal, *Springfield*
Mohamed Al-Shabrawey, *Augusta*
Pascale Alard, *Louisville*
Omar Ali, *Milwaukee*
Judith Aponte, *New York*
Balamurugan N Appakalai, *Minneapolis*
Hwyda A Arafat, *Philadelphia*
Carl V Asche, *Salt Lake*
Sanford A Asher, *Pittsburgh*
Anthony Atala, *Winston-Salem*
Sami Toufic Azar, *Beirut*

George Louis Bakris, *Chicago*
Alistair J Barber, *Hershey*
Daniel C Battle, *Chicago*
David SH Bell, *Birmingham*
Rita Bortell, *Worcester*
Sebastien G Bouret, *Los Angeles*
David Lloyd Brown, *Stony Brook*
Lu Cai, *Louisville*
Jack D Caldwell, *Erie*
Anna C Calkin, *Los Angeles*
Roberto A Calle, *Groton*
R Keith Campbell, *Pullman*
Carlos Campos, *New Braunfels*
Heping Cao, *New Orleans*
Krista Casazza, *Birmingham*
Aaron Brandon Caughey, *Portland*
Eileen R Chasens, *Pittsburgh*
Munmun Chattopadhyay, *Ann Arbor*
Xiao-Li Chen, *St Paul*
Sheri Renee Colberg, *Norfolk*
Craig Ian Coleman, *Hartford*
Robert Russell Conley, *Indianapolis*
Colleen M Croniger, *Cleveland*
Doyle M Cummings, *Greenville*
William C Cushman, *Memphis*
Patricia Ann D'Amore, *Boston*
Patricia Darbishire, *West Lafayette*
Guillaume Darrasse-Jèze, *New York*
Ravi M Dasu, *Sacramento*
Michael Harvey Davidson, *Chicago*
Prakash Deedwania, *San Francisco*
Hong-Wen Deng, *Kansas City*
Teresa P DiLorenzo, *Bronx*
Scot E Dowd, *Lubbock*
Samuel C Durso, *Baltimore*
Krystal L Edwards, *Dallas*
Alexander M Efanov, *Indianapolis*
Azza B El-Remessy, *Augusta*
Amy Zhihong Fan, *Atlanta*
Melissa Spezia Faulkner, *Tucson*
George S Ferzli, *Staten Island*
Paolo Fiorina, *Boston*
James Edward Foley, *East Hanover*
Samuel N Forjuoh, *Temple*
Alessia Fornoni, *Miami*
Martha M Funnell, *Ann Arbor*
Trudy Gaillard, *Columbus*
Pietro Galassetti, *Irvine*
Claudia Gragnoli, *Hershey*
Jennifer B Green, *Durham*
Gary J Grover, *Piscataway*
Alok Kumar Gupta, *Baton Rouge*
Werner K Gurr, *New Haven*
Samy L Habib, *San Antonio*
Abdel Rahim Hamad, *Baltimore*
Daniel M Herron, *New York*
Tiffany Hilton, *Rochester*
Raimund Hirschberg, *Torrance*
Michael Francis Holick, *Boston*
Zhaoyong Hu, *Houston*
Rachel Mary Hudacko, *New Brunswick*
Yasuo Ido, *Boston*
Brian K Irons, *Lubbock*
Pamela Itkin-Ansari, *La Jolla*
Hieronim Jakubowski, *Newark*
Hong-Lin Jiang, *Blacksburg*
Ping Jiao, *Providence*
Shengkan Jin, *Piscataway*
Arpita Kalla, *St Louis*
Richard Evers Katholi, *Springfield*

Melina Rae Kibbe, *Chicago*
Bhumsoo Kim, *Ann Arbor*
Tomoshige Kino, *Bethesda*
Julienne K Kirk, *Winston-Salem*
Renu A Kowluru, *Detroit*
Lewis H Kuller, *Pittsburgh*
Rajesh Kumar, *Temple*
Blandine Laferrère, *New York*
Sang Yeoup Lee, *Mayo*
Cong-Jun Li, *Beltsville*
Ching-Shwun Lin, *San Francisco*
Julie Lin, *Boston*
Shuo Lin, *Los Angeles*
Peter Lindgren, *San Diego*
James F List, *Princeton*
Dong-Min Liu, *Blacksburg*
Zhen-Qi Liu, *Charlottesville*
George William Lysterly, *Conway*
Jian-Xing Ma, *Oklahoma City*
Rong Ma, *Fort Worth*
Xin-Laing Ma, *Philadelphia*
David Maggs, *San Diego*
Kenneth Maiese, *Detroit*
Kevin C Maki, *Glen Ellyn*
Sridhar Mani, *Bronx*
Suresh Mathews, *Auburn*
Lauraar McCabe, *East Lansing*
Sarah E Messiah, *Miami*
Thomas O Metz, *Richland*
Shannon A Miller, *Orlando*
Murielle Mimeault, *Omaha*
Raghavendra G Mirmira, *Indianapolis*
Prasun J Mishra, *Bethesda*
Reema Mody, *Grayslake*
Arshag D Mooradian, *Jacksonville*
Mohammad Reza Movahed, *Tucson*
James Mu, *Rahway*
Muraleedharan G Nair, *East Lansing*
Manuel F Navedo, *Seattle*
Charles B Nemeroff, *Atlanta*
Joshua J Neumiller, *Spokane*
Steven Nissen, *Cleveland*
Hirofumi Noguchi, *Fort Worth*
Craig Nunemake, *Charlottesville*
Patrick J O'Connor, *Minneapolis*
Erin St Onge, *Apopka*
Wei-Hong Pan, *Baton Rouge*
Naushira Pandya, *Fort Lauderdale*
Michael R Peluso, *Corvallis*
Inga Peter, *New York*
Axel Pflueger, *Rochester*
Gretchen A Piatt, *Pittsburgh*
John D Piette, *Ann Arbor*
Leonid Poretsky, *New York*
Walter J Pories, *Greenville*
Parviz M Pour, *Omaha*
Wei Qiao Qiu, *Boston*
Teresa Quattrin, *Buffalo*
Cristina Rabadán-Diehl, *Bethesda*
Rajendra S Raghov, *Memphis*
Swapnil Rajpathak, *Bronx*
Armin Rashidi, *Norfolk*
Mohammed S Razzaque, *Boston*
Beverly A S Reyes, *Philadelphia*
David Rodbard, *Potomac*
Helena W Rodbard, *Rockville*
June Hart Romeo, *Cleveland*
Raul J Rosenthal, *Fort Lauderdale*
Juan M Saavedra, *Bethesda*
Stephen W Schaffer, *Mobile*

Frank AJL Scheer, *Boston*
Richard E Scranton, *Tiverton*
Vallabh (Raj) O Shah, *Albuquerque*
Aziz Shaibani, *Houston*
Jin-Xiong She, *Augusta*
Guo-Ping Shi, *Boston*
Carol Ann Shively, *Winston-Salem*
Anders AF Sima, *Detroit*
Pramil N Singh, *Loma Linda*
Rajan Singh, *Los Angeles*
Jay S Skyler, *Miami*
Dawn Smiley, *Atlanta*
Matthew D Solomon, *Stanford*
Mark A Sperling, *Pittsburgh*
Rakesh K Srivastava, *Tyler*
Bangyan Stiles, *Los Angeles*
Yu-Xiang Sun, *Houston*
Salim Surani, *Corpus Christi*
Arthur L M Swislocki, *Martinez*
Ya-Xiong Tao, *Auburn*
John A Tayek, *Torrance*
John Gaylord Teeter, *New Haven*
Carlos Marcelo Telleria, *Vermillion*
Christopher Gordon Thanos, *Providence*
Ronald G Tilton, *Galveston*
Serena Tonstad, *Loma Linda*
Michael Lawrence Traub, *Staten Island*
Guillermo E Umpierrez, *Atlanta*
Margrit Urbanek, *Chicago*
Vladimir N Uversky, *Indianapolis*
Gabriel I Uwaifo, *Baton Rouge*
Volker Vallon, *San Diego*
Shambhu D Varma, *Baltimore*
Maria Virella, *Charleston*
Hong-Jun Wang, *Boston*
Mark E Williams, *Boston*
Nathan D Wong, *Irvine*
Guangyu Wu, *New Orleans*
Zhong-Jian Xie, *San Francisco*
Ming-Zhao Xing, *Baltimore*
Hariom Yadav, *Bethesda*
Lijun Yang, *Gainesville*
Ruoqing Yang, *Rahway*
Subhashini Yaturu, *Albany*
Joseph Yeboah, *Charlottesville*
Dengping Yin, *Nashville*
Yisang Yoon, *Rochester*
Yi-Hao Yu, *New York*
Kevin CJ Yuen, *Portland*
Ian Stuart Zagon, *Hershey*
Robert Yuk-Lung Zee, *Boston*
Cui-Lin Zhang, *Rockville*
James Xuejie Zhang, *Richmond*
Sarah Zhang, *Oklahoma*
Guixiang Zhao, *Atlanta*
Yang Zhao, *Indianapolis*
Ming-Hui Zou, *Oklahoma City*



Venezuela

Fuad Lechin, *Caracas*



Yemen

Khaled Abdul-Aziz Ahmed, *Ibb*

**REVIEW**

- 1285** New perspectives on exploitation of incretin peptides for the treatment of diabetes and related disorders
Irwin N, Flatt PR
- 1296** Role of adiponectin and some other factors linking type 2 diabetes mellitus and obesity
Chakraborti CK

Contents

World Journal of Diabetes
Volume 6 Number 15 November 10, 2015

ABOUT COVER

Editorial Board Member of *World Journal of Diabetes*, Massimo Collino, PhD, Department of Anatomy, Pharmacology and Forensic Medicine, University of Turin, via P. Giuria 9, 10125 Torino, Italy

AIM AND SCOPE

World Journal of Diabetes (*World J Diabetes*, *WJD*, online ISSN 1948-9358, DOI: 10.4239), is a peer-reviewed open access academic journal that aims to guide clinical practice and improve diagnostic and therapeutic skills of clinicians.

WJD covers topics concerning α , β , δ and PP cells of the pancreatic islet, the effect of insulin and insulinresistance, pancreatic islet transplantation, adipose cells and obesity.

We encourage authors to submit their manuscripts to *WJD*. We will give priority to manuscripts that are supported by major national and international foundations and those that are of great clinical significance.

INDEXING/ ABSTRACTING

World Journal of Diabetes is now indexed in Thomson Reuters Web of Science Emerging Sources Citation Index, PubMed Central, PubMed, Digital Object Identifier, and Directory of Open Access Journals.

FLYLEAF

I-V Editorial Board

EDITORS FOR THIS ISSUE

Responsible Assistant Editor: *Xiang Li*
Responsible Electronic Editor: *Ya-Jing Lu*
Proofing Editor-in-Chief: *Lian-Sheng Ma*

Responsible Science Editor: *Shui Qiu*
Proofing Editorial Office Director: *Xiu-Xia Song*

NAME OF JOURNAL

World Journal of Diabetes

ISSN

ISSN 1948-9358 (online)

LAUNCH DATE

March 15, 2010

FREQUENCY

Biweekly

EDITORS-IN-CHIEF

Lu Qi, MD, PhD, Assistant Professor, Department of Nutrition, Harvard School of Public Health, 665 Huntington Ave., Boston, MA 02115, United States

Jingbo Zhao, PhD, Associate Professor, Aalborg Hospital Science and Innovation Centre, Aalborg Hospital, Aarhus University Hospital, Aalborg 9000, Denmark

EDITORIAL OFFICE

Jin-Lei Wang, Director

Xiu-Xia Song, Vice Director

World Journal of Diabetes

Room 903, Building D, Ocean International Center, No. 62 Dongsihuan Zhonglu, Chaoyang District, Beijing 100025, China

Telephone: +86-10-85381891

Fax: +86-10-85381893

E-mail: editorialoffice@wjgnet.com

Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>

<http://www.wjgnet.com>

PUBLISHER

Baishideng Publishing Group Inc

8226 Regency Drive,

Pleasanton, CA 94588, USA

Telephone: +1-925-223-8242

Fax: +1-925-223-8243

E-mail: bpgoffice@wjgnet.com

Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>

<http://www.wjgnet.com>

PUBLICATION DATE

November 10, 2015

COPYRIGHT

© 2015 Baishideng Publishing Group Inc. Articles published by this Open-Access journal are distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits use, distribution, and reproduction in any medium, provided the original work is properly cited, the use is non-commercial and is otherwise in compliance with the license.

SPECIAL STATEMENT

All articles published in journals owned by the Baishideng Publishing Group (BPG) represent the views and opinions of their authors, and not the views, opinions or policies of the BPG, except where otherwise explicitly indicated.

INSTRUCTIONS TO AUTHORS

Full instructions are available online at http://www.wjgnet.com/1948-9358/g_info_20100107165233.htm

ONLINE SUBMISSION

<http://www.wjgnet.com/esps/>

New perspectives on exploitation of incretin peptides for the treatment of diabetes and related disorders

Nigel Irwin, Peter R Flatt

Nigel Irwin, Peter R Flatt, SAAD Centre for Pharmacy and Diabetes, University of Ulster, Coleraine BT52 1SA, Northern Ireland, United Kingdom

Author contributions: All authors contributed to the manuscript.

Conflict-of-interest statement: No potential conflicts of interest relevant to this article were reported.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Correspondence to: Dr. Nigel Irwin, SAAD Centre for Pharmacy and Diabetes, University of Ulster, Cromore Road, Coleraine BT52 1SA, Northern Ireland, United Kingdom. n.irwin@ulster.ac.uk
 Telephone: +44-28-70324574
 Fax: +44-28-70323939

Received: May 20, 2015
 Peer-review started: May 21, 2015
 First decision: September 17, 2015
 Revised: September 25, 2015
 Accepted: October 20, 2015
 Article in press: October 27, 2015
 Published online: November 10, 2015

Abstract

The applicability of stable gut hormones for the treatment of obesity-related diabetes is now undisputable. This is based predominantly on prominent and sustained glucose-lowering actions, plus evidence that these peptides can augment insulin secretion and pancreatic islet function over time. This review highlights the therapeutic potential of glucagon-like peptide-1 (GLP-1), glucose-dependent insulintropic polypeptide (GIP), oxyntomodulin (OXM) and cholecystokinin (CCK) for obesity-related diabetes.

Stable GLP-1 mimetics have already been successfully adopted into the diabetic clinic, whereas GIP, CCK and OXM molecules offer promise as potential new classes of antidiabetic drugs. Moreover, recent studies have shown improved therapeutic effects following simultaneous modulation of multiple receptor signalling pathways by combination therapy or use of dual/triple agonist peptides. However, timing and composition of injections may be important to permit interludes of beta-cell rest. The review also addresses the possible perils of incretin based drugs for treatment of prediabetes. Finally, the unanticipated utility of stable gut peptides as effective treatments for complications of diabetes, bone disorders, cognitive impairment and cardiovascular dysfunction is considered.

Key words: Diabetes; Obesity; Incretin; Prediabetes; Gut hormones

© **The Author(s) 2015.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Stable gut hormones have well defined therapeutic actions for type 2 diabetes mellitus. In addition, simultaneous modulation of gut hormone receptors could increase therapeutic efficacy, but timing and receptor activation profile may be important. Finally, gut-derived peptides could possess benefits for bone disorders, cognitive impairment and cardiovascular dysfunction.

Irwin N, Flatt PR. New perspectives on exploitation of incretin peptides for the treatment of diabetes and related disorders. *World J Diabetes* 2015; 6(15): 1285-1295 Available from: URL: <http://www.wjgnet.com/1948-9358/full/v6/i15/1285.htm> DOI: <http://dx.doi.org/10.4239/wjd.v6.i15.1285>

INTRODUCTION

The human gastrointestinal tract (GIT) comprises the stomach, as well as the small (duodenum, jejunum

and ileum) and large (caecum, colon and rectum) intestines. Aside from nutrient digestion, absorption and assimilation, the GIT also has significant endocrine functions^[1]. To date, the most important endocrine function of the gut relates to evidence that intestinal derived peptides are fundamentally involved in post-prandial insulin release^[2]. This action is termed the “incretin effect”, and relates to the direct beta-cell insulin secretory effect of two hormones, namely glucagon-like peptide-1 (GLP-1) and glucose-dependent insulinotropic polypeptide (GIP) that are secreted from L- and K-cells, respectively (Figure 1)^[3]. A number of other enteric peptide hormones released in response to feeding also have a role in energy regulation and possibly insulin secretion, including cholecystokinin (CCK) and oxyntomodulin (OXM) (Figure 1)^[4,5]. However, only GLP-1 and GIP fulfil the criteria of a true incretin hormone that stimulates glucose-induced insulin secretion at physiological circulating concentrations^[3]. Despite the obvious potential of incretin and incretin-like peptides for the treatment of conditions such as diabetes and obesity, the extremely short biological half-life of these peptides, due to efficient enzymatic degradation and subsequent renal filtration, severely limits therapeutic applicability^[4,5]. However, interest in gut peptides has increased in recent years with knowledge that modified versions of these compounds, with vastly improved pharmacokinetic properties, have sustained beneficial physiological effects^[6].

GLP-1

The biological actions of GLP-1 are largely preserved in type 2 diabetes and pharmacological doses of the peptide evoke robust insulin-releasing and antihyperglycaemic effects^[7]. GLP-1 exerts its beta-cell effects through interaction with specific surface receptors that activate signal transduction pathways including the stimulation of intracellular cAMP mediated events^[8]. GLP-1 also promotes beta-cell proliferation and islet cell neogenesis as well as inhibiting beta-cell apoptosis and alpha-cell glucagon secretion^[8]. Notably, both GLP-1 and GIP expression and secretion has been described in islet alpha cells^[9,10]. Indeed, it is feasible that intra-islet, rather than gut derived, GLP-1 and GIP make a significant contribution to these direct beneficial islet effects^[11-13]. However, it should be noted that positive direct islets effects are still noted in rodents following prolonged exogenous delivery of stable GLP-1 mimetics^[8].

GLP-1 not only targets pancreatic islet cells, but imparts positive actions in terms of inhibition of gastric emptying, suppression of appetite and weight loss^[8]. Given this advantageous biological action profile, there are now several GLP-1 related enzyme-resistant, long-acting analogues available for clinical use in diabetes (Table 1), ranging from regimens that require twice daily injection to those that necessitate only once weekly administration^[14]. Development of

new GLP-1 mimetics, such as those conjugated to an antithrombin III-binding pentasaccharide, are also in the pipeline^[15]. Interestingly, a recent commentary highlights that differences in the structure and pharmacokinetics of currently available GLP-1 mimetics could significantly alter immunogenicity, CNS signalling and overall therapeutic effect^[16]. Thus, physicians may need to re-evaluate the most appropriate GLP-1 treatment strategy for each patient. Encouragingly however, GLP-1-R agonists demonstrate an efficacy approaching that of insulin treatment, but unlike insulin have the added benefits of promoting weight loss with minimal risk of hypoglycaemia^[17].

Despite the widespread use of GLP-1 mimetics (Table 1), there have been recent safety concerns regarding the ability of sustained GLP-1-R activation to cause pancreatitis, pancreatic and thyroid cancer, as well as glucagon-producing neuroendocrine tumours in man^[18,19]. As such, it is well recognised that pancreatitis is a risk factor for pancreatic cancer^[20]. However, a recent meta-analysis did not support increased risk of pancreatitis or cancer associated with GLP-1 therapy^[21]. Indeed, issues with poorly matched patient groups treated with incretin-based vs non-incretin-based medications and problems with specifically identifying glucagon-producing cells also calls into question the validity of these safety concerns^[22]. Thyroid cancer fears appear to stem largely from rodent studies^[23], and reduced expression of the GLP-1 receptors in human, as opposed to rodent, thyroid cells is the likely explanation for this^[24]. The most frequently reported side effect of GLP-1 therapy is dose-dependent and transient mild to moderate nausea, vomiting and diarrhoea^[16]. Thus, taken together the safety profile of GLP-1 based therapeutics is largely reassuring. However, pharmacovigilance with GLP-1 drugs is still required, especially in relation to patients with a history, or increased risk, of pancreatitis or thyroid cancer.

GLUCOSE-DEPENDENT INSULINOTROPIC POLYPEPTIDE

Although initially thought to play a role in impeding histamine induced gastric acid secretion^[25], the primary physiological role of GIP is now considered to be stimulation of postprandial insulin secretion^[13]. The insulinotropic action of GIP, mediated by specific receptors on the surface of pancreatic beta-cells, is initiated largely by intracellular cAMP generation (Figure 1) and subsequent Ca²⁺ ion influx leading to insulin granule exocytosis^[13]. An additional beneficial action of GIP involves enhanced survival of beta-cells, which is also mediated through cAMP dependent cell signaling pathways^[26,27]. GIP also acts as beta-cell growth factor by stimulating mitogen-activated protein kinase pathways^[28] and modulating K_{ATP} channel expression^[29]. Given this impressive bioactive profile at the level of the beta-cell, there has been significant interest in the potential for GIP-based pharmaceuticals as antidiabetic

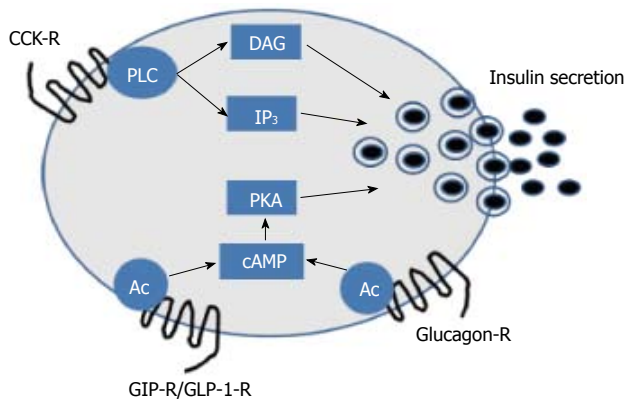


Figure 1 Schematic depicting the major signalling pathways involved in glucose-dependent insulinotropic polypeptide, glucagon-like peptide-1, glucagon and cholecystokinin induced insulin secretion from pancreatic beta-cells. AC: Adenyl cyclase; cAMP: Adenosine 3'-5'-cyclic monophosphate; DAG: Diacyl-glycerol; IP3: Inositol 1,4,5-trisphosphate; PKA: Protein kinase A; PLC: Phospholipase C; CCK: Cholecystokinin.

drugs. However, like GLP-1 the pharmacokinetic profile GIP is severely hindered due to rapid plasma degradation by the ubiquitous enzyme dipeptidyl peptidase 4 (DPP-4), and clearance cleared from the body by efficient renal filtration^[30]. In addition to this, the biological effects of GIP appear to be markedly reduced in patients with type 2 diabetes when compared to normal individuals^[7].

The first of these barriers has been conquered, as with GLP-1 mimetics, through generation of N-terminally modified enzyme-resistant, long-acting GIP molecules, and these molecules has been reviewed extensively elsewhere^[31,32]. However, the issue of reduced GIP responsiveness in type 2 diabetes still remains, and is thought to be linked to GIP receptor (GIP-R) down-regulation or desensitisation^[7]. However, it is highly likely that that GIP desensitisation is a pathophysiological consequence as opposed to an aetiological factor of type 2 diabetes. In keeping with this, studies correcting hyperglycaemia using insulin or sulphonylureas indicate that GIP sensitivity can be restored^[33,34]. It has also been demonstrated that a K-cell derived peptide co-secreted from the intestine with GIP, xenin-25, can potentiate the insulinotropic action of GIP^[35,36]. As such, a novel long-acting palmitate-derivatised analogue of xenin-25 was shown to significantly augment GIP action *in vitro* and *in vivo*^[37]. Moreover, sustained administration of this acylated xenin peptide exerted a spectrum of beneficial metabolic effects in high-fat-fed mice^[38]. This presumably relates to restoration of GIP action in these diabetic mice^[38]. In harmony with this, a recent study indicates that the impaired insulinotropic response to GIP under diabetic milieu involves mechanisms beyond simple expression of the GIP-R^[39], further highlighting a potential role for xenin. Therefore, there still appears to be significant, as yet untapped, therapeutic potential for GIP-based compounds, especially in combination with molecules that can enhance GIP sensitivity directly or counter hyperglycaemia through other actions.

Table 1 Incretin-based drugs currently approved by the European Medicines Agency

Drug name	Primary mechanism of action	EMA approval date
Exenatide	GLP-1 receptor agonist	Nov-06
Sitagliptin	DPP-4 inhibitor	Mar-07
Vildagliptin	DPP-4 inhibitor	Sep-07
Liraglutide	GLP-1 receptor agonist	Jun-09
Saxagliptin	DPP-4 inhibitor	Oct-09
Exenatide-LAR	GLP-1 receptor agonist	Jun-11
Linagliptin	DPP-4 inhibitor	Aug-11
Lixisenatide	GLP-1 receptor agonist	Feb-13
Alogliptin	DPP-4 inhibitor	Sep-13
Dulaglutide	GLP-1 receptor agonist	Jan-15

DPP-4: Dipeptidyl peptidase 4; GLP-1: Glucagon-like peptide-1; LAR: Long-acting release; EMA: European medicines agency.

OXYNTOMODULIN

Similar to GLP-1, OXM is an L-cell derived proglucagon gene product secreted in response to feeding^[40]. To date a specific OXM receptor has not been described, and the biological actions of OXM are attributed to binding and activation of GLP-1 and glucagon receptors (Figure 1), albeit with reduced potency compared to the native ligands^[41]. *In vitro* and *in vivo* rodent studies suggest that through glucagon receptor agonism, OXM induces catabolic effects that favour weight loss and subsequent improved metabolic control, while glucose homeostasis and insulin resistance are improved through activation of GLP-1 receptors^[5]. Promisingly, data from small clinical studies implies that beneficial effects on energy intake and weight loss also occur in humans^[42,43]. However, as is this case for the incretin hormones, the therapeutic potential of OXM-based molecules is hindered by rapid cleavage of the first two N-terminal amino acids of OXM by DPP-4 in plasma, rendering the peptide inactive^[44]. Nonetheless, structure-function studies show that N-terminal modification can protect against DPP-4 degradation without disproportionately affecting bioactivity of the molecule^[44,45]. Indeed, a recent study of six novel OXM analogues has revealed that Oxm-based peptides with specific N-terminal position 2 modifications are stable and show particular promise for the treatment of diabetes^[46]. These data suggest that further exploration of dual agonism of the GLP-1 and glucagon receptor is required for human diabetes. It is notable that co-administration of GLP-1 and glucagon in humans can replicate the beneficial actions of OXM^[47], although this approach may be more cumbersome in clinical practice.

CHOLECYSTOKININ

CCK is an intestinal I-cell derived gut hormone secreted in response to meal ingestion^[48]. CCK binds to specific CCK₁ receptors present on gastric mucosa and vagal afferent neurons which collectively leads to gallbladder secretions, release of pancreatic digestive juices, satiety and slowing

of gut motility^[1]. CCK₂ receptors are mainly confined to the gastrointestinal tract and brain and may have a role in regulating anxiety and locomotion^[49]. Importantly, CCK has also been shown to stimulate insulin secretion in rodents and man (Figure 1)^[50,51], and act as a growth and anti-apoptotic factor for pancreatic beta-cells^[52]. Thus, CCK agonists could have noteworthy potential for diabetes therapy, since their biological action profile is similar to the incretin hormones. However, native CCK is rapidly degraded by serum aminopeptidases upon secretion into the bloodstream^[53], which hinders therapeutic potential. However, early studies have clearly shown that both N-terminal modification through glycation, or PEGylation, can prevent enzymatic degradation of CCK and extend biological action and therapeutic potential^[53,54]. Following on from this, a more recently developed enzymatically stable, N-terminally modified, CCK analogue, namely (pGlu-Gln)-CCK-8, has been shown to have an improved pharmacodynamic profile, and to both alleviate and protect against obesity-related diabetes in animal models^[51,55], with an encouraging safety profile^[56]. The mechanism of action of (pGlu-Gln)-CCK-8 likely revolves around prominent and sustained reductions of energy intake, possibly related to modulation of central neuropeptide Y and melanocortin related pathways, and enhanced insulin release^[57]. Encouragingly, a PEGylated version of (pGlu-Gln)-CCK-8 has now been fully characterised, that would be resistant to kidney filtration, and suitable for possible once daily dosing in man^[58]. Further investigations relating to translation of beneficial effects to human type 2 diabetes together with safety evaluation are still required, but initial observations with specific and stable CCK₁ receptor agonists are encouraging.

MULTI-TARGET HYBRID PEPTIDE THERAPIES FOR DIABETES

Given the beneficial effects of OXM-based peptides, it follows that design of hybrid peptides capable of modulating more than one receptor pathway could have distinct therapeutic benefits for the treatment of obesity-related diabetes. By utilising the correct ratio of receptor pathway interactions, efficacy should be enhanced with the potential for administration of lower doses, thereby reducing, or removing, adverse side effects. The most logical starting point for design of a synthetic dual acting hybrid peptide would inevitably involve a modified incretin hormones capable of activating both GIP and GLP-1 receptors. As such, GIP/GLP-1 chimeric peptides were characterised almost 20 years ago, and the structural requirements for specific ligand-receptor interactions well defined^[59]. Combined administration of individual long-acting GIP and GLP-1 mimetics has been considered in preclinical studies, with some success^[60]. However, issues of separate drug formulation and dosing still remain, although these may not be insurmountable as indicated by recent

introduction of IDegLira for combined insulin and GLP-1 therapy in type 1 diabetes^[61]. In terms of a single hybrid peptide that can directly activate both GIP and GLP-1 receptors, only MAR701, Marcadia Biotech (now Roche) has progressed to the evaluation of beneficial effects in man. However, since the clinical benefits of DPP-4 inhibitors clearly involves increased circulating levels of both incretin peptides^[62], concomitant activation of GIP and GLP-1 receptors does appear to have promise for the treatment of type 2 diabetes (Table 1).

Further studies have investigated the effects of GLP-1 receptor agonism combined with either glucagon receptor agonism or antagonism^[63,64]. Although somewhat contradictory in nature, these contrasting regimens both utilise the beneficial glucose-lowering effects of GLP-1, combined with either inhibition of glucagon-mediated gluconeogenesis and glycogenolysis^[65], or activation of glucagon pathways involved in energy turnover and weight loss^[64], as is this case for OXM. Other modified hybrid peptides for dual activation of regulatory peptide receptors include, ZP3022, a combined GLP-1-gastrin agonist^[66]. Through activation of GLP-1 and CCK₂ receptors, this peptide improved glycaemic control in *db/db* mice *via* enhancement of beta-cell mass^[66]. However, perhaps more appealing is the potential for combined and sustained activation of GLP-1 and CCK₁ receptors. As such, two independent studies have clearly shown pronounced synergistic metabolic benefits with combined administration of long-acting GLP-1 and CCK₁ receptor agonists in rodent models of type 2 diabetes^[67,68]. These extremely positive effects are believed to occur through activation of complementary pathways that lead to significant weight loss and dramatically improved metabolic control^[67,68]. Furthermore, a novel CCK/GLP-1 hybrid peptide, based on the chemical structures of (pGlu-Gln)-CCK-8 and exenatide, has recently been described and shown to have significant therapeutic potential in high-fat fed mice^[69]. This molecule clearly warrants further study as a potential new treatment option for type 2 diabetes.

Considering the evident therapeutic efficacy offered by dual peptide receptor interactions, single compounds with the ability to concurrently activate three or more regulatory peptide receptors could deliver even greater beneficial effects. Moreover, the celebrated success of bariatric surgery for restoring metabolic control in type 2 diabetic patients, independent of weight loss^[70], results from a culmination of reduced energy intake and modulation of the secretion and biological action of numerous gut-derived peptides^[71]. Thus, there is now significant enthusiasm arising from designer modified peptides with the ability to concurrently modulate GIP, GLP-1 and glucagon receptor signalling^[72,73]. These triple-acting peptides have resulted in dramatic improvements in glucose homeostasis and overall metabolic control in high fat fed mice^[72,73]. Despite their obvious potential, issues regarding the ratio of GIP, GLP-1 and glucagon receptor activation still need to be addressed, As such,

a subsequent study has reported the distinct beneficial effects of a balanced glucagon, GLP-1 and GIP receptor tri-agonist to correct obesity and diabetes in high fat fed mice^[74]. Taken together, there is a clear and attractive rationale for further testing of combinatorial hormone therapies for the treatment of obesity and diabetes in humans.

Although the future trend for peptide-based anti-diabetic drugs seems to be development dual or triple agonists, treatment modalities that incorporate periods of beta-cell rest could be important for glycaemic control^[75]. Thus, antidiabetic drugs that induce direct beta-cell stimulatory effects can erode beta-cell mass over time^[76]. As such, intermittent periods of beta-cell rest may be useful to preserve long-term beta-cell function and lasting glycaemic control^[75]. In contrast to sulphonylureas and meglitinides, incretin based drugs stimulate insulin secretion in a glucose-dependent fashion that should help preserve beta-cell mass and function^[8]. Nonetheless, adequate periods of rest might still allow chronically stimulated pancreatic beta-cells to replenish both cell surface receptors and the immediately secretable insulin granule pool^[77]. Such effects, together with the positive actions of incretins on beta-cell stimulus-secretion coupling, survival and growth, could be highly beneficial. Accordingly, the timing of injections of dual or triple acting therapies, as well as the profile of receptor pathways activated, could be of valuable clinical relevance. In relation to this, inhibition of GIP-R signalling has been shown to improve metabolic control and glycaemic status in animal models of obesity-related diabetes by enhancing insulin action and diminishing insulin secretion^[78,79]. Thus a key aspect underlying the beneficial effects could be related to the induction of pancreatic beta-cell rest. Consistent with this, combination of morning injection of liraglutide, with stable GIP antagonist peptide in the evening, greatly improved glycaemic control in *db/db* mice compared with reciprocal administration or twice daily injection of liraglutide^[80]. Further investigation of this potentially important treatment paradigm, in combination with other agents that stimulate and/or relieve beta cell insulin release, is required to fully explore therapeutic relevance and applicability.

INCRETIN THERAPIES AND PREDIABETES

Prediabetes describes to a situation where blood sugar is high, but not elevated sufficiently to classify as overt type 2 diabetes. However, the condition represents a high risk state for future development of diabetes, most likely linked to progressive beta-cell decline^[81]. Thus, it follows that the positive effects of incretin mimetics on beta-cell function, including possible benefits for beta-cell proliferation and survival, plus additional weight-lowering and extrapancreatic actions^[8], could hold significant promise for prediabetic patients. Moreover, patients with prediabetes have been shown to have

an impaired incretin effect in response to oral nutrient delivery^[82].

To date, there have been several tentative clinical studies conducted on the potential beneficial effects of incretin-based drugs for prediabetes. Studies with DPP-4 inhibitors (Table 1), which prevent incretin peptide degradation and increase active circulating levels of GIP and GLP-1, reported modest positive effects^[83-85]. However, treatment with the stable incretin mimetics, exenatide or liraglutide, generated more positive outcomes^[86,87]. This included significant reductions in the prevalence of prediabetes with reversion to normal glucose tolerance^[86,87]. The inconsistency between DPP-4 inhibitors and GLP-1 mimetics most likely relates to differences in the circulating levels of active hormones achieved. However, issues of oral vs injectable delivery of DPP-4 inhibitors and GLP-1 mimetics, respectively, could significantly affect compliance in this patient subgroup. In addition, the potential adverse side-effect profile of incretin based therapies, as discussed above, would also have to be fully considered. Finally, the cost of therapy with DPP-4 inhibitors and particularly GLP-1 mimetics is greater when compared to other glucose-lowering agents^[88]. Thus, given the limited experience to date regarding the effect of incretin therapies in prediabetes, future clinical trials would be recommended. In terms of GIP, CCK and OXM therapies, clinical effectiveness in type 2 diabetes would need to be fully established before beneficial actions in prediabetic patients could be considered.

UNEXPECTED THERAPEUTIC POTENTIAL OF INCRETIN BASED DRUGS

Bone

Although incretin hormones have been studied extensively for therapeutic effectiveness in diabetes, research has uncovered unexpected benefits in various other tissues. For instance, a role for gastrointestinal derived hormones in bone remodeling is suspected since serum levels of bone biomarkers rapidly alter after a meal^[89]. Indeed, functional GIP receptors have been evidenced on the surface of bone cells^[90]. Notably, GIP has been shown to inhibit bone resorption in humans under both euglycaemic and hyperglycaemic states^[91]. Thus, the beneficial effects of GIP on bone could be independent of feeding state. Indeed, exogenous prolonged administration of an N-terminally modified stable GIP receptor agonist imparted various beneficial effects on tissue-level bone material properties of rats^[92]. In terms of GLP-1 effects on bone, the picture is less clear. This mostly relates to data from animal models being clouded by the fact that GLP-1 receptors are highly expressed on rodent thyroid cells, resulting alterations of circulating calcitonin levels^[93]. Nonetheless, GLP-1 receptors have been found on the surface of human osteoblast-like cells^[94]. Moreover, very recent data suggest that liraglutide has anabolic effects on bone

in diabetic rats^[95]. In keeping with this, a study in double incretin receptor knockout mice^[89], reported a combination of detrimental bone abnormalities that mimicked observations from both GIP^[96,97] and GLP-1^[98] receptor knockout mice. Despite these observations in rodents, a preliminary meta-analysis suggests that GLP-1 mimetics do not modify the increased bone fracture risk in humans with type 2 diabetes^[99], or could even potentially increase fracture risk in this population^[100]. In keeping with this, a retrospective population based cohort study has suggested that DPP-4 inhibition is not associated with reduced fracture risk in humans^[101], whereas bone loss and strength were significantly improved by sitagliptin therapy in diabetic rats^[102]. Care is required therefore when extrapolating data on the effects of incretin-like drugs on bone from rodents to man, particularly in the case of GLP-1. However, actions of GIP are particularly promising and further research is required to determine if incretin hormones can be useful to treat abnormalities of bone encountered in diabetes and osteoporosis.

Brain

In terms of the central nervous system, expression of functional GIP and GLP-1 receptors has been demonstrated in several brain regions^[103]. Much of the therapeutic interest for incretin-like molecules in the CNS revolves around neuroprotective effects for the treatment of Alzheimer's and Parkinson's diseases, as well as cognitive impairments in diabetes^[3,104]. Accordingly, GIP receptor knockout mice exhibit impaired memory learning, synaptic plasticity, and neurogenesis^[105]. In agreement, transgenic mice that over-express GIP exhibit enhanced sensorimotor coordination and memory recognition^[106]. Earlier studies have already shown that stable forms of GIP can beneficially modulate synaptic transmission and enhance the induction of long-term potentiation, an important physiological cellular means of monitoring learning processes^[107]. In addition, prolonged GIP receptor activation improved cognitive function, hippocampal synaptic plasticity and glucose homeostasis in obese-diabetic high-fat fed mice^[108]. In agreement with this, GLP-1 receptor knockout mice display an impairment of synaptic plasticity and memory formation^[109]. Furthermore, sustained treatment with long-acting GLP-1 agonists improves memory and learning in various rodent models of neurodegeneration and diabetes^[108,110,111]. Moreover, liraglutide treatment has recently been shown to restore cerebral and systemic microvascular architecture in a rodent model of genetically-induced cognitive dysfunction^[112]. Based on the positive neuroprotective effects of incretin compounds, there are several ongoing clinical trials with these drugs that should reveal encouraging effects for the potential treatment of Alzheimer's and Parkinson's diseases^[104]. Finally, in harmony with the positive effects of incretin molecules on brain function, sitagliptin treatment was recently shown to improve recognition memory, oxidative

stress and hippocampal neurogenesis in diabetic mice^[113]. Collectively, these observations strengthen the possibility that incretin peptides play a direct role in modulating aspects of brain function and could possess key clinical pharmacological benefits for patients with diabetes and neurodegenerative disorders.

Heart and vasculature

The GLP-1 receptor has been demonstrated in the heart^[114]. Although some controversy still exists as to the exact location of the receptor within the heart, various studies confirm the presence of GLP-1 receptor mRNA transcripts in rodent and human cardiac tissue^[115]. In cardiomyocytes GLP-1 receptor signalling induced elevations in cAMP levels, but surprisingly this was not coupled to an increase in intracellular Ca²⁺ concentrations and cardiomyocyte contractility^[116]. Indeed, there could be a paradoxical reduction in cardiomyocyte contractility despite elevated cAMP levels^[116]. Moreover, GLP-1 receptor knockout mice present with decreased ventricular contractile function^[117]. As such, the exact mechanism of action and physiological relevance of GLP-1 receptor signalling in the heart requires further detailed investigation. Despite this, and similar to the situation in pancreatic beta-cells, GLP-1 appears to have anti-apoptotic effects in cardiomyocytes and improves overall outcomes in mice after myocardial infarction^[118]. Further to this, GLP-1 receptor protein has also been detected in human coronary artery endothelial cells and encouragingly, activation is believed to improve endothelial cell function in diabetic patients^[119]. Thus, prospective clinical trials are ongoing to assess the cardiovascular safety profile of GLP-1 based peptides, and initial observations in humans with diabetes are positive^[120]. Whilst the GIP receptor is believed to be present in the heart and on vasculature^[103], there is a paucity of knowledge in relation to GIP effects on these tissues. Stimulation of GIP receptors may induce conflicting effects in different vascular beds^[121], and this could explain for its unaccounted physiological effects in these tissues. In keeping with this, the overall effect of DPP-4 inhibition on cardiovascular function is still not clear^[122].

FUTURE DIRECTIONS

Stable gut hormones have considerable potential for the treatment of obesity-related diabetes, and possibly other related pathologies. Whilst disorders of bone, cognitive function and the cardiovascular system can be considered as complications of diabetes, they are also standalone distinct illnesses in their own right. Thus, the therapeutic outlook of incretin mimetics may stretch well beyond diabetes. However, to date only GLP-1 based drugs are clinically available, exclusively for the treatment of type 2 diabetes and associated obesity. Concerns regarding the safety of GLP-1 analogues in man appear to have been allayed, but pharmacovigilance is still required. The potential promise of incretin based drugs

such as GLP-1 mimetics for the treatment of prediabetes still requires detailed investigation. Stable forms of GIP, OXM and CCK also appear to offer distinct therapeutic possibilities for the treatment of type 2 diabetes based on data from animal models and preliminary human studies. Given this, and the multifactorial pathological nature of diabetes, it is not unexpected that concurrent activation of more than one regulatory peptide receptor signalling pathway appears to have promise for the future treatment of diabetes. This may be achieved through the development of double or triple acting agonists or use of a cocktail of existing peptidergic drugs. However, note should be taken of emerging evidence suggesting the utility of sequential peptide exposures to facilitate essential periods of beta-cell rest. Taken together, future advances in our understanding of gut peptide biology, coupled with therapeutic application, should lead to an expansion of clinically available gut peptide-based drugs with far-reaching benefits to the patient.

ACKNOWLEDGMENTS

The authors work on incretin peptides has been supported over many years by Diabetes United Kingdom, European Foundation for the Study of Diabetes, Invest Northern Ireland, Irish Endocrine Society, SAAD Trading and Contracting Company, Department of Education and Learning Northern Ireland, Diabetes Research Wellness Foundation and University of Ulster strategic research funding.

REFERENCES

- 1 **Rehfeld JF**. Gastrointestinal hormones and their targets. *Adv Exp Med Biol* 2014; **817**: 157-175 [PMID: 24997033 DOI: 10.1007/978-1-4939-0897-4_7]
- 2 **Perley MJ**, Kipnis DM. Plasma insulin responses to oral and intravenous glucose: studies in normal and diabetic subjects. *J Clin Invest* 1967; **46**: 1954-1962 [PMID: 6074000 DOI: 10.1172/JCI105685]
- 3 **Irwin N**, Flatt PR. Enteroendocrine hormone mimetics for the treatment of obesity and diabetes. *Curr Opin Pharmacol* 2013; **13**: 989-995 [PMID: 24064397 DOI: 10.1016/j.coph.2013.09.009]
- 4 **Dockray GJ**. Cholecystokinin. *Curr Opin Endocrinol Diabetes Obes* 2012; **19**: 8-12 [PMID: 22157397 DOI: 10.1097/MED.0b-013e32834eb77d]
- 5 **Pocai A**. Unraveling oxyntomodulin, GLP1's enigmatic brother. *J Endocrinol* 2012; **215**: 335-346 [PMID: 23019069 DOI: 10.1530/JOE-12-0368]
- 6 **Gribble FM**. The gut endocrine system as a coordinator of postprandial nutrient homeostasis. *Proc Nutr Soc* 2012; **71**: 456-462 [PMID: 22906726 DOI: 10.1017/S0029665112000705]
- 7 **Nauck MA**, Heimesaat MM, Orskov C, Holst JJ, Ebert R, Creutzfeldt W. Preserved incretin activity of glucagon-like peptide 1 [7-36 amide] but not of synthetic human gastric inhibitory polypeptide in patients with type-2 diabetes mellitus. *J Clin Invest* 1993; **91**: 301-307 [PMID: 8423228 DOI: 10.1172/JCI116186]
- 8 **Drucker DJ**. Deciphering metabolic messages from the gut drives therapeutic innovation: the 2014 Banting Lecture. *Diabetes* 2015; **64**: 317-326 [PMID: 25614665 DOI: 10.2337/db14-1514]
- 9 **Fujita Y**, Wideman RD, Asadi A, Yang GK, Baker R, Webber T, Zhang T, Wang R, Ao Z, Warnock GL, Kwok YN, Kieffer TJ. Glucose-dependent insulinotropic polypeptide is expressed in pancreatic islet alpha-cells and promotes insulin secretion. *Gastroenterology* 2010; **138**: 1966-1975 [PMID: 20138041 DOI: 10.1053/j.gastro.2010.01.049]
- 10 **Whalley NM**, Pritchard LE, Smith DM, White A. Processing of proglucagon to GLP-1 in pancreatic α -cells: is this a paracrine mechanism enabling GLP-1 to act on β -cells? *J Endocrinol* 2011; **211**: 99-106 [PMID: 21795304 DOI: 10.1530/JOE-11-0094]
- 11 **Vasu S**, Moffett RC, Thorens B, Flatt PR. Role of endogenous GLP-1 and GIP in beta cell compensatory responses to insulin resistance and cellular stress. *PLoS One* 2014; **9**: e101005 [PMID: 24967820 DOI: 10.1371/journal.pone.0101005]
- 12 **Moffett RC**, Vasu S, Thorens B, Drucker DJ, Flatt PR. Incretin receptor null mice reveal key role of GLP-1 but not GIP in pancreatic beta cell adaptation to pregnancy. *PLoS One* 2014; **9**: e96863 [PMID: 24927416 DOI: 10.1371/journal.pone.0096863]
- 13 **Moffett RC**, Vasu S, Flatt PR. Functional GIP receptors play a major role in islet compensatory response to high fat feeding in mice. *Biochim Biophys Acta* 2015; **1850**: 1206-1214 [PMID: 25688757 DOI: 10.1016/j.bbagen.2015.02.006]
- 14 **Scott DA**, Boye KS, Timlin L, Clark JF, Best JH. A network meta-analysis to compare glycaemic control in patients with type 2 diabetes treated with exenatide once weekly or liraglutide once daily in comparison with insulin glargine, exenatide twice daily or placebo. *Diabetes Obes Metab* 2013; **15**: 213-223 [PMID: 22958381 DOI: 10.1111/dom.12007]
- 15 **Patterson S**, de Kort M, Irwin N, Moffett RC, Dokter WH, Bos ES, Miltenburg AM, Flatt PR. Pharmacological characterisation and antidiabetic activity of a long-acting GLP-1 analogue conjugated to an ATH1-binding pentasaccharide. *Diabetes Obes Metab* 2015; **17**: 760-770 [PMID: 25929155 DOI: 10.1111/dom.12483]
- 16 **Lund A**, Knop FK, Vilsbøll T. Glucagon-like peptide-1 receptor agonists for the treatment of type 2 diabetes: differences and similarities. *Eur J Intern Med* 2014; **25**: 407-414 [PMID: 24694879 DOI: 10.1016/j.ejim.2014.03.005]
- 17 **Balena R**, Hensley IE, Miller S, Barnett AH. Combination therapy with GLP-1 receptor agonists and basal insulin: a systematic review of the literature. *Diabetes Obes Metab* 2013; **15**: 485-502 [PMID: 23061470 DOI: 10.1111/dom.12025]
- 18 **Gier B**, Butler PC, Lai CK, Kirakossian D, DeNicola MM, Yeh MW. Glucagon like peptide-1 receptor expression in the human thyroid gland. *J Clin Endocrinol Metab* 2012; **97**: 121-131 [PMID: 22031513 DOI: 10.1210/jc.2011-2407]
- 19 **Butler AE**, Campbell-Thompson M, Gurlo T, Dawson DW, Atkinson M, Butler PC. Marked expansion of exocrine and endocrine pancreas with incretin therapy in humans with increased exocrine pancreas dysplasia and the potential for glucagon-producing neuroendocrine tumors. *Diabetes* 2013; **62**: 2595-2604 [PMID: 23524641 DOI: 10.2337/db12-1686]
- 20 **Butler PC**, Elashoff M, Elashoff R, Gale EA. A critical analysis of the clinical use of incretin-based therapies: Are the GLP-1 therapies safe? *Diabetes Care* 2013; **36**: 2118-2125 [PMID: 23645885 DOI: 10.2337/dc12-2713]
- 21 **Alves C**, Batel-Marques F, Macedo AF. A meta-analysis of serious adverse events reported with exenatide and liraglutide: acute pancreatitis and cancer. *Diabetes Res Clin Pract* 2012; **98**: 271-284 [PMID: 23010561 DOI: 10.1016/j.diabres.2012.09.008]
- 22 **Nauck MA**, Meier JJ. Studying pancreatic risks caused by incretin-based therapies: is it a game? It's not a game! *J Diabetes Sci Technol* 2014; **8**: 895-897 [PMID: 24876434 DOI: 10.1177/1932296814532874]
- 23 **Bjerre Knudsen L**, Madsen LW, Andersen S, Almholt K, de Boer AS, Drucker DJ, Gotfredsen C, Egerod FL, Hegelund AC, Jacobsen H, Jacobsen SD, Moses AC, Møllek AM, Nielsen HS, Nowak J, Solberg H, Thi TD, Zdravkovic M, Moerch U. Glucagon-like Peptide-1 receptor agonists activate rodent thyroid C-cells causing calcitonin release and C-cell proliferation. *Endocrinology* 2010; **151**: 1473-1486 [PMID: 20203154 DOI: 10.1210/en.2009-1272]
- 24 **Nauck MA**, Friedrich N. Do GLP-1-based therapies increase cancer risk? *Diabetes Care* 2013; **36** Suppl 2: S245-S252 [PMID: 23882053 DOI: 10.2337/dcS13-2004]

- 25 **Ross SA**, Dupre J. Effects of ingestion of triglyceride or galactose on secretion of gastric inhibitory polypeptide and on responses to intravenous glucose in normal and diabetic subjects. *Diabetes* 1978; **27**: 327-333 [PMID: 640238 DOI: 10.2337/diab.27.3.327]
- 26 **Ehses JA**, Casilla VR, Doty T, Pospisilik JA, Winter KD, Demuth HU, Pederson RA, McIntosh CH. Glucose-dependent insulinotropic polypeptide promotes beta-(INS-1) cell survival via cyclic adenosine monophosphate-mediated caspase-3 inhibition and regulation of p38 mitogen-activated protein kinase. *Endocrinology* 2003; **144**: 4433-4445 [PMID: 12960055 DOI: 10.1210/en.2002-0068]
- 27 **Renner S**, Fehlings C, Herbach N, Hofmann A, von Waldhausen DC, Kessler B, Ulrichs K, Chodnevskaja I, Moskalenko V, Amselgruber W, Göke B, Pfeifer A, Wanke R, Wolf E. Glucose intolerance and reduced proliferation of pancreatic beta-cells in transgenic pigs with impaired glucose-dependent insulinotropic polypeptide function. *Diabetes* 2010; **59**: 1228-1238 [PMID: 20185813 DOI: 10.2337/db09-0519]
- 28 **Kubota A**, Yamada Y, Yasuda K, Someya Y, Ihara Y, Kagimoto S, Watanabe R, Kuroe A, Ishida H, Seino Y. Gastric inhibitory polypeptide activates MAP kinase through the wortmannin-sensitive and -insensitive pathways. *Biochem Biophys Res Commun* 1997; **235**: 171-175 [PMID: 9196057 DOI: 10.1006/bbrc.1997.6743]
- 29 **Kim SJ**, Winter K, Nian C, Tsuneoka M, Koda Y, McIntosh CH. Glucose-dependent insulinotropic polypeptide (GIP) stimulation of pancreatic beta-cell survival is dependent upon phosphatidylinositol 3-kinase (PI3K)/protein kinase B (PKB) signaling, inactivation of the forkhead transcription factor Foxo1, and down-regulation of bax expression. *J Biol Chem* 2005; **280**: 22297-22307 [PMID: 15817464 DOI: 10.1074/jbc.M500540200]
- 30 **Deacon CF**, Åhrén B. Physiology of incretins in health and disease. *Rev Diabet Stud* 2011; **8**: 293-306 [PMID: 22262068 DOI: 10.1900/RDS.2011.8.293]
- 31 **Gault VA**, O'Harte FP, Flatt PR. Glucose-dependent insulinotropic polypeptide (GIP): anti-diabetic and anti-obesity potential? *Neuropeptides* 2003; **37**: 253-263 [PMID: 14607102 DOI: 10.1016/j.npep.2003.09.002]
- 32 **Aaboe K**, Knop FK, Vilsbøll T, Vølund A, Simonsen U, Deacon CF, Madsbad S, Holst JJ, Krarup T. KATP channel closure ameliorates the impaired insulinotropic effect of glucose-dependent insulinotropic polypeptide in patients with type 2 diabetes. *J Clin Endocrinol Metab* 2009; **94**: 603-608 [PMID: 19050053 DOI: 10.1210/jc.2008-1731]
- 33 **Irwin N**, Gault V, Flatt PR. Therapeutic potential of the original incretin hormone glucose-dependent insulinotropic polypeptide: diabetes, obesity, osteoporosis and Alzheimer's disease? *Expert Opin Investig Drugs* 2010; **19**: 1039-1048 [PMID: 20698813 DOI: 10.1517/13543784.2010.513381]
- 34 **Højberg PV**, Vilsbøll T, Rabøl R, Knop FK, Bache M, Krarup T, Holst JJ, Madsbad S. Four weeks of near-normalisation of blood glucose improves the insulin response to glucagon-like peptide-1 and glucose-dependent insulinotropic polypeptide in patients with type 2 diabetes. *Diabetologia* 2009; **52**: 199-207 [PMID: 19037628 DOI: 10.1007/s00125-008-1195-5]
- 35 **Taylor AI**, Irwin N, McKillop AM, Patterson S, Flatt PR, Gault VA. Evaluation of the degradation and metabolic effects of the gut peptide xenin on insulin secretion, glycaemic control and satiety. *J Endocrinol* 2010; **207**: 87-93 [PMID: 20631047 DOI: 10.1677/JOE-10-0085]
- 36 **Wice BM**, Wang S, Crimmins DL, Diggs-Andrews KA, Althage MC, Ford EL, Tran H, Ohlendorf M, Griest TA, Wang Q, Fisher SJ, Ladenson JH, Polonsky KS. Xenin-25 potentiates glucose-dependent insulinotropic polypeptide action via a novel cholinergic relay mechanism. *J Biol Chem* 2010; **285**: 19842-19853 [PMID: 20421298 DOI: 10.1074/jbc.M110.129304]
- 37 **Martin CM**, Gault VA, McClean S, Flatt PR, Irwin N. Degradation, insulin secretion, glucose-lowering and GIP additive actions of a palmitate-derivatised analogue of xenin-25. *Biochem Pharmacol* 2012; **84**: 312-319 [PMID: 22561048 DOI: 10.1016/j.bcp.2012.04.015]
- 38 **Gault VA**, Martin CM, Flatt PR, Parthasarathy V, Irwin N. Xenin-25[Lys13PAL]: a novel long-acting acylated analogue of xenin-25 with promising antidiabetic potential. *Acta Diabetol* 2015; **52**: 461-471 [PMID: 25374384 DOI: 10.1007/s00592-014-0681-0]
- 39 **Pathak V**, Vasu S, Flatt PR, Irwin N. Effects of chronic exposure of clonal β -cells to elevated glucose and free fatty acids on incretin receptor gene expression and secretory responses to GIP and GLP-1. *Diabetes Obes Metab* 2014; **16**: 357-365 [PMID: 24164718 DOI: 10.1111/dom.12227]
- 40 **Wren AM**, Bloom SR. Gut hormones and appetite control. *Gastroenterology* 2007; **132**: 2116-2130 [PMID: 17498507 DOI: 10.1053/j.gastro.2007.03.048]
- 41 **Karra E**, Batterham RL. The role of gut hormones in the regulation of body weight and energy homeostasis. *Mol Cell Endocrinol* 2010; **316**: 120-128 [PMID: 19563862 DOI: 10.1016/j.mce.2009.06.010]
- 42 **Cohen MA**, Ellis SM, Le Roux CW, Batterham RL, Park A, Patterson M, Frost GS, Ghatei MA, Bloom SR. Oxyntomodulin suppresses appetite and reduces food intake in humans. *J Clin Endocrinol Metab* 2003; **88**: 4696-4701 [PMID: 14557443 DOI: 10.1210/jc.2003-030421]
- 43 **Wynne K**, Park AJ, Small CJ, Patterson M, Ellis SM, Murphy KG, Wren AM, Frost GS, Meeran K, Ghatei MA, Bloom SR. Subcutaneous oxyntomodulin reduces body weight in overweight and obese subjects: a double-blind, randomized, controlled trial. *Diabetes* 2005; **54**: 2390-2395 [PMID: 16046306 DOI: 10.2337/diabetes.54.8.2390]
- 44 **Druce MR**, Minnion JS, Field BC, Patel SR, Shillito JC, Tilby M, Beale KE, Murphy KG, Ghatei MA, Bloom SR. Investigation of structure-activity relationships of Oxyntomodulin (Oxm) using Oxm analogs. *Endocrinology* 2009; **150**: 1712-1722 [PMID: 19074579 DOI: 10.1210/en.2008-0828]
- 45 **Kerr BD**, Flatt PR, Gault VA. (D-Ser2)Oxm[mPEG-PAL]: a novel chemically modified analogue of oxyntomodulin with antihyperglycaemic, insulinotropic and anorexigenic actions. *Biochem Pharmacol* 2010; **80**: 1727-1735 [PMID: 20735990 DOI: 10.1016/j.bcp.2010.08.010]
- 46 **Lynch AM**, Pathak N, Flatt YE, Gault VA, O'Harte FP, Irwin N, Flatt PR. Comparison of stability, cellular, glucose-lowering and appetite suppressing effects of oxyntomodulin analogues modified at the N-terminus. *Eur J Pharmacol* 2014; **743**: 69-78 [PMID: 25246014 DOI: 10.1016/j.ejphar.2014.09.018]
- 47 **Cegla J**, Troke RC, Jones B, Tharakan G, Kenkre J, McCullough KA, Lim CT, Parvizi N, Hussein M, Chambers ES, Minnion J, Cuenco J, Ghatei MA, Meeran K, Tan TM, Bloom SR. Coinfusion of low-dose GLP-1 and glucagon in man results in a reduction in food intake. *Diabetes* 2014; **63**: 3711-3720 [PMID: 24939425 DOI: 10.2337/db14-0242]
- 48 **Cummings DE**, Overduin J. Gastrointestinal regulation of food intake. *J Clin Invest* 2007; **117**: 13-23 [PMID: 17200702 DOI: 10.1172/JCI30227]
- 49 **Wank SA**. Cholecystokinin receptors. *Am J Physiol* 1995; **269**: G628-G646 [PMID: 7491953]
- 50 **Åhrén B**, Pettersson M, Uvnäs-Moberg K, Gutniak M, Efendic S. Effects of cholecystokinin (CCK)-8, CCK-33, and gastric inhibitory polypeptide (GIP) on basal and meal-stimulated pancreatic hormone secretion in man. *Diabetes Res Clin Pract* 1991; **13**: 153-161 [PMID: 1683622 DOI: 10.1016/0168-8227(91)90059-M]
- 51 **Irwin N**, Frizelle P, Montgomery IA, Moffett RC, O'Harte FP, Flatt PR. Beneficial effects of the novel cholecystokinin agonist (pGlu-Gln)-CCK-8 in mouse models of obesity/diabetes. *Diabetologia* 2012; **55**: 2747-2758 [PMID: 22814764 DOI: 10.1007/s00125-012-2654-6]
- 52 **Lavine JA**, Raess PW, Stapleton DS, Rabaglia ME, Suhonen JI, Schueler KL, Koltjes JE, Dawson JA, Yandell BS, Samuelson LC, Beinfeld MC, Davis DB, Hellerstein MK, Keller MP, Attie AD. Cholecystokinin is up-regulated in obese mouse islets and expands beta-cell mass by increasing beta-cell survival. *Endocrinology* 2010; **151**: 3577-3588 [PMID: 20534724 DOI: 10.1210/en.2010-0233]
- 53 **O'Harte FP**, Mooney MH, Kelly CM, Flatt PR. Glycated cholecystokinin-8 has an enhanced satiating activity and is protected against enzymatic degradation. *Diabetes* 1998; **47**: 1619-1624 [PMID: 9753301 DOI: 10.2337/diabetes.47.10.1619]

- 54 **Verbaeys I**, León-Tamariz F, Buyse J, Decuypere E, Pottel H, Cokelaere M. Lack of tolerance development with long-term administration of PEGylated cholecystokinin. *Peptides* 2009; **30**: 699-704 [PMID: 19084041 DOI: 10.1016/j.peptides.2008.11.010]
- 55 **Irwin N**, Montgomery IA, Moffett RC, Flatt PR. Chemical cholecystokinin receptor activation protects against obesity-diabetes in high fat fed mice and has sustainable beneficial effects in genetic ob/ob mice. *Biochem Pharmacol* 2013; **85**: 81-91 [PMID: 23085436 DOI: 10.1016/j.bcp.2012.10.008]
- 56 **Irwin N**, Frizelle P, O'Harte FP, Flatt PR. Metabolic effects of activation of CCK receptor signaling pathways by twice-daily administration of the enzyme-resistant CCK-8 analog, (pGlu-Gln)-CCK-8, in normal mice. *J Endocrinol* 2013; **216**: 53-59 [PMID: 23055353 DOI: 10.1530/JOE-12-0353]
- 57 **Montgomery IA**, Irwin N, Flatt PR. Beneficial effects of (pGlu-Gln)-CCK-8 on energy intake and metabolism in high fat fed mice are associated with alterations of hypothalamic gene expression. *Horm Metab Res* 2013; **45**: 471-473 [PMID: 23315994 DOI: 10.1055/s-0032-1331767]
- 58 **Irwin N**, Frizelle P, O'Harte FP, Flatt PR. (pGlu-Gln)-CCK-8[mPEG]: a novel, long-acting, mini-PEGylated cholecystokinin (CCK) agonist that improves metabolic status in dietary-induced diabetes. *Biochim Biophys Acta* 2013; **1830**: 4009-4016 [PMID: 23583730 DOI: 10.1016/j.bbagen.2013.04.004]
- 59 **Gallwitz B**, Witt M, Morys-Wortmann C, Fölsch UR, Schmidt WE. GLP-1/GIP chimeric peptides define the structural requirements for specific ligand-receptor interaction of GLP-1. *Regul Pept* 1996; **63**: 17-22 [PMID: 8795084 DOI: 10.1016/0167-0115(96)00019-5]
- 60 **Irwin N**, Flatt PR. Evidence for beneficial effects of compromised gastric inhibitory polypeptide action in obesity-related diabetes and possible therapeutic implications. *Diabetologia* 2009; **52**: 1724-1731 [PMID: 19533083 DOI: 10.1007/s00125-009-1422-8]
- 61 **Gough SC**, Bode B, Woo V, Rodbard HW, Linjawi S, Poulsen P, Damgaard LH, Buse JB; NN9068-3697 (DUAL-I) trial investigators. Efficacy and safety of a fixed-ratio combination of insulin degludec and liraglutide (IDegLira) compared with its components given alone: results of a phase 3, open-label, randomised, 26-week, treat-to-target trial in insulin-naïve patients with type 2 diabetes. *Lancet Diabetes Endocrinol* 2014; **2**: 885-893 [DOI: 10.1016/S2213-8587(14)70174-3]
- 62 **Gallwitz B**. Emerging DPP-4 inhibitors: focus on linagliptin for type 2 diabetes. *Diabetes Metab Syndr Obes* 2013; **6**: 1-9 [PMID: 23319869 DOI: 10.2147/DMSO.S23166]
- 63 **Pan CQ**, Buxton JM, Yung SL, Tom I, Yang L, Chen H, MacDougall M, Bell A, Claus TH, Clairmont KB, Whelan JP. Design of a long acting peptide functioning as both a glucagon-like peptide-1 receptor agonist and a glucagon receptor antagonist. *J Biol Chem* 2006; **281**: 12506-12515 [PMID: 16505481 DOI: 10.1074/jbc.M60012700]
- 64 **Pocai A**, Carrington PE, Adams JR, Wright M, Eiermann G, Zhu L, Du X, Petrov A, Lassman ME, Jiang G, Liu F, Miller C, Tota LM, Zhou G, Zhang X, Sountis MM, Santoprete A, Capito E, Chicchi GG, Thornberry N, Bianchi E, Pessi A, Marsh DJ, SinhaRoy R. Glucagon-like peptide 1/glucagon receptor dual agonism reverses obesity in mice. *Diabetes* 2009; **58**: 2258-2266 [PMID: 19602537 DOI: 10.2337/db09-0278]
- 65 **Sinclair EM**, Drucker DJ. Proglucagon-derived peptides: mechanisms of action and therapeutic potential. *Physiology* (Bethesda) 2005; **20**: 357-365 [PMID: 16174875 DOI: 10.1152/physiol.00030.2005]
- 66 **Fosgerau K**, Jessen L, Lind Tolborg J, Østerlund T, Schæffer Larsen K, Rolsted K, Brorson M, Jelsing J, Skovlund Ryge Neerup T. The novel GLP-1-gastrin dual agonist, ZP3022, increases β -cell mass and prevents diabetes in db/db mice. *Diabetes Obes Metab* 2013; **15**: 62-71 [PMID: 22862961 DOI: 10.1111/j.1463-1326.2012.01676.x]
- 67 **Irwin N**, Hunter K, Montgomery IA, Flatt PR. Comparison of independent and combined metabolic effects of chronic treatment with (pGlu-Gln)-CCK-8 and long-acting GLP-1 and GIP mimetics in high fat-fed mice. *Diabetes Obes Metab* 2013; **15**: 650-659 [PMID: 23388064 DOI: 10.1111/dom.12079]
- 68 **Trevaskis JL**, Sun C, Athanacio J, D'Souza L, Samant M, Tatarkiewicz K, Griffin PS, Wittmer C, Wang Y, Teng CH, Forood B, Parkes DG, Roth JD. Synergistic metabolic benefits of an exenatide analogue and cholecystokinin in diet-induced obese and leptin-deficient rodents. *Diabetes Obes Metab* 2015; **17**: 61-73 [PMID: 25204356 DOI: 10.1111/dom.12390]
- 69 **Irwin N**, Pathak V, Flatt PR. A novel CCK-8/GLP-1 hybrid peptide exhibiting prominent insulinotropic, glucose-lowering and satiety actions with significant therapeutic potential in high-fat fed mice. *Diabetes* 2015; **64**: 2996-3009 [PMID: 25883113 DOI: 10.2337/db15-0220]
- 70 **Flatt PR**. Dorothy Hodgkin Lecture 2008. Gastric inhibitory polypeptide (GIP) revisited: a new therapeutic target for obesity-diabetes? *Diabet Med* 2008; **25**: 759-764 [PMID: 18513308 DOI: 10.1111/j.1464-5491.2008.02455.x]
- 71 **Knop FK**, Taylor R. Mechanism of metabolic advantages after bariatric surgery: it's all gastrointestinal factors versus it's all food restriction. *Diabetes Care* 2013; **36** Suppl 2: S287-S291 [PMID: 23882061 DOI: 10.2337/dcS13-2032]
- 72 **Bhat VK**, Kerr BD, Flatt PR, Gault VA. A novel GIP-oxyntomodulin hybrid peptide acting through GIP, glucagon and GLP-1 receptors exhibits weight reducing and anti-diabetic properties. *Biochem Pharmacol* 2013; **85**: 1655-1662 [PMID: 23518155 DOI: 10.1016/j.bcp.2013.03.009]
- 73 **Bhat VK**, Kerr BD, Vasu S, Flatt PR, Gault VA. A DPP-IV-resistant triple-acting agonist of GIP, GLP-1 and glucagon receptors with potent glucose-lowering and insulinotropic actions in high-fat-fed mice. *Diabetologia* 2013; **56**: 1417-1424 [PMID: 23503814 DOI: 10.1007/s00125-013-2892-2]
- 74 **Finan B**, Yang B, Ottaway N, Smiley DL, Ma T, Clemmensen C, Chabenne J, Zhang L, Habegger KM, Fischer K, Campbell JE, Sandoval D, Seeley RJ, Bleicher K, Uhles S, Riboulet W, Funk J, Hertel C, Belli S, Sebokova E, Conde-Knape K, Konkari A, Drucker DJ, Gelfanov V, Pfluger PT, Müller TD, Perez-Tilve D, DiMarchi RD, Tschöp MH. A rationally designed monomeric peptide triagonist corrects obesity and diabetes in rodents. *Nat Med* 2015; **21**: 27-36 [PMID: 25485909 DOI: 10.1038/nm.3761]
- 75 **Brown RJ**, Rother KI. Effects of beta-cell rest on beta-cell function: a review of clinical and preclinical data. *Pediatr Diabetes* 2008; **9**: 14-22 [PMID: 18221429 DOI: 10.1111/j.1399-5448.2007.00272.x]
- 76 **Maedler K**, Carr RD, Bosco D, Zuellig RA, Berney T, Donath MY. Sulfonylurea induced beta-cell apoptosis in cultured human islets. *J Clin Endocrinol Metab* 2005; **90**: 501-506 [PMID: 15483097 DOI: 10.1210/jc.2004-0699]
- 77 **Ritzel RA**, Hansen JB, Veldhuis JD, Butler PC. Induction of beta-cell rest by a Kir6.2/SUR1-selective K(ATP)-channel opener preserves beta-cell insulin stores and insulin secretion in human islets cultured at high (11 mM) glucose. *J Clin Endocrinol Metab* 2004; **89**: 795-805 [PMID: 14764798 DOI: 10.1210/jc.2003-031120]
- 78 **Irwin N**, Hunter K, Frizzell N, Flatt PR. Antidiabetic effects of sub-chronic activation of the GIP receptor alone and in combination with background exendin-4 therapy in high fat fed mice. *Regul Pept* 2009; **153**: 70-76 [PMID: 19073224 DOI: 10.1016/j.regpep.2008.11.007]
- 79 **Nasteska D**, Harada N, Suzuki K, Yamane S, Hamasaki A, Joo E, Iwasaki K, Shibue K, Harada T, Inagaki N. Chronic reduction of GIP secretion alleviates obesity and insulin resistance under high-fat diet conditions. *Diabetes* 2014; **63**: 2332-2343 [PMID: 24584548 DOI: 10.2337/db13-1563]
- 80 **Pathak V**, Vasu S, Gault VA, Flatt PR, Irwin N. Sequential induction of beta cell rest and stimulation using stable GIP inhibitor and GLP-1 mimetic peptides improves metabolic control in C57BL/KsJ db/db mice. *Diabetologia* 2015; **58**: 2144-2153 [PMID: 26048235 DOI: 10.1007/s00125-015-3653-1]
- 81 **Ahmadi H**, Azar ST. The role of incretin-based therapies in prediabetes: a review. *Prim Care Diabetes* 2014; **8**: 286-294 [PMID: 24666932 DOI: 10.1016/j.pcd.2014.02.005]
- 82 **Laakso M**, Zilinskaite J, Hansen T, Boesgaard TW, Vanttinen M,

- Stancáková A, Jansson PA, Pellmé F, Holst JJ, Kuulasmaa T, Hribal ML, Sesti G, Stefan N, Fritsche A, Häring H, Pedersen O, Smith U. Insulin sensitivity, insulin release and glucagon-like peptide-1 levels in persons with impaired fasting glucose and/or impaired glucose tolerance in the EUGENE2 study. *Diabetologia* 2008; **51**: 502-511 [PMID: 18080106 DOI: 10.1007/s00125-007-0899-2]
- 83 **Utzschneider KM**, Tong J, Montgomery B, Udayasankar J, Gerchman F, Marcovina SM, Watson CE, Ligueros-Saylan MA, Foley JE, Holst JJ, Deacon CF, Kahn SE. The dipeptidyl peptidase-4 inhibitor vildagliptin improves beta-cell function and insulin sensitivity in subjects with impaired fasting glucose. *Diabetes Care* 2008; **31**: 108-113 [PMID: 17909087 DOI: 10.2337/dc07-1441]
- 84 **Rosenstock J**, Foley JE, Rendell M, Landin-Olsson M, Holst JJ, Deacon CF, Rochotte E, Baron MA. Effects of the dipeptidyl peptidase-IV inhibitor vildagliptin on incretin hormones, islet function, and postprandial glycemia in subjects with impaired glucose tolerance. *Diabetes Care* 2008; **31**: 30-35 [PMID: 17947341 DOI: 10.2337/dc07-1616]
- 85 **Bock G**, Dalla Man C, Micheletto F, Basu R, Giesler PD, Laugen J, Deacon CF, Holst JJ, Toffolo G, Cobelli C, Rizza RA, Vella A. The effect of DPP-4 inhibition with sitagliptin on incretin secretion and on fasting and postprandial glucose turnover in subjects with impaired fasting glucose. *Clin Endocrinol (Oxf)* 2010; **73**: 189-196 [PMID: 20039889]
- 86 **Rosenstock J**, Klaff LJ, Schwartz S, Northrup J, Holcombe JH, Wilhelm K, Trautmann M. Effects of exenatide and lifestyle modification on body weight and glucose tolerance in obese subjects with and without pre-diabetes. *Diabetes Care* 2010; **33**: 1173-1175 [PMID: 20332357 DOI: 10.2337/dc09-1203]
- 87 **Astrup A**, Rössner S, Van Gaal L, Rissanen A, Niskanen L, Al Hakim M, Madsen J, Rasmussen MF, Lean ME. Effects of liraglutide in the treatment of obesity: a randomised, double-blind, placebo-controlled study. *Lancet* 2009; **374**: 1606-1616 [PMID: 19853906 DOI: 10.1016/S0140-6736(09)61375-1]
- 88 **Aroda VR**, Ratner R. The safety and tolerability of GLP-1 receptor agonists in the treatment of type 2 diabetes: a review. *Diabetes Metab Res Rev* 2011; **27**: 528-542 [PMID: 21484979 DOI: 10.1002/dmrr.1202]
- 89 **Mieczkowska A**, Mansur S, Bouvard B, Flatt PR, Thorens B, Irwin N, Chappard D, Mabilletau G. Double incretin receptor knock-out (DIRKO) mice present with alterations of trabecular and cortical micromorphology and bone strength. *Osteoporos Int* 2015; **26**: 209-218 [PMID: 25127672 DOI: 10.1007/s00198-014-2845-8]
- 90 **Bollag RJ**, Zhong Q, Phillips P, Min L, Zhong L, Cameron R, Mulloy AL, Rasmussen H, Qin F, Ding KH, Isaacs CM. Osteoblast-derived cells express functional glucose-dependent insulinotropic peptide receptors. *Endocrinology* 2000; **141**: 1228-35 [DOI: 10.1210/en.141.3.1228]
- 91 **Nissen A**, Christensen M, Knop FK, Vilsbøll T, Holst JJ, Hartmann B. Glucose-dependent insulinotropic polypeptide inhibits bone resorption in humans. *J Clin Endocrinol Metab* 2014; **99**: E2325-E2329 [PMID: 25144635 DOI: 10.1210/jc.2014-2547]
- 92 **Mabilletau G**, Mieczkowska A, Irwin N, Simon Y, Audran M, Flatt PR, Chappard D. Beneficial effects of a N-terminally modified GIP agonist on tissue-level bone material properties. *Bone* 2014; **63**: 61-68 [PMID: 24594344 DOI: 10.1016/j.bone.2014.02.013]
- 93 **Yamada C**, Yamada Y, Tsukiyama K, Yamada K, Udagawa N, Takahashi N, Tanaka K, Drucker DJ, Seino Y, Inagaki N. The murine glucagon-like peptide-1 receptor is essential for control of bone resorption. *Endocrinology* 2008; **149**: 574-579 [PMID: 18039776 DOI: 10.1210/en.2007-1292]
- 94 **Pacheco-Pantoja EL**, Ranganath LR, Gallagher JA, Wilson PJ, Fraser WD. Receptors and effects of gut hormones in three osteoblastic cell lines. *BMC Physiol* 2011; **11**: 12 [PMID: 21801348 DOI: 10.1186/1472-6793-11-12]
- 95 **Sun HX**, Lu N, Luo X, Zhao L, Liu JM. Liraglutide, the glucagon-like peptide-1 receptor agonist, has anabolic bone effects in diabetic Goto-Kakizaki rats. *J Diabetes* 2015; **7**: 584-588 [PMID: 25753217 DOI: 10.1111/1753-0407.12282]
- 96 **Mieczkowska A**, Irwin N, Flatt PR, Chappard D, Mabilletau G. Glucose-dependent insulinotropic polypeptide (GIP) receptor deletion leads to reduced bone strength and quality. *Bone* 2013; **56**: 337-342 [PMID: 23851294 DOI: 10.1016/j.bone.2013.07.003]
- 97 **Gaudin-Audrain C**, Irwin N, Mansur S, Flatt PR, Thorens B, Baslé M, Chappard D, Mabilletau G. Glucose-dependent insulinotropic polypeptide receptor deficiency leads to modifications of trabecular bone volume and quality in mice. *Bone* 2013; **53**: 221-230 [PMID: 23220186 DOI: 10.1016/j.bone.2012.11.039]
- 98 **Mabilletau G**, Mieczkowska A, Irwin N, Flatt PR, Chappard D. Optimal bone mechanical and material properties require a functional glucagon-like peptide-1 receptor. *J Endocrinol* 2013; **219**: 59-68 [PMID: 23911987 DOI: 10.1530/JOE-13-0146]
- 99 **Mabilletau G**, Mieczkowska A, Chappard D. Use of glucagon-like peptide-1 receptor agonists and bone fractures: a meta-analysis of randomized clinical trials. *J Diabetes* 2014; **6**: 260-266 [PMID: 24164867 DOI: 10.1111/1753-0407.12102]
- 100 **Su B**, Sheng H, Zhang M, Bu L, Yang P, Li L, Li F, Sheng C, Han Y, Qu S, Wang J. Risk of bone fractures associated with glucagon-like peptide-1 receptor agonists' treatment: a meta-analysis of randomized controlled trials. *Endocrine* 2015; **48**: 107-115 [PMID: 25074632 DOI: 10.1007/s12020-014-0361-4]
- 101 **Driessen JH**, van Onzenoort HA, Henry RM, Lalmohamed A, van den Bergh JP, Neef C, Leufkens HG, de Vries F. Use of dipeptidyl peptidase-4 inhibitors for type 2 diabetes mellitus and risk of fracture. *Bone* 2014; **68**: 124-130 [PMID: 25093264 DOI: 10.1016/j.bone.2014.07.030]
- 102 **Glorie L**, Behets GJ, Baerts L, De Meester I, D'Haese PC, Verhulst A. DPP IV inhibitor treatment attenuates bone loss and improves mechanical bone strength in male diabetic rats. *Am J Physiol Endocrinol Metab* 2014; **307**: E447-E455 [PMID: 25053403 DOI: 10.1152/ajpendo.00217.2014]
- 103 **Usdin TB**, Mezey E, Button DC, Brownstein MJ, Bonner TI. Gastric inhibitory polypeptide receptor, a member of the secretin-vasoactive intestinal peptide receptor family, is widely distributed in peripheral organs and the brain. *Endocrinology* 1993; **133**: 2861-2870 [PMID: 8243312]
- 104 **Hölscher C**. New drug treatments show neuroprotective effects in Alzheimer's and Parkinson's diseases. *Neural Regen Res* 2014; **9**: 1870-1873 [PMID: 25558231 DOI: 10.4103/1673-5374.145342]
- 105 **Faivre E**, Gault VA, Thorens B, Hölscher C. Glucose-dependent insulinotropic polypeptide receptor knockout mice are impaired in learning, synaptic plasticity, and neurogenesis. *J Neurophysiol* 2011; **105**: 1574-1580 [PMID: 21273318 DOI: 10.1152/jn.00866.2010]
- 106 **Ding KH**, Zhong Q, Xie D, Chen HX, Della-Fera MA, Bollag RJ, Bollag WB, Gujral R, Kang B, Sridhar S, Baile C, Curl W, Isaacs CM. Effects of glucose-dependent insulinotropic peptide on behavior. *Peptides* 2006; **27**: 2750-2755 [PMID: 16822587 DOI: 10.1016/j.peptides.2006.05.011]
- 107 **Gault VA**, Hölscher C. Protease-resistant glucose-dependent insulinotropic polypeptide agonists facilitate hippocampal LTP and reverse the impairment of LTP induced by beta-amyloid. *J Neurophysiol* 2008; **99**: 1590-1595 [PMID: 18234983 DOI: 10.1152/jn.01161.2007]
- 108 **Porter DW**, Irwin N, Flatt PR, Hölscher C, Gault VA. Prolonged GIP receptor activation improves cognitive function, hippocampal synaptic plasticity and glucose homeostasis in high-fat fed mice. *Eur J Pharmacol* 2011; **650**: 688-693 [PMID: 21050845 DOI: 10.1016/j.ejphar.2010.10.059]
- 109 **Abbas T**, Faivre E, Hölscher C. Impairment of synaptic plasticity and memory formation in GLP-1 receptor KO mice: Interaction between type 2 diabetes and Alzheimer's disease. *Behav Brain Res* 2009; **205**: 265-271 [PMID: 19573562 DOI: 10.1016/j.bbr.2009.06.035]
- 110 **McClean PL**, Hölscher C. Lixisenatide, a drug developed to treat type 2 diabetes, shows neuroprotective effects in a mouse model of Alzheimer's disease. *Neuropharmacology* 2014; **86**: 241-258 [PMID: 25107586 DOI: 10.1016/j.neuropharm.2014.07.015]
- 111 **McGovern SF**, Hunter K, Hölscher C. Effects of the glucagon-like polypeptide-1 analogue (Val⁸)GLP-1 on learning, progenitor cell proliferation and neurogenesis in the C57B/16 mouse brain.

- Brain Res* 2012; **1473**: 204-213 [PMID: 22867941 DOI: 10.1016/j.brainres.2012.07.029]
- 112 **Kelly P**, McClean PL, Ackermann M, Konerding MA, Hölscher C, Mitchell CA. Restoration of cerebral and systemic microvascular architecture in APP/PS1 transgenic mice following treatment with Liraglutide™. *Microcirculation* 2015; **22**: 133-145 [PMID: 25556713 DOI: 10.1111/micc.12186]
 - 113 **Gault VA**, Lennox R, Flatt PR. Sitagliptin, a dipeptidyl peptidase-4 inhibitor, improves recognition memory, oxidative stress and hippocampal neurogenesis and upregulates key genes involved in cognitive decline. *Diabetes Obes Metab* 2015; **17**: 403-413 [PMID: 25580570 DOI: 10.1111/dom.12432]
 - 114 **Bullock BP**, Heller RS, Habener JF. Tissue distribution of messenger ribonucleic acid encoding the rat glucagon-like peptide-1 receptor. *Endocrinology* 1996; **137**: 2968-2978 [PMID: 8770921]
 - 115 **Campbell JE**, Drucker DJ. Pharmacology, physiology, and mechanisms of incretin hormone action. *Cell Metab* 2013; **17**: 819-837 [PMID: 23684623 DOI: 10.1016/j.cmet.2013.04.008]
 - 116 **Vila Petroff MG**, Egan JM, Wang X, Sollott SJ. Glucagon-like peptide-1 increases cAMP but fails to augment contraction in adult rat cardiac myocytes. *Circ Res* 2001; **89**: 445-452 [PMID: 11532906 DOI: 10.1161/hh1701.095716]
 - 117 **Gros R**, You X, Baggio LL, Kabir MG, Sadi AM, Mungrue IN, Parker TG, Huang Q, Drucker DJ, Husain M. Cardiac function in mice lacking the glucagon-like peptide-1 receptor. *Endocrinology* 2003; **144**: 2242-2252 [PMID: 12746281 DOI: 10.1210/en.2003-0007]
 - 118 **Noyan-Ashraf MH**, Momen MA, Ban K, Sadi AM, Zhou YQ, Riazi AM, Baggio LL, Henkelman RM, Husain M, Drucker DJ. GLP-1R agonist liraglutide activates cytoprotective pathways and improves outcomes after experimental myocardial infarction in mice. *Diabetes* 2009; **58**: 975-983 [PMID: 19151200 DOI: 10.2337/db08-1193]
 - 119 **Nyström T**, Gutniak MK, Zhang Q, Zhang F, Holst JJ, Ahrén B, Sjöholm A. Effects of glucagon-like peptide-1 on endothelial function in type 2 diabetes patients with stable coronary artery disease. *Am J Physiol Endocrinol Metab* 2004; **287**: E1209-E1215 [PMID: 15353407 DOI: 10.1152/ajpendo.00237.2004]
 - 120 **Avogaro A**, Vigili de Kreutzenberg S, Fadini GP. Cardiovascular actions of GLP-1 and incretin-based pharmacotherapy. *Curr Diab Rep* 2014; **14**: 483 [DOI: 10.1007/s11892-014-0483-3]
 - 121 **Ding KH**, Zhong Q, Xu J, Isaacs CM. Glucose-dependent insulinotropic peptide: differential effects on hepatic artery vs. portal vein endothelial cells. *Am J Physiol Endocrinol Metab* 2004; **286**: E773-E779 [PMID: 14709420 DOI: 10.1152/ajpendo.00507.2003]
 - 122 **Wu S**, Hopper I, Skiba M, Krum H. Dipeptidyl peptidase-4 inhibitors and cardiovascular outcomes: meta-analysis of randomized clinical trials with 55,141 participants. *Cardiovasc Ther* 2014; **32**: 147-158 [PMID: 24750644 DOI: 10.1111/1755-5922.12075]

P- Reviewer: Ali O, Collino M **S- Editor:** Qi Y **L- Editor:** A
E- Editor: Lu YJ



Role of adiponectin and some other factors linking type 2 diabetes mellitus and obesity

Chandra Kanti Chakraborti

Chandra Kanti Chakraborti, Kanak Manjari Institute of Pharmaceutical Sciences, Rourkela 769015, Odisha, India

Author contributions: Chakraborti CK solely contributed to this manuscript.

Conflict-of-interest statement: The author has no conflict of interests.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Correspondence to: Dr. Chandra Kanti Chakraborti, Professor, Kanak Manjari Institute of Pharmaceutical Sciences, Chhend, Rourkela 769015, Odisha, India. chandrakanti_12@rediffmail.com
 Telephone: +91-0977-6145092
 Fax: +91-0661-2480752

Received: July 9, 2015
 Peer-review started: July 14, 2015
 First decision: August 25, 2015
 Revised: September 14, 2015
 Accepted: October 23, 2015
 Article in press: October 27, 2015
 Published online: November 10, 2015

Abstract

Because of the intimate association of obesity with type 2 diabetes mellitus (T2DM), during the last two decades, extensive research work is being conducted to find out whether the coexistence of the two is a simple association or there is a positive correlating link between the two. In this article, an attempt has been made to collect and analyse the recent developments in this

field and to arrive at a conclusion on the subject. The possible role of several important factors (obtained from adipocytes/not of adipocyte origin) in linking the two has been discussed in detail. Some of the agents, specifically adiponectin, are beneficial (*i.e.*, reduce the incidence of both), while others are harmful (*i.e.*, increase their incidence). From the analysis, it appears that obesity and T2DM are intimately linked.

Key words: Obesity; Insulin; Insulin resistance; Type 2 diabetes mellitus; Adipocyte

© **The Author(s) 2015.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: The objective of this article is to establish the connection of obesity with that of insulin resistance (IR) and type 2 diabetes mellitus (T2DM) by analyzing the recent developments in this field. The factors linking the three have been found to be some adipocytokines as well as certain other factors not of adipocyte origin. Of these, adiponectin appears to play the most beneficial role (so also leptin, peroxisome proliferator-activated receptors, apelin, *etc.*), while others (tumour necrosis factor- α , interleukin-6, resistin, retinol binding protein-4, dipeptidyl peptidase-4, plasminogen activator inhibitor-1, visfatin, free fatty acid, angiotensin II and toll-like receptors) are harmful. Agonists and antagonists of these factors may be designed to fight against obesity, thereby achieving protection for IR and T2DM.

Chakraborti CK. Role of adiponectin and some other factors linking type 2 diabetes mellitus and obesity. *World J Diabetes* 2015; 6(15): 1296-1308 Available from: URL: <http://www.wjgnet.com/1948-9358/full/v6/i15/1296.htm> DOI: <http://dx.doi.org/10.4239/wjd.v6.i15.1296>

INTRODUCTION

It is practically established that type 1 diabetes

mellitus is an autoimmune disorder where the tissue-specific antibodies target and cause complete or near complete destruction of islet- β -cells, leading to absolute insulin deficiency. In contrast, type 2 diabetes mellitus (T2DM) is usually a hereditary disorder, commonly (80%) associated with obesity, where deficient insulin action may be due to a real deficiency of insulin or a relative one associated with normal or even elevated plasma concentrations of insulin, *i.e.*, insulin resistance (IR). Such simultaneous occurrence of the two (T2DM and obesity) suggests the possibility of a strong link between them, and during the past two decades several positive correlations between them have been established by many workers^[1-4]. Besides obesity which is directly linked to T2DM *via* adipocytokines, some nonadipocytokines have been found to be related with T2DM indirectly by interfering with the growth, development and functions of adipocytes (mentioned later). In this article, an attempt has been made to collect and analyse some such authentic work-results together that will help the reader to comprehend and assess the developments in this field.

The intimate association of T2DM and obesity is a world-wide phenomenon. Though much knowledge about the pathophysiology, course and consequences of T2DM has been gathered, it is not so with obesity, which was almost practically considered as a cosmetic problem. But recently, because of its frequent association with T2DM as well as with hypertension, extensive work is being continued on the adipocyte anatomy, distribution pattern, physiological function, pathological role and its possible link with T2DM and hypertension.

PHYSIOLOGICAL ROLE OF ADIPOCYTES AND ADIPOSE TISSUE

Primary physiological role of adipose tissue is to insulate and cushion the body, to store fat when it is in excess and to supply it when needed^[5]. The exogenous and endogenous pathways of lipid metabolism, during which free fatty acids (FFAs) are released from the lipoprotein (chylomicron, very low density lipoprotein, *etc.*) - triglyceride (TG) content upon hydrolysis by the enzyme lipoprotein lipase (LPL), their (FFAs) subsequent storage in fat depots as TG again, and their remobilisation into the periphery by hydrolysis of these stored TGs by the hormone sensitive lipase (HSL), is well established^[5,6]. Insulin plays a major role for maintenance of adipocyte-fat content as it is a potent activator and inhibitor of LPL and HSL, respectively^[5].

SECRETIONS OF ADIPOCYTES (ADIPOCYTOKINES)

Recently, adipocytes are considered as endocrine structures because of their wide variety of chemical secretions (adipocytokines), which affect many diverse physiological functions and related pathological processes

of the body, like metabolism of carbohydrates and lipids, coagulation of blood, maintenance of blood pressure, feeding behaviour and inflammation, affecting almost all the organs of the body. Increased adipocyte number and adipose-tissue mass have been found to result in increased plasma adipocytokine level except adiponectin, whose plasma concentration is actually low in obesity^[5]. Diseases like obesity, T2DM and metabolic syndrome are associated with altered plasma adipokine levels.

A brief discussion of the adipocytokines known till-date along with their possible roles in genesis or amelioration of IR and T2DM is made below. Besides the adipokines, possible involvement of certain other factors (not of adipocyte origin) has also been taken into account (Figure 1).

Leptin

Several physiological functions of leptin along with its source and metabolism have been extensively discussed. This adipokine, which is a product of "*ob*" gene but mediates its function through the receptor coded by "*db*" gene, is involved in energy homeostasis of the body by interfering with the food-behaviour of the animal centrally (hypothalamus) *via* several hormones^[7].

Many studies on mice and human beings have shown a beneficial and balancing complementary relationship between leptin and insulin where leptin has been found to reduce appetite, obesity and IR along with improvement of metabolic disturbances associated with T2DM. Moreover, mice with *db/db* gene (deficient leptin action) have been found to be obese and diabetic^[7].

Though the receptors for insulin and leptin are different, both of them mediate their action through some common second messengers. Therefore, it is possible that leptin may trigger some of the same downstream events triggered by insulin. Increase in tissue sensitivity of insulin by leptin may be due to later's action on oxidation of FFAs which is increased in skeletal muscles leading to its (FFAs) decreased blood concentrations^[7].

Because of such functional cooperation, it may be assumed that obesity due to inadequate leptin action may predispose or get associated with IR and T2DM.

Tumour necrosis factor- α

The role of tumour necrosis factor- α (TNF- α) as a pro-inflammatory cytokine is well established^[8]. It is produced by macrophages (mainly) as well as by some other cell types including visceral adipocytes^[8-10]. Recently, it has been shown that besides its pro-inflammatory property, increased TNF- α inhibits insulin transduction mechanism, resulting in inadequate glucose metabolism, IR and obesity. Because visceral fat is a source of TNF- α , increase in such fat (obesity) leads to increased production of this cytokine, which aggravates obesity and a vicious cycle is established leading to predisposition, onset and progression of T2DM along with IR. Hence, reduction of obesity, which in

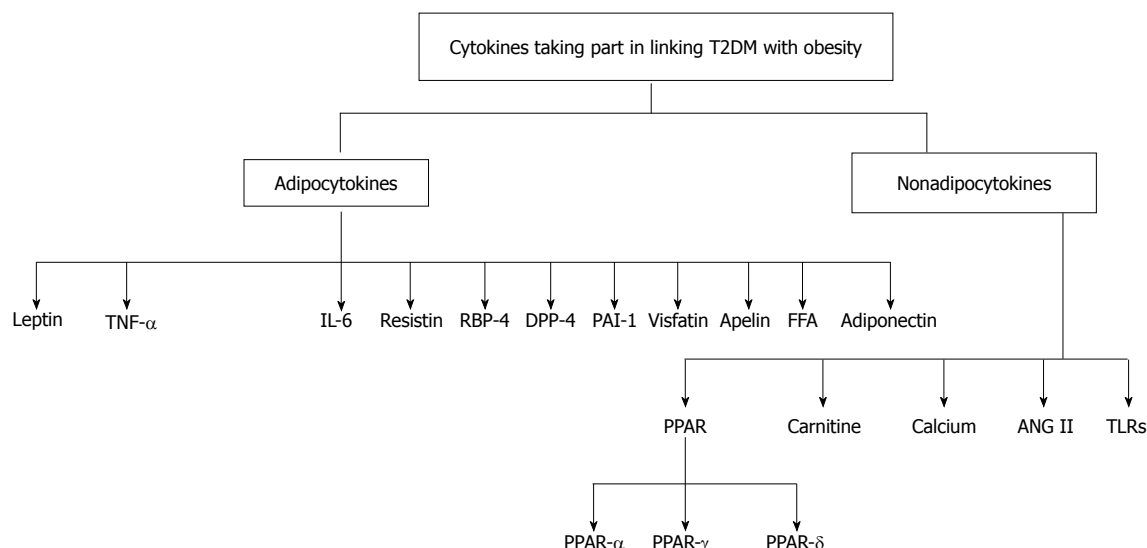


Figure 1 Cytokines linking type 2 diabetes mellitus with obesity. TNF- α : Tumour necrosis factor- α ; IL-6: Interleukin-6; RBP-4: Retinol binding protein-4; DPP-4: Dipeptidyl peptidase-4; FFA: Free fatty acid; PPAR: Peroxisome proliferator-activated receptor; Ang II: Angiotensin II; TLRs: Toll-like receptors; T2DM: Type 2 diabetes mellitus; PAI-1: Plasminogen activator inhibitor-1.

turn may lead to decreased formation of TNF- α , may help to prevent genesis, progression and complications of T2DM^[8]. Besides inhibiting insulin signalling mechanism, TNF- α also has been found to inhibit glucose-induced insulin secretion from β -cells, cause damage to insulin strand and enhance β -cell apoptosis. However, such functions of TNF- α have been demonstrated *in vitro* with concentrations of the cytokine, which was much higher than *in vivo* plasma concentrations^[5]. Moreover, besides visceral adipocytes, macrophages and other cells also produce TNF- α , which may contribute towards the elevated level of this cytokine in obesity^[10]. Therefore, obesity and increased TNF- α levels cannot be directly and definitely implicated with T2DM, although they seem to have a role which needs further investigations^[5,8,9].

Interleukin-6

It is another pro-inflammatory cytokine produced by many cell types (fibroblast, endothelial cells, monocytes) in the body including adipocytes, the production (by adipocytes) being increased in obesity. *In vitro* studies as well as investigations on mice have shown interleukin (IL)-6 to upregulate the production of vascular endothelial growth factor, which is thought to support angiogenesis during adipose tissue growth, leading to increase in the production of IL-6 further (similar to TNF- α)^[5,10].

IL-6 action is mediated through a cytokine class one receptor subtype involving Janus kinase/signal transducers and activators of transcription (JAK/STAT) signal transduction pathway, whereas insulin action is mediated through a receptor family having intrinsic tyrosine kinase activity, signal transduction being carried out through insulin receptor substrate (IRS) proteins. It has been clearly demonstrated that inspite of entirely different receptor involvement, a strong interaction

occurs between the receptor signalling pathway of IL-6 and insulin, leading to impaired biological effect of the later. Though not fully clear, the interaction may involve activation of tyrosine phosphatase, leading to dephosphorylation and inactivation of tyrosine kinase activity or an interaction between suppressor of cytokine signalling proteins and insulin receptors, resulting in deficient insulin action^[10]. Therefore, it appears that elevated plasma levels of IL-6 due to any cause (not necessarily of body fat) may get associated with IR and hence, increased risk of diabetes^[5].

Resistin

This pro-inflammatory cytokine, besides monocytes and macrophages, is also produced by adipocytes. It is so named, because of its capacity to resist insulin action^[1,10,11]. It has a molecular weight of 12.5 kDa and possesses 108 amino acid residues in humans. Unlike adiponectin, this polypeptide has a low circulatory level, which is increased in subjects with IR, T2DM and metabolic syndrome^[3].

Several workers have demonstrated a definite role of resistin in linking obesity to T2DM, during which the cytokine has been found to modulate the insulin signalling pathway, leading to development of IR^[2]. Increased production of resistin has been found to be a result of adipocyte differentiation as well as increase in their number. Locally (from adipocytes) released resistin may play a paracrine role, resulting in inhibition of insulin-induced glucose uptake by adipocytes, which prevents their (adipocytes) further differentiation, thereby reducing its own synthesis and release. This observation may suggest a reciprocal relationship between the two hormones which may further be supported by the fact that rosiglitazone (an oral antidiabetic drug) decreases the circulating concentration of resistin, whereas diet-

induced and genetic forms of obesity increases it^[11]. Moreover, neutralization of resistin has been found to increase the insulin-induced uptake of glucose by adipocytes, whereas resistin itself decreased it.

Recently, it has been observed that resistin-knockout mice show lower fasting blood sugar with increased glucose tolerance and insulin sensitivity associated with reduced hepatic output of glucose. The possible mechanism of this observation may be an overactivity of AMP-activated protein kinase (AMPK) resulting from lack of resistin, leading to reduced expression of genes responsible for hepatic neoglucogenesis. This possible mechanism suggests an opposite role of resistin to that of adiponectin. Again, it was observed that when these resistin-knockout mice were fed with high fat diet, they became obese and IR like their wild counterparts^[10]. All these observations suggest a potential positive link between obesity and T2DM^[11].

Retinol-binding protein-4

This adipocytokine, which is primarily a vitamin A -transport protein, has been recently shown to be linked with IR. Down-regulation of adipocyte GLUT-4 (glucose transporter) has been found to increase the secretion of retinol-binding protein-4 (RBP-4) from adipocytes. In mice, increased serum levels of RBP-4 has been found to be associated with decreased uptake of glucose by skeletal muscles and increased hepatic neoglucogenesis. On the other hand, insulin sensitivity was found to be increased when serum RBP-4 levels were low^[12]. Similar positive correlations between raised plasma RBP-4 level and IR, plasma glucose, BMI and homeostatic model assessment-IR have also been shown in nondiabetics with a high genetic predisposition for T2DM. Interestingly, in this experiment, it was observed that serum RBP-4 levels were raised before significant appearance of diabetic markers^[13]. Such an observation indicates the "elevated plasma RBP-4 level" to be a signal for development of insulin resistance and subsequent T2DM in future^[12,13]. In another experiment, it has been shown that excess of RBP-4 relative to retinol (RBP to retinol ratio) is more accurate in predicting the development of T2DM than raised RBP-4 levels alone^[14].

Dipeptidyl peptidase-4

The incretins (glucagon-like peptide-1 and glucose-dependent insulinotropic hormone) are known to possess favourable effect on carbohydrate and lipid metabolism as they increase postprandial insulin release along with a decrease in release of glucagon. The two incretins, like several other glycoprotein and peptide substrates, are metabolically degraded by the enzyme dipeptidyl peptidase-4 (DPP-4), which reduces their favourable metabolic effects in relation to diabetes and therefore may be considered as diabetogenic. Hence, DPP-4 inhibitors (sitagliptin, vildagliptin, etc.) are now used extensively for management of T2DM along with other antidiabetic agents^[15].

Recently, it has been shown that like other cells, adipocytes also express DPP-4 and substantial over-expression is found in visceral fat of obese persons. Experiments have demonstrated that DPP-4 expression and circulating DPP-4 concentration are well-correlated with adipocyte size and adipose tissue inflammation. This may suggest a stimulatory role of pro-inflammatory adipokines on expression of DPP-4 from adipocytes and other tissues. Thus, increased release of DPP-4 from visceral adipocytes of obese persons may enhance the metabolic degradation of incretins in an autocrine or paracrine manner, thereby reducing their favourable effect on carbohydrate and lipid metabolism which in turn may predispose the concerned obese person for development of T2DM and metabolic syndrome. In another study, it has been shown that explants from subjects release more DPP-4 and the release is reduced after weight reduction^[15]. Moreover, in insulin-sensitive obese patients, plasma concentration of DPP-4 has been found to be lower than those of insulin-resistant obese diabetics^[16]. All these physiological and experimental observations suggest a strong link between T2DM and obesity, where the linking factor appears to be DPP-4.

Plasminogen activator inhibitor-1

This prothrombotic cytokine, besides being produced by vascular endothelial cells, is also produced by adipocytes, production being more from omental adipose tissue than that of subcutaneous adipocytes^[17]. Some recent studies have found a direct contribution of this cytokine towards the complications of obesity like T2DM and coronary thrombosis, as well as increased accumulation of visceral fat^[18]. Nowadays, plasminogen activator inhibitor-1 (PAI-1) is being considered as a strong predictor of T2DM, and has been found to stimulate adipocyte differentiation, which may be mediated through reducing *peroxisome proliferator-activated receptor* (PPAR)- γ activity, resulting in more production of resistin. It has been demonstrated that adipocyte-PAI-1 increases the production of TNF- α (an autocrine action) in adipocytes that reduces insulin action and predisposes to T2DM. Moreover, PPAR- γ receptor has been found to be downregulated both by PAI-1 and TNF- α . Hence, inhibition of PAI-1 action on adipocytes may prevent obesity and IR, and retard adipocyte differentiation and fat accumulation by removing not only its (of PAI-1) own antiinsulin action but also that of resistin and TNF- α ^[7,17].

Visfatin

This adipocytokine, a pro-inflammatory marker of adipose tissue, is mainly produced by visceral adipocytes of humans and mice, whose plasma concentration increases along with the progression of obesity^[19-21]. Its production is upregulated by hypoxia, inflammation and hyperglycemia, and downregulated by insulin, somatostatin and cholesterol reducing statins. Besides visceral fat, intracellular presence of visfatin has also

been demonstrated in many other tissues and organs, the location being both cytoplasmic and nuclear^[21].

Functions of visfatin are difficult to explain as they appear to be contradictory. The cytokine has been found to possess insulinomimetic effect in cultured cells^[19,20] and lowers plasma glucose concentration in mice^[19]. It has also been shown to cause hypoglycaemia by reducing hepatic output of glucose and increasing utilisation of glucose in adipocytes and monocytes^[21]. In spite of such favourable insulinomimetic action^[19,20], this cytokine has been found to be associated with IR and possesses a direct relationship between its plasma concentration and T2DM^[21,22]. This anomaly may be explained by the fact that it also produces hyperlipidemia, which may be responsible for IR and hence T2DM (As T2DM may either be due to deficiency of insulin or IR)^[22]. The resultant effect seems to be favouring the development of T2DM, which in turn suggests the pernicious role of visceral adipose tissue (VAT) in human obesity-related T2DM and accompanying metabolic disorders^[20].

Besides these T2DM-related pathological functions, visfatin, by its endocrine, autocrine as well as paracrine function, has been found to cause increase in cell proliferation and biosynthesis of nicotinamide mono- and dinucleotides^[16], significance of which is yet to be ascertained.

Apelin

Apelin, a small peptide adipokine, has also been found to be present in a number of tissues. It is the ligand of the G-protein-coupled receptor (GPCR) APJ, and has several active forms, which include apelin 13, apelin 17 and apelin 36. It is considered as a beneficial adipokine as it has been found to possess antiobesity and anti-diabetic properties, because of its potent positive role^[23,24] in energy metabolism and insulin sensitivity improvement^[24]. Such actions appear to be due to promotion of complete lipid combustion^[23] in muscle of IR mice through mitochondrial biogenesis and tighter matching between fatty acid oxidation and TCA cycle. Such apelin-stimulated improvement of FA oxidation led to decreased levels of acyl-carnitines and enhanced insulin-stimulated glucose uptake in soleus muscle^[25]. For such beneficial actions, apelin may be considered as a promising useful therapeutic agent for T2DM and other metabolic disorders^[23].

FFA

FFAs, which are produced during the metabolism of exogenous and endogenous lipids, play an important role in the development of IR and hence, genesis of T2DM, when their plasma concentration is abnormally raised^[26].

Mechanisms of FFA-induced IR include inhibition of insulin-induced release of NO from endothelial cells, resulting in decreased blood flow, inhibition of insulin-stimulated glucose transport across the cell membrane and/or inhibition of intracellular phosphorylation of

glucose by interfering with insulin signal transduction pathway. Acute elevation of FFA in plasma has been found to be associated with IR, which may account for 50% of IR in obese individuals with T2DM^[27].

Intracellular mechanism of FFA-induced IR has been demonstrated both *in vivo* and *in vitro*, where there was an activation of pro-inflammatory nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) pathway. It has been shown *in vivo* that acute increase in FFA level resulted in activation of NF- κ B pathway in human skeletal muscle and rat liver, leading to increased production of pro-inflammatory cytokines, i.e., TNF- α , IL-1 β and IL-6, in both the test organs along with an increase in the concentration of macrophage chemoattractant protein-1 (MCP-1) in circulation. In health as well as in T2DM, insulin tends to reduce FFA-induced-IR by lowering the plasma concentration of FFAs through its lipogenic as well as antilipolytic action along with increased intracellular oxidation of FFA. However, in obesity, which is considered as an inflammatory state, there is not only an increase in FFA, but also an increase in the plasma concentration of pro-inflammatory cytokines, which together are liable to cause IR and T2DM^[27].

Thus, obesity alone or along with increased FFA, can create and maintain a low grade inflammatory state by production of pro-inflammatory cytokines (TNF- α , IL-6, etc.), which may induce IR and T2DM. The condition may be further aggravated by antiinsulin action of FFA on glucose metabolism^[27].

Adiponectin

This adipocytokine is being extensively studied worldwide since the past decade because of its remarkable insulin sensitizing property (IR is the major problem in T2DM) as well as antiatherogenic action (dyslipidemia, commonly associated with T2DM, is responsible for atherosclerotic complications of T2DM), thereby playing an important role in delaying and suppressing the metabolic derangements, which result in IR, T2DM, metabolic syndrome and complications of diabetes including vascular and cardiac. These two important functions of adiponectin involves myriads of interrelated molecular mechanisms, which interconnect it with other diabetogenic/antidiabetic adipokines as well as with many physiological and biochemical processes associated with maintenance of energy balance from metabolism of carbohydrates and lipids^[3]. Because of such widespread metabolic involvement, an attempt has been made to discuss the pathophysiological role of this key adipocytokine in detail, which in concert with its siblings appears to play an important role in linking T2DM with obesity.

Source and location: Adiponectin, secreted by both white and brown adipose tissue, has several other names like gelatin binding protein-28, AdipoQ, Adipocyte complement-related protein-30 and OP-MI.

Adiponectin mRNA presence is lower in VAT than that of subcutaneous adipose tissue^[4,10]. Normal plasma concentration of this cytokine varies from 5–30 µg/mL and is inversely proportional to abdominal obesity, IR and T2DM. In some animal models, a decrease in plasma adiponectin concentration was found to precede the onset of T2DM and was parallel with decreased insulin sensitivity. The cytokine circulates in blood in multimeric forms, like trimeric, hexameric and high molecular mass species, each of which plays a specific role in maintenance of energy homeostasis^[4,7].

Control of secretion: Control of adiponectin secretion is effected by: (1) some hormones; (2) many adipokines; (3) certain receptor families including its own; (4) endoplasmic reticulum (ER) and oxidative stresses; and (5) several other factors.

Hormonal control: Sex hormone: Adiponectin plasma concentration has been found to be higher in women than men, which may be due to the difference in concentration of oestrogen and androgen, suggesting a presumably stimulating role of oestrogen on synthesis and secretion of this adipokine^[5,7]; Insulin: The relationship between plasma insulin concentration and adiponectin secretion appears to be peculiar, confusing and contradictory, as the experimental observations do not correlate as expected.

Though insulin favours adiponectin biosynthesis through PPAR-γ *via* inhibition of FoxO1 (an inhibitor of PPAR-γ), type I diabetic patients who practically have no circulating insulin, contrary to the expectations, show elevated levels of plasma adiponectin. Moreover, patients, with defective insulin receptors due to abnormal genes coding for them, also show raised circulating adiponectin levels. Furthermore, adiponectin concentration, unlike other insulin-resistance-inducing adipokines, has been found to be decreased in obesity and insulin-resistant models. From such observations it seems that IR decreases plasma adiponectin concentration. This may be explained by taking into account the role of oxidative stress which is known to increase IR and to decrease adiponectin production. In obesity, adipocytes may develop oxidative stress, leading to decreased expression of adiponectin by them. That IR decreases adiponectin expression may further be supported by the observation that hyperinsulinemia associated with euglycemia (an IR state) significantly decreases the plasma adiponectin concentration and selectively downregulates its high molecular weight (HMW) form. The disparity between the above mentioned experimental observations in relation to role of insulin on adiponectin formation is not known and appears to be complicated^[4].

Control by adipokines: TNF-α and IL-6 are considered to be established inhibitors of adiponectin synthesis^[28]. As their synthesis and secretion increase in obesity, adiponectin plasma concentration decreases

accordingly^[4,29].

Control by certain receptors: PPAR-γ: This PPAR subfamily transcription factor, which is mainly found in adipocytes, has been shown to possess a positive regulatory role on adiponectin gene expression leading to increased production of proteins like Erol-La and DsbA-L, which take part in synthesis and secretion of adiponectin^[4]; Own receptors: Circulating adiponectin concentration has been found to be inversely related to muscle AdipoR_{1/2} (receptor subtypes), but directly related to subcutaneous AdipoR₂^[4,30].

ER and oxidative stresses: ER is known to be an intracellular fine network of microtubules. It is continuous with the nuclear membrane, and is called sarcoplasmic reticulum in muscles. It controls intracellular calcium ion uptake and release besides its other functions, thereby effecting muscular contraction and relaxation. ER stress, which is produced in obesity, has been shown to be negatively related to adiponectin production by adipocytes. The molecular mechanism involved has been studied in 3T3LI-cells, where oxidative stress in ER lead to increased production of H₂O₂, which, *via* protein kinase B (Akt) and JAK/STAT pathway, appreciably suppressed the expression of adiponectin mRNA and consequent reduction in synthesis of proteins required for adiponectin formation. Moreover, in this model H₂O₂ has been found to increase the production of PAI-1 and IL-6, which are known to inhibit adiponectin synthesis^[31].

Other factors: Obesity: Unlike other adipokines, adiponectin secretion has been found to be decreased in obesity. Though the exact cause of such reduction is not known, the suggested causes include increased production of TNF-α and IL-6^[28], generation of a hypoxicmicroenvironment in the adipocytes of increased fat mass, and obesity-induced increased production of insulin like growth factor binding protein-3, which inhibits adiponectin transcription *via* hypoxia inducible factor-1α dependent pathway^[4,32]; Drugs: PPAR-γ agonists (thiazolidinediones-TZDs), which increase insulin sensitivity, have been found to increase the plasma concentration of adiponectin, whereas anti-HIV drugs like protease inhibitors decrease it^[29].

Physiological functions of adiponectin: Adiponectin, along with other adipokines, interferes in several metabolic functions, like lipid synthesis and storage, neoglucogenesis and peripheral utilisation of glucose, which have been demonstrated in skeletal and cardiac muscles, adipocytes and hepatocytes^[31]. But, it differs from other adipokines in several aspects. Unlike others, its circulating concentration has been found to be decreased in obesity (particularly abdominal obesity) and T2DM, and instead of increasing insulin resistance, it decreases it in addition to possessing antiatherosclerotic effect. In animal models and in

patients with obesity and T2DM, the cytokine has been shown to stimulate fatty acid (FA) oxidation, reduce lipid accumulation in muscles, decrease plasma FA concentration and increase insulin sensitivity. Because of such beneficial involvement in metabolic functions (lipids and carbohydrates), IR and atherosclerosis, this adipokine is expected to impart protection against coronary heart diseases, steatohepatitis, non-alcoholic fatty liver diseases and a wide variety of cancers^[33].

Cellular basis of mechanism of action: Functions of adiponectin have been found to be mediated by three receptor subtypes namely, AdipoR₁, AdipoR₂ and T-cadherin. AdipoR₁ and AdipoR₂ are 7 transmembrane proteins but dissimilar to GPCRs. Its receptor distribution pattern varies from cell type to cell type - AdipoR₁ being found abundantly in muscles, while AdipoR₂ is mainly expressed in hepatocytes. Both the receptors are present in almost every tissue, but in a particular tissue, usually one type predominates. Moreover, degree of affinity of these receptors for different forms of adiponectin also varies^[4,7]. AdipoR₁ has high affinity for globular adiponectin (a cleaved part of full-length adiponectin) but low affinity for full-length adiponectin, whereas AdipoR₂ has intermediate affinity for both forms. Hypoadiponectinemia, associated with IR, upregulates both the receptor types. Such upregulation also occurs in physical activity, suggesting an association between adiponectin hormone system and exercise-induced improvement in IR^[7]. Adiponectin, binding to its cell surface receptors, activates several intracellular signalling molecules like p38MAPK, PPAR, the RAS-associated protein Rab5, PI3K, Akt and AMP-activated protein kinase (AMPK), of which AMPK system and PPARs play an important and dominant role leading to modification of lipid and carbohydrate metabolism^[4,7,29].

As has already been mentioned, in this article emphasis would be given on two important protective physiological functions of adiponectin, *i.e.*, protection against IR and atherosclerosis. Obesity, T2DM, dyslipidemia and IR are intimately related, where one leads to the other and once developed, aggravate each other thereby establishing a vicious cycle, leading to development of practically all the dangerous complications of T2DM^[34]. Increased fatmass, as found in obesity, not only increases the production of bad adipokines who enhance this cycle further but also decreases the production of the good one-adiponectin, deficiency of which contributes significantly towards the development, continuation and aggravation of this cycle. Adiponectin has been shown to prevent the development as well as to break this dangerous cycle, thereby posing itself as a potential therapeutic agent in such condition.

Mechanisms of antiatherosclerotic and IR preventing actions of adiponectin: As these two actions are interrelated, it is convenient to discuss them together. It has already been mentioned that

adiponectin increases FA oxidation in mitochondria that leads to a decrease in plasma concentration of FA. Reduced level of FA in circulation prevents the development and progression of atherosclerosis and IR. Multiple biochemical actions at cellular level are modified for this action of adiponectin that needs an extensive discussion and correlation between them to arrive at a conclusion.

Adiponectin-induced FA oxidation is primarily mediated by phosphorylation (activation) of AMPK - a multi-subunit protein kinase, which appears to be a sensor of intracellular energy status through activation of PPAR- α receptor. It has been demonstrated that when muscles were treated with adiponectin or when its receptors were expressed ectopically, there occurred an increase in AMPK phosphorylation and FA oxidation in the muscles that was abolished by dominant-negative AMPK use. Stressful conditions, like heat shock, hypoxia, starvation and exercise, *etc.*, which need expenditure of more energy (denoted by high AMP - to - ATP ratio) have been found to cause AMPK activation. This important signalling molecule (AMPK) is also directly activated by other upstream kinases, where they cause phosphorylation of its threonine residue in the kinase domain. In skeletal muscle, activated AMPK increases FA oxidation by stimulating the phosphorylation (leading to inactivation) of the key enzyme acetyl-CoA carboxylase (ACC). Reduced ACC activity, in turn, decreases intracellular malonyl-CoA concentration along with stimulation of carnitine palmitoyl transferase 1 (CPT1) activity, leading to increased entry of long-chain FAs into mitochondria and hence, more of their peripheral oxidation. The fact, that adiponectin increases insulin sensitivity by decreasing plasma FA concentration, has been demonstrated in obese and T2DM patients, where serum adiponectin concentration is low. In such patients, administration of adiponectin has been found to increase insulin sensitivity by decreasing their plasma FA and TG^[33].

Metabolic stressful conditions like muscle contraction, hypoxia, ischemia and hyperosmolality, *etc.*, not only increase AMPK activation (as mentioned before), but also stimulate the activity of p38MAPK (a signalling molecule activated by inflammatory cytokines). This indicates an association between the two signalling molecules during signal transduction, though the agonists (adiponectin, inflammatory cytokines) inducing the signals are different. In fact, adiponectin has been found to stimulate the activity of not only AMPK but also that of p38MAPK and PPAR- α in target tissue though the subsequent signal transduction pathway following these three activations is not fully known. Other evidences in muscles suggest a sequential activity of these three, leading to increased FA oxidation and increased glucose uptake by muscles. But it has been shown that when primary hepatocytes are treated with adiponectin, their FA oxidation is not increased, which suggests a differential effect of the cytokine on FA oxidation of muscles and liver^[33].

ER stress decreases adiponectin secretion:

Several workers have shown that ER stress in adipocytes decreases adiponectin secretion. It has been demonstrated that properly integrated mitochondrial function in adipocytes is necessary for adequate secretion of adiponectin. Like other cells, growth and development of adipocytes occur through differentiation and hypertrophy, which need increased mitochondrial function, because of greater energy requirement. Newly differentiated adipocytes are small in size, because of less accumulated TG due to increased FA oxidation in them, as the mitochondrial content and activity are more^[29].

It has been shown that these small adipocytes synthesise and secrete more adiponectin because of their high mitochondrial functional level, whereas large hypertrophied fat cells, as found in obesity, produced the cytokine to lesser extent because of impaired mitochondrial function. Though till now, adiponectin synthesis has not been properly correlated with increased mitochondrial function, it may be due to much greater consumption of energy for the synthesis of this cytokine protein in comparison with other proteins. Therefore, it appears that synthesis of adiponectin in adipocytes needs high consumption of energy, which is produced by elevated (adequate) mitochondrial function. In support of this, it has been shown that rosiglitazone and others agents like Ad-NFR-1, which increase mitochondrial biogenesis, also cause an increase in adiponectin synthesis. This observation points the finger towards mitochondrial dysfunction as the cause of low adiponectin level in obesity^[29].

Moreover, several evidences have been put forward where obesity-induced mitochondrial dysfunction has resulted in ER stress, which decreases adiponectin secretion and development of IR. Both ER stress and mitochondrial dysfunction have been demonstrated to activate a series of reactions involving sequential activation of JNK and activating transcription factor 3 (ATF3), which in turn decrease the transcription of adiponectin. When JNK and ATF3 are inhibited, adiponectin transcription is restored. It has also been suggested that ER stress and impaired mitochondrial function are separately responsible for genesis of IR in various tissues of obese persons^[29].

Adiponectin-induced increase in FA oxidation *via* activation of AMPK and phosphorylation of ACC is of short duration, as ACC phosphorylation is short-lived. Hence, this pathway cannot be considered to be fully responsible for the long term effect of adiponectin in causing weight loss and FA oxidation, for which action through PPAR- α is thought to be involved, because PPAR- α action has been found to persist even after initial signalling is over. This is so, because adiponectin has been found to increase transcriptional activity of PPAR- α and subsequent expression of its target genes *via* activation of AMPK. Involvement of AMPK is supported by the fact that when PPAR- α agonists were administered to obese animals, there occurred an

equivalent and sufficient lowering of lipids, as was found with adiponectin. This fact was further supported by *in vivo* administration of 5-Aminoimidazole-4-carboxamide ribonucleotide (AICAR) to lean and obese Zucker rats where the compound was found to decrease plasma FA and TG levels significantly, because AICAR is known to increase the transcriptional activity of PPAR- α *via* activation of AMPK^[33].

Anti-inflammatory action of adiponectin: Mention has been made about the decreased secretion of inflammatory cytokine TNF- α by adiponectin from macrophages that contribute towards its antiatherogenic effect. This anti-inflammatory property is also likely to be involved in its IR reducing action, because TNF- α and IL-6 are known to decrease adiponectin formation and to induce IR^[10,28].

Recently, it has been shown that NF- κ B activation in endothelial and monocytic cells, which is involved in causation of inflammation and metabolic alteration in obesity, is suppressed in these cells by adiponectin. Moreover, both forms of adiponectin-globular as well as full-length, have been found to decrease the production of pro-inflammatory cytokines IL-6 and MCP1 from inflamed adipocytes that may be due to inhibition of NF- κ B activity as well as PPAR- α expression^[35].

Insulin sensitizing actions of adiponectin: Adiponectin aids to insulin sensitivity by several novel mechanisms, which include - increased FA oxidation, decreased ER stress, improvement in insulin signalling pathway, increased (improved) mitochondrial number and function, increased insulin secretion, decreased hepatic output of glucose, increased uptake of glucose by liver and muscle, and increased glucose metabolism.

Adiponectin-induced increase in FA oxidation has been demonstrated by several workers^[5,10,33,36]. This action of adiponectin contributes significantly towards its insulin sensitizing action and prevention of development of IR, as increased plasma FA concentration is the most important cause of IR. In some animal models, adiponectin has been shown to decrease FFA concentration in plasma by increasing its uptake and oxidation in skeletal muscles. On the other hand, acute reduction of plasma FFA has been found to be associated with low adiponectin concentration, though the exact role of FFA in such action is not known^[36]. It is well documented that the key enzyme responsible for FA oxidation is AMPK, which is activated by adiponectin^[4,10,31,33]. It has already been mentioned that once activated, AMPK inhibits the activity of ACC, which not only leads to reduced contents of intracellular malonyl-CoA but also increases activity of CPTI. Such an action increases the entry of long-chain FAs into mitochondria and hence, an increase in their oxidation. Works on FA oxidation in skeletal muscles have shown a sequential activation of AMPK, p38MAPK and PPAR- α to be responsible for increased FA oxidation. But the signalling pathways and components involved in such sequential activation is

not known. PPAR- α , a ligand-activated nuclear receptor, plays an important role in FA oxidation. This receptor is abundantly expressed in tissues like liver, heart, kidney and skeletal muscles, who meet their metabolic energy consumption from oxidation of FAs. It has been shown that HMW adiponectin fraction increases the PPAR- α target gene expression. Moreover, in IR rodent models, PPAR- α ligands have been found to reduce lipid levels and to improve insulin sensitivity. Several studies on humans and rodents have shown that both forms of adiponectin, HMW as well as low molecular weight (LMW), not only increase target gene expression of PPAR- α but also increase the phosphorylation of AMPK and p38MAPK. But such activity is more pronounced and better correlated with HMW fraction than that of LMW, suggesting a differential efficacy between the two fractions or involvement of multiple pathways in increasing FA oxidation in muscles^[33].

It has already been mentioned that mitochondrial dysfunction in adipocytes induces ER stress, which in turn reduces adiponectin transcription, leading to decreased production of this adipokine along with development of IR^[29,31]. Moreover, as discussed earlier, adiponectin, *via* activated AMPK, also improves mitochondrial number and function in skeletal muscles^[29]. From these two observations it may be inferred that adiponectin, by counteracting mitochondrial dysfunction (through improvement of mitochondrial function), decreases ER stress and improves its own secretion, which in turn may contribute towards reduction of IR. Mention has already been made about the IR-inducing and diabetogenic adipocytokine resistin^[1,10,11], whose plasma concentration is high in IR, T2DM, metabolic syndrome and cardiovascular diseases^[3]. In contrast, its sibling adiponectin plasma concentration is low in such conditions^[4], and it has favourable effects on them. Such contrasting effects of the two adipokines may be due to their comparable domain architecture, assembled in a multimeric form, which suggests a common regulatory mechanism (opposite to each other) on insulin-signalling pathway, as well as on mechanisms involved in glucose and lipid homeostasis. In IR and T2DM, hypoadiponectinemia along with hyperresistinemia have been found to antagonise insulin signalling by causing dephosphorylation and deactivation of the key enzyme AMPK in skeletal muscles and liver along with increased expression of genes coding for the synthesis of neoglucogenic enzymes as well as reduced expression of IRS-2 and glucose transporter, GLUT-2. The resultant effects of such action were decreased FFA oxidation in muscles, decreased hepatic uptake of glucose, increased neoglucogenesis and glycogenolysis leading to hyperglycemia and increased plasma FFA. Impaired FFA oxidation may be further aggravated by downregulated PPAR- α action^[3].

It is well established that insulin resistance is very often associated with inadequate functioning of post receptor signalling molecules including IRS. It has been demonstrated that adiponectin upregulates

IRS-2 by activation of STAT-3 in liver. Such activation was also associated with increased production of IL-6 from macrophages - an adiponectin action mediated through activation of NF- κ B, which does not require activation of classical AdipoR₁ and AdipoR₂ receptors. Upregulation of IRS-2 definitely improves insulin sensitivity, but exact mechanisms of such upregulation are not known. Probably, it is effected by an IL-6 dependent pathway, which is initiated by adiponectin, through its combination with yet another unidentified adiponectin receptor. Moreover, though adiponectin activates AMPK and PPAR- α through activation of its classical AdipoR₁ and AdipoR₂ receptors leading to increased FA oxidation and insulin sensitisation, it has not been possible to link AMPK and PPAR- α activation with the proper functioning of post-receptor insulin signalling molecules^[37]. Experiments on skeletal muscles have demonstrated that AMPK activation by adiponectin occurs by two pathways, out of which one is a major one while the other plays a minor role. In the major pathway (the APPL1/LKB1-dependent pathway), AMPK activation needs the binding of adapter protein APPL1, which promotes the translocation of APPL1-dependent LKB1 into the cytosol where it is anchored. The same pathway has been found to be followed by the insulin sensitising drug metformin. Through the minor pathway (the phospholipase C/Ca²⁺/Ca²⁺/calmodulin-dependent protein kinase kinase-dependent pathway), *via* activation of phospholipase C, Ca²⁺ is released from the intracellular calcium ion stores that plays a minor role in activation of AMPK^[38].

Works on skeletal muscles have shown that adiponectin, through AMPK activation, not only increases mitochondrial function, but also increases their number. As activated AMPK in skeletal muscles has been found to stimulate mitochondrial biogenesis under conditions of chronic energy deprivation or endurance training, it appears that adiponectin-induced-increase in mitochondrial number is due to stimulation of mitochondrial biogenesis. This action of the adipokine points towards its insulin sensitising action, because mitochondrial function in skeletal muscles is taken as an indicator of whole-body insulin sensitivity. Thus, it may be presumed that adipocyte-mitochondrial action, which regulates adiponectin synthesis in adipocytes (already discussed), also regulates skeletal muscle-mitochondrial (or metabolic) activity and insulin action in skeletal muscles through adiponectin^[29].

Though adiponectin does not have any effect on normal insulin secretion, the adipokine has been found to increase it in insulin resistant mice fed with high fat diet. But in such mice, augmentation of secretion occurs only in response to high plasma glucose, but actually inhibited when plasma glucose was low. Adiponectin appears to possess a protective effect on islet- β -cells, as it has been found to reduce the pro-apoptotic effect of FFAs and other cytokines on β -cells^[5].

Several workers have demonstrated the capacity of adiponectin to decrease hepatic output of glucose,

thereby contributing towards reduction of plasma glucose concentration and hence, increased insulin sensitivity^[5,10]. One of the important causes of increased hepatic output of glucose in diabetes mellitus is increased neoglucogenesis due to inadequate insulin action. As mentioned earlier, adiponectin inhibits hepatic neoglucogenesis by decreasing the formation of two important enzymes concerned, through interference with the mRNA expression that is necessary for the synthesis of these enzymes^[10]. Moreover, adiponectin, by increasing the oxidation of FAs, decreases their availability for utilization in the process of neoglucogenesis.

Adiponectin has been found to increase the uptake of glucose by liver and muscles which appears to result from improvement in insulin signalling pathway, leading to better insulin action and hence, decreased blood sugar and increased insulin sensitivity^[5].

It has been observed that in obese individuals with IR and in patients having metabolic syndrome (who are IR), adiponectin receptors are downregulated, which suggests inadequate adiponectin action as the cause of IR. *In vitro* and *in vivo* experiments on skeletal muscles have shown adiponectin to increase glucose metabolism and insulin sensitivity *via* activation of AMPK^[31].

AR (adiponectin: Resistin) and IR_{AR} indices: Upregulated resistin, which is followed by PPAR- α downregulation, has been found to impair adipocyte differentiation, leading to dramatic decrease in adiponectin formation. Because of such inverse relationship with respect to both secretion and function, it seems to be more predictive to use their ratio (AR index-adiponectin: Resistin) in linking obesity with T2DM than using either of them alone^[3].

Besides AR index another novel IR_{AR} index has been coined that seems to be a strong indicator of degree of IR in T2DM. The index appears to relate IR with AR. As expected, AR index value gets smaller and smaller according to the degree of obesity (which determines the magnitude of hypoadiponectinemia with hyperresistinemia), resulting in a parallel rise of IR. Hence, greater the IR_{AR} index value, more is the degree of IR in T2DM.

As IR in T2DM is the major determinant of progression into metabolic syndrome, which in turn, lays the foundation for other complications of diabetes, this index may also be used to predict the arrival of T2DM complications^[3].

FACTORS NOT OF ADIPOCYTE ORIGIN

In addition to these adipokines, there are some other factors (not of adipocyte origin), whose role in linking obesity with T2DM cannot be ignored. These factors include PPARs, carnitine, calcium, angiotensin II and toll-like receptors (TLRs).

PPARs

This nuclear receptor family, consisting of PPAR- α , PPAR- γ and PPAR- δ , are primarily related with lipid metabolism

having fatty acids and their derivatives as their endogenous ligands.

PPAR- α : Besides interference with several steps of lipid metabolism, the main results of this receptor activation is increased oxidation of FA that leads to decreased plasma level of TG by decreasing its synthesis and storage in adipocytes. Moreover, PPAR- α activation, along with activation of PPAR- γ , has been found not only to increase the formation and secretion of adiponectin but also to upregulate AdipoR₁/AdipoR₂^[7].

PPAR- γ : These receptors, mainly expressed in liver and adipose tissue, on stimulation, cause gene expression necessary for differentiation of fibroblasts into adipocytes, and for lipid synthesis and storage in adipocytes. Because of their lipogenicity, they seem to decrease insulin sensitivity rather than increase it. But, their exogenous agonists-TZDs, have been found to decrease IR and increase insulin sensitivity. Such paradoxical actions of TZDs, have been shown to be due to reduced lipotoxicity in liver and skeletal muscles because of lipid storage in adipocytes, and increase in number of small adipocytes, which are not only more sensitive to insulin action, but also secrete large quantity of adiponectin (insulin-sensitising), while decreasing the release of resistin and TNF- α (both are IR-inducing)^[7].

PPAR- δ : Main result of this receptor activation is increased FA oxidation, which contributes towards decreasing IR and increasing insulin sensitivity^[7].

It may be noted that the results of activation of these three receptors, particularly activation of those of PPAR- α and PPAR- γ , are beneficial in IR and insulin sensitivity through their interference with adipocyte number (increased number of small adipocytes) and function (increased production of adiponectin and decreased production of resistin and TNF- α), FA oxidation (which decreases TG formation in adipocytes resulting in decreased obesity) and upregulation of AdipoR₁ and AdipoR₂ (decreased IR and increased insulin sensitivity). As all these functions finally lead to reduced obesity, this receptor family can be considered to play a role in linking obesity and T2DM.

Carnitine

This vitamin and amino acid, which is derived from yeast, milk, liver and muscles (in large quantities), increases FFA oxidation through carnitine shuttle reactions. In this reaction, carnitine has been found not only to favour entry of long-chain FFAs across the mitochondrial membrane, but also facilitate the transport of fatty acyl-CoA into mitochondrial matrix for β -oxidation. Therefore, carnitine deficiency, which is commonly found in several IR cases, leads to increased concentration of plasma FFA and hence, their increased conversion into TG in adipocytes, resulting in obesity and further aggravation of IR. Moreover, relative carnitine deficiency may occur in prolonged metabolic stress, which may add to mito-

chondrial dysfunction, leading to reduced glucose tolerance. These two factors may contribute towards obesity-associated IR in T2DM. Therefore, like PPAR-receptor family action, carnitine function in the body may contribute towards linking obesity with diabetes as its deficiency is reflected upon genes of obesity and IR^[7].

Calcium

Role of calcium in various cellular secretory processes^[39], including secretion of insulin from islet β -cells, is well established. Improper regulation of intracellular calcium has been found to affect insulin secretion and its tissue sensitivity adversely^[40]. High calcium intake alone or with vitamin D has been shown to reduce not only body weight and fat mass, but also to decrease weight gain and adipocyte fat accumulation. The mechanisms suggested for such beneficial actions include adipocyte apoptosis and reduced adipogenesis along with deranged lipid metabolism^[40,41]. Moreover, epidemiological studies have shown that low calcium intake and poor vitamin D status are associated with increased risk of obesity^[36]. From such observations, it may be inferred that obesity, thus developed, may lead to increased production of IR-inducing and diabetogenic adipokines, thereby linking it (obesity) with IR and T2DM.

Angiotensin II

Renin-angiotensin-aldosterone system, whose primary function is to maintain water and electrolyte balance of the body and to regulate blood pressure, is known to mediate its function by formation of angiotensin II (Ang II). Ang II formation occurs through several steps where renin of renal origin converts angiotensinogen of hepatic origin to Ang I, which is then converted to Ang II by the enzyme angiotensin-converting enzyme (ACE) of endothelial cell origin^[42]. But recently, a local RAAS has been demonstrated in several tissues of the body including adipose tissue, which is involved in several functions of the adipocytes including adipose tissue growth and cell differentiation. It has been shown that when AT₂ receptors (one of the subtypes of angiotensin receptor) are deleted from adipocytes, the cell size is reduced, and there is protection from diet-induced obesity and IR^[43]. Such observations suggest an additional beneficial role of ACE inhibitors and AT₂ receptor blockers, when used as antihypertensives in patients having hypertension with obesity and T2DM^[44]. Moreover, like low Ca²⁺ and poor vitamin D status, locally generated Ang II, via its action on adipocytes, may link obesity with T2DM.

TLRs

TLRs are transmembrane glycoprotein receptors whose known function is antigen recognition^[6,45]. Recently, substantial evidences have been put forward which suggest their pathological role in genesis of obesity. In this respect, both TLR-2 and TLR-4 have been found

to be overexpressed on adipocytes in obese persons having T2DM. Such overexpressed TLR receptors along with similarly overexpressed adipokines in adipose tissue of obese individuals may play an important role in obesity-associated meta inflammation resulting in IR and T2DM. It has been demonstrated that inhibition of TLR-2 in skeletal muscles and white adipose tissue of mice fed with high fat diet, improves insulin sensitivity and signalling^[43].

Moreover, overexpression of TLRs on adipocytes may also suggest an important role of adipose tissue in the regulation of inflammation and innate immunity in human beings by modulating TLR/NF- κ B regulatory pathway. Such observations suggest a modulatory role of TLRs in the interaction between the pathways of inflammation and metabolism^[43]. The above-discussed roles of TLRs in genesis of obesity, reduction of insulin signalling and sensitivity, and modulation of the interacting pathways of inflammation and metabolism appear to support the correlation between obesity and T2DM.

CONCLUSION

From the discussions made so far, it may be observed that results obtained from extensive research work on the factors supposed to link obesity with T2DM, very clearly show an intimate relationship between the two, for which both adipocytokines as well as some factors not derived from adipocytes have been implicated. Of them, few (Adiponectin, Leptin, PPAR, Carnitine, Apelin and Calcium) are beneficial, while others (TNF- α , IL-6, Resistin, RBP-4, DPP-4, PAI-1, Visfatin, FFA, Ang II and TLR) are harmful, but all of them play a definite role in linking obesity with T2DM (mentioned earlier). Among these, adiponectin has been found to play a crucial and seemingly complicated but definite role. Such studies may be extended to all concerned factors giving emphasis on mitochondrial and ER stresses. Finally, using these agents, drugs may be designed which will be helpful to prevent the development of obesity, thereby producing a beneficial response in prevention, progression and treatment of T2DM.

REFERENCES

- 1 **Steppan CM**, Bailey ST, Bhat S, Brown EJ, Banerjee RR, Wright CM, Patel HR, Ahima RS, Lazar MA. The hormone resistin links obesity to diabetes. *Nature* 2001; **409**: 307-312 [PMID: 11201732 DOI: 10.1038/35053000]
- 2 **Sentinelli F**, Romeo S, Arca M, Filippi E, Leonetti F, Banchieri M, Di Mario U, Baroni MG. Human resistin gene, obesity, and type 2 diabetes: mutation analysis and population study. *Diabetes* 2002; **51**: 860-862 [PMID: 11872692 DOI: 10.2337/diabetes.51.3.860]
- 3 **Lau CH**, Muniandy S. Novel adiponectin-resistin (AR) and insulin resistance (IRAR) indexes are useful integrated diagnostic biomarkers for insulin resistance, type 2 diabetes and metabolic syndrome: a case control study. *Cardiovasc Diabetol* 2011; **10**: 8 [PMID: 21251282 DOI: 10.1186/1475-2840-10-8]
- 4 **Shehzad A**, Iqbal W, Shehzad O, Lee YS. Adiponectin: regulation of its production and its role in human diseases. *Hormones (Athens)* 2012; **11**: 8-20 [PMID: 22450341]
- 5 **Hajer GR**, van Haeften TW, Visseren FL. Adipose tissue

- dysfunction in obesity, diabetes, and vascular diseases. *Eur Heart J* 2008; **29**: 2959-2971 [PMID: 18775919 DOI: 10.1093/eurheartj/ehn387]
- 6 **Rang HP**, Dale MM, Ritter JM, Flower RJ, Handerson G. Rang and Dale's Pharmacology. 7th ed. Edinburgh: Churchill Livingstone, 2012: 77-88
 - 7 **Chakraborti CK**. Possible links between obesity and type 2 diabetes mellitus. *Int J Pharm Sci Res* 2012; **3**: 1935-1945 Available from: URL: <http://www.ijpsr.com>
 - 8 **Swaroop JJ**, Rajarajeswari D, Naidu JN. Association of TNF- α with insulin resistance in type 2 diabetes mellitus. *Indian J Med Res* 2012; **135**: 127-130 [PMID: 22382194 DOI: 10.4103/0971-5916.93435]
 - 9 **Borst SE**. The role of TNF-alpha in insulin resistance. *Endocrine* 2004; **23**: 177-182 [PMID: 15146098 DOI: 10.1385/ENDO.23:2-3:177]
 - 10 **Bastard JP**, Maachi M, Lagathu C, Kim MJ, Caron M, Vidal H, Capeau J, Feve B. Recent advances in the relationship between obesity, inflammation, and insulin resistance. *Eur Cytokine Netw* 2006; **17**: 4-12 [PMID: 16613757]
 - 11 **Kusminski CM**, McTernan PG, Kumar S. Role of resistin in obesity, insulin resistance and Type II diabetes. *Clin Sci (Lond)* 2005; **109**: 243-256 [PMID: 16104844 DOI: 10.1042/CS20050078]
 - 12 **Wolf G**. Serum retinol-binding protein: a link between obesity, insulin resistance, and type 2 diabetes. *Nutr Rev* 2007; **65**: 251-256 [PMID: 17566551 DOI: 10.1111/j.1753-4887.2007.tb00302.x]
 - 13 **Bose KS**, Gupta SK, Singh S. Is serum retinol binding protein-4: A predictor for diabetes in genetically high risk population? *J Res Med Sci* 2012; **17**: 1015-1019 [PMID: 23833574]
 - 14 **Erikstrup C**, Mortensen OH, Nielsen AR, Fischer CP, Plomgaard P, Petersen AM, Krogh-Madsen R, Lindegaard B, Erhardt JG, Ullum H, Benn CS, Pedersen BK. RBP-to-retinol ratio, but not total RBP, is elevated in patients with type 2 diabetes. *Diabetes Obes Metab* 2009; **11**: 204-212 [PMID: 19215278 DOI: 10.1111/j.1463-1326.2008.00901.x]
 - 15 **Lamers D**, Famulla S, Wronkowitz N, Hartwig S, Lehr S, Ouwers DM, Eckardt K, Kaufman JM, Ryden M, Müller S, Hanisch FG, Ruige J, Arner P, Sell H, Eckel J. Dipeptidyl peptidase 4 is a novel adipokine potentially linking obesity to the metabolic syndrome. *Diabetes* 2011; **60**: 1917-1925 [PMID: 21593202 DOI: 10.2337/db10-1707]
 - 16 **Blüher M**. Adipokines - removing road blocks to obesity and diabetes therapy. *Mol Metab* 2014; **3**: 230-240 [PMID: 24749053 DOI: 10.1016/j.molmet.2014.01.005]
 - 17 **Correia ML**, Haynes WG. A role for plasminogen activator inhibitor-1 in obesity: from pie to PAI? *Arterioscler Thromb Vasc Biol* 2006; **26**: 2183-2185 [PMID: 16990563 DOI: 10.1161/01.ATV.0000244018.24120.70]
 - 18 **De Taeye B**, Smith LH, Vaughan DE. Plasminogen activator inhibitor-1: a common denominator in obesity, diabetes and cardiovascular disease. *Curr Opin Pharmacol* 2005; **5**: 149-154 [PMID: 15780823 DOI: 10.1016/j.coph.2005.01.007]
 - 19 **Fukuhara A**, Matsuda M, Nishizawa M, Segawa K, Tanaka M, Kishimoto K, Matsuki Y, Murakami M, Ichisaka T, Murakami H, Watanabe E, Takagi T, Akiyoshi M, Ohtsubo T, Kihara S, Yamashita S, Makishima M, Funahashi T, Yamanaka S, Hiramatsu R, Matsuzawa Y, Shimomura I. Visfatin: a protein secreted by visceral fat that mimics the effects of insulin. *Science* 2005; **307**: 426-430 [PMID: 15604363 DOI: 10.1126/science.1097243]
 - 20 **Sethi JK**, Vidal-Puig A. Visfatin: the missing link between intra-abdominal obesity and diabetes? *Trends Mol Med* 2005; **11**: 344-347 [PMID: 16005682 DOI: 10.1016/j.molmed.2005.06.010]
 - 21 **Adeghate E**. Visfatin/PBEF- a new natural insulin-mimetic adipokine. *Curr Med Chem* 2008; **15**: 1851-1862
 - 22 **Chang YC**, Chang TJ, Lee WJ, Chuang LM. The relationship of visfatin/pre-B-cell colony-enhancing factor/nicotinamide phosphoribosyltransferase in adipose tissue with inflammation, insulin resistance, and plasma lipids. *Metabolism* 2010; **59**: 93-99 [PMID: 19765775 DOI: 10.1016/j.metabol.2009.07.011]
 - 23 **Castan-Laurell I**, Dray C, Attané C, Duparc T, Knauf C, Valet P. Apelin, diabetes, and obesity. *Endocrine* 2011; **40**: 1-9 [PMID: 21725702 DOI: 10.1007/s12020-011-9507-9]
 - 24 **Castan-Laurell I**, Dray C, Knauf C, Kunduzova O, Valet P. Apelin, a promising target for type 2 diabetes treatment? *Trends Endocrinol Metab* 2012; **23**: 234-241 [PMID: 22445464 DOI: 10.1016/j.tem.2012.02.005]
 - 25 **Attané C**, Foussal C, Le Gonidec S, Benani A, Daviaud D, Wanecq E, Guzmán-Ruiz R, Dray C, Bezaire V, Rancoule C, Kuba K, Ruiz-Gayo M, Levade T, Penninger J, Burcelin R, Pénicaud L, Valet P, Castan-Laurell I. Apelin treatment increases complete Fatty Acid oxidation, mitochondrial oxidative capacity, and biogenesis in muscle of insulin-resistant mice. *Diabetes* 2012; **61**: 310-320 [PMID: 22210322 DOI: 10.2337/db11-0100]
 - 26 **Huang Y**, Li X, Wang M, Ning H, A L, Li Y, Sun C. Lipoprotein lipase links vitamin D, insulin resistance, and type 2 diabetes: a cross-sectional epidemiological study. *Cardiovasc Diabetol* 2013; **12**: 17 [PMID: 23320821 DOI: 10.1186/1475-12-17]
 - 27 **Boden G**. Obesity, insulin resistance and free fatty acids. *Curr Opin Endocrinol Diabetes Obes* 2011; **18**: 139-143 [PMID: 21297467 DOI: 10.1097/MED.0b013e3283444b09]
 - 28 **Lubos E**, Handy DE, Loscalzo J. Role of oxidative stress and nitric oxide in atherothrombosis. *Front Biosci* 2008; **13**: 5323-5344 [PMID: 18508590]
 - 29 **Koh EH**, Park JY, Park HS, Jeon MJ, Ryu JW, Kim M, Kim SY, Kim MS, Kim SW, Park IS, Youn JH, Lee KU. Essential role of mitochondrial function in adiponectin synthesis in adipocytes. *Diabetes* 2007; **56**: 2973-2981 [PMID: 17827403 DOI: 10.2337/db07-0510]
 - 30 **Blüher M**, Williams CJ, Klötting N, Hsi A, Ruschke K, Oberbach A, Fasshauer M, Berndt J, Schön MR, Wolk A, Stumvoll M, Mantzoros CS. Gene expression of adiponectin receptors in human visceral and subcutaneous adipose tissue is related to insulin resistance and metabolic parameters and is altered in response to physical training. *Diabetes Care* 2007; **30**: 3110-3115 [PMID: 17878241 DOI: 10.2337/dc07-1257]
 - 31 **Boddu NJ**, Theus S, Luo S, Wei JY, Ranganathan G. Is the lack of adiponectin associated with increased ER/SR stress and inflammation in the heart? *Adipocyte* 2014; **3**: 10-18 [PMID: 24575364 DOI: 10.4161/adip.26684]
 - 32 **Marcotorchino J**, Gouranton E, Romier B, Tourniaire F, Astier J, Malezet C, Amiot MJ, Landrier JF. Vitamin D reduces the inflammatory response and restores glucose uptake in adipocytes. *Mol Nutr Food Res* 2012; **56**: 1771-1782 [PMID: 23065818 DOI: 10.1002/mnfr.201200383]
 - 33 **Yoon MJ**, Lee GY, Chung JJ, Ahn YH, Hong SH, Kim JB. Adiponectin increases fatty acid oxidation in skeletal muscle cells by sequential activation of AMP-activated protein kinase, p38 mitogen-activated protein kinase, and peroxisome proliferator-activated receptor alpha. *Diabetes* 2006; **55**: 2562-2570 [PMID: 16936205 DOI: 10.2337/db05-1322]
 - 34 **Shu AD**, Myer MG. Pharmacology of the endocrine pancreas. In: Golan DE, Tashjian AH, Armstrong EJ, Galanter JM, Armstrong AW, Arnaout RA, Rose HS. Principles of Pharmacology: The pathophysiologic basis of drug therapy. Philadelphia: Lippincott Williams and Wilkins, 2005: 457-469
 - 35 **Zoico E**, Garbin U, Olivos D, Mazzali G, Fratta Pasini AM, Di Francesco V, Sepe A, Cominacini L, Zamboni M. The effects of adiponectin on interleukin-6 and MCP-1 secretion in lipopolysaccharide-treated 3T3-L1 adipocytes: role of the NF-kappaB pathway. *Int J Mol Med* 2009; **24**: 847-851 [PMID: 19885628 DOI: 10.3892/ijmm.00000302]
 - 36 **Bernstein EL**, Koutkia P, Ljungquist K, Breu J, Canavan B, Grinspoon S. Acute regulation of adiponectin by free fatty acids. *Metabolism* 2004; **53**: 790-793 [PMID: 15164330]
 - 37 **Awazawa M**, Ueki K, Inabe K, Yamauchi T, Kubota N, Kaneko K, Kobayashi M, Iwane A, Sasako T, Okazaki Y, Ohsugi M, Takamoto I, Yamashita S, Asahara H, Akira S, Kasuga M, Kadowaki T. Adiponectin enhances insulin sensitivity by increasing hepatic IRS-2 expression via a macrophage-derived IL-6-dependent pathway. *Cell Metab* 2011; **13**: 401-412 [PMID: 21459325 DOI: 10.1016/j.cmet.2011.02.010]
 - 38 **Zhou L**, Deepa SS, Etzler JC, Ryu J, Mao X, Fang Q, Liu DD, Torres

- JM, Jia W, Lechleiter JD, Liu F, Dong LQ. Adiponectin activates AMP-activated protein kinase in muscle cells via APPL1/LKB1-dependent and phospholipase C/Ca2+/Ca2+/calmodulin-dependent protein kinase kinase-dependent pathways. *J Biol Chem* 2009; **284**: 22426-22435 [PMID: 19520843 DOI: 10.1074/jbc.M109.028357]
- 39 **Penner R**, Neher E. The role of calcium in stimulus-secretion coupling in excitable and non-excitable cells. *J Exp Biol* 1988; **139**: 329-345 [PMID: 2850338]
- 40 **Pittas AG**, Dawson-Hughes B, Li T, Van Dam RM, Willett WC, Manson JE, Hu FB. Vitamin D and calcium intake in relation to type 2 diabetes in women. *Diabetes Care* 2006; **29**: 650-656 [PMID: 16505521 DOI: 10.2337/diacare.29.03.06.dc05-1961]
- 41 **Song Q**, Sergeev IN. Calcium and vitamin D in obesity. *Nutr Res Rev* 2012; **25**: 130-141 [PMID: 22588363 DOI: 10.1017/S0954422412000029]
- 42 **Jackson EK**. Renin and angiotensin. In: Brunton LL, Lazo JS, Parker KL. Goodman and Gilman's The Pharmacological Basis of Therapeutics. New York: Mc Graw-Hill Medical Publishing Division, 2006: 789-821
- 43 **vinh quốc Lu'ông K**, Nguyễn LT. The beneficial role of vitamin D in obesity: possible genetic and cell signaling mechanisms. *Nutr J* 2013; **12**: 89 [PMID: 23800102 DOI: 10.1186/1475-2891-12-89]
- 44 **Johnston CI**. Angiotensin receptor antagonists for the treatment of hypertension. *Aust Prescr* 1998; **21**: 95-97
- 45 **Stafford JL**, Ellestad KK, Magor KE, Belosevic M, Magor BG. A toll-like receptor (TLR) gene that is up-regulated in activated goldfish macrophages. *Dev Comp Immunol* 2003; **27**: 685-698 [PMID: 12798365 DOI: 10.1016/S0145-305X(03)00041-7]

P- Reviewer: Chang ST, Tarantino G **S- Editor:** Ji FF **L- Editor:** A
E- Editor: Lu YJ





Published by **Baishideng Publishing Group Inc**

8226 Regency Drive, Pleasanton, CA 94588, USA

Telephone: +1-925-223-8242

Fax: +1-925-223-8243

E-mail: bpgoffice@wjgnet.com

Help Desk: <http://www.wjgnet.com/esps/helpdesk.aspx>

<http://www.wjgnet.com>

